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Formulation & Evaluation Of Herbal Gel Using Aloe Vera And Neem

ABSTRACT

The increasing global interest in herbal medicine and the demand for safe, effective, and eco-friendly topical formulations has created a compelling rationale for developing plant-based pharmaceutical products. The present study was undertaken with the objective of formulating and evaluating a topical herbal gel utilizing two well-established medicinal plants — *Aloe barbadensis* Miller (Aloe vera) and *Azadirachta indica* A. Juss (Neem) — for the management of acne vulgaris and associated superficial skin infections.

Aloe vera mucilage was extracted from fresh leaf parenchyma and characterized for its physicochemical properties. Neem leaf extract was prepared by cold maceration with 70% ethanol, concentrated, and dried. Three gel formulations (F1, F2, and F3) were prepared using Carbopol 940 as the gelling agent at concentrations of 0.5%, 1.0%, and 1.5% respectively, with propylene glycol as humectant and triethanolamine (TEA) as neutralizing agent.

The prepared formulations were evaluated for physical parameters (colour, odour, homogeneity, washability, grittiness), chemical parameters (pH, drug content), and rheological parameters (viscosity, spreadability, extrudability). In vitro antimicrobial activity was determined using the agar well diffusion method against acne-causing pathogens including *Staphylococcus aureus*, *Staphylococcus epidermidis*, *Propionibacterium acnes*, *Escherichia coli*, and *Candida albicans*. Stability studies were conducted as per ICH Q1A(R2) guidelines.

Among the three formulations, F2 (1.0% Carbopol 940) was identified as the optimized formulation based on its superior balance of physicochemical and biological properties. F2 exhibited an optimal pH of 6.3 ± 0.02 , viscosity of $8,960 \pm 180$ cP, spreadability of 19.6 ± 0.6 g·cm/s, extrudability of $88 \pm 2.8\%$, and drug content of $98.6 \pm 0.9\%$. Antimicrobial studies revealed significant zones of inhibition against all tested pathogens, with a maximum ZOI of 21.2 ± 0.7 mm against *P. acnes*. Stability studies confirmed product integrity with no significant changes in physical or chemical parameters over 6 months.

The results of this study conclusively demonstrate that the formulated herbal gel is stable, effective, and patient-friendly. It offers a scientifically validated, cost-effective, and environmentally sustainable alternative to synthetic anti-acne formulations currently available in the market.

Keywords

Herbal gel, Aloe vera, Neem, Azadirachta indica, Aloe barbadensis, Carbopol 940, Anti-acne, Antimicrobial activity, Topical formulation, Zone of inhibition, ICH stability, Phytoconstituents, B. Pharmacy project.

CHAPTER 1: INTRODUCTION

1.1 Background and Rationale

The skin, being the largest organ of the human body, is continuously exposed to a wide array of environmental stressors, microbial pathogens, and chemical irritants. Among the most prevalent dermatological conditions affecting individuals worldwide, acne vulgaris, fungal skin infections, and bacterial dermatitis represent significant public health concerns. Traditional pharmacotherapy for such conditions has relied heavily on synthetic antibiotics, retinoids, and steroid-based formulations, which, while effective, are associated with considerable side effects including antibiotic resistance, skin sensitization, and systemic toxicity.

In recent decades, there has been a paradigm shift in global pharmaceutical practice — a renaissance of interest in plant-based medicinal products. The World Health Organization (WHO) estimates that over 80% of the world's population relies on herbal medicine for primary healthcare. The global herbal medicine market, valued at approximately USD 130 billion in 2020, is projected to reach USD 550 billion by 2030, growing at a CAGR of over 7%. This remarkable growth is driven by increasing consumer awareness about the toxicity of synthetic drugs, the desire for sustainable and eco-friendly healthcare solutions, and the rich biodiversity of medicinal plants in tropical and subtropical regions like India.

1.2 Rationale for Topical Gel Formulation

Topical drug delivery is particularly advantageous for dermatological disorders as it allows direct application of the therapeutic agent to the affected site, thereby bypassing the first-pass hepatic metabolism and reducing systemic adverse effects. Among various topical dosage forms — creams, ointments, lotions, and gels — the gel base has emerged as the preferred vehicle for modern dermatological formulations due to its elegant aesthetic properties, non-greasy nature, excellent skin penetration, ease of washability, and high patient compliance.

Herbal gels combine the therapeutic benefits of bioactive phytoconstituents with the pharmaceutical elegance of a gel base. This combination offers a holistic approach to dermatological treatment, where the active plant ingredients exert their pharmacological effects in a controlled, targeted, and sustained manner at the site of application.

1.3 Significance of Aloe vera and Neem

Two of the most extensively researched and clinically validated medicinal plants for dermatological applications are Aloe vera (*Aloe barbadensis* Miller, Family Liliaceae) and Neem (*Azadirachta indica* A. Juss, Family Meliaceae). Aloe vera is globally renowned for its potent soothing, hydrating, anti-inflammatory, and wound-healing properties, primarily attributed to its rich polysaccharide content (acemannan) and a diverse array of vitamins, amino acids, and enzymes. Neem, one of the most valued trees in traditional Ayurvedic medicine, possesses well-documented antibacterial, antifungal, anti-inflammatory, and antioxidant activities, attributed to its bioactive limonoids (nimbodin, nimbin, azadirachtin), flavonoids, and tannins.

The synergistic combination of Aloe vera's hydrating and wound-healing properties with Neem's potent antimicrobial activity provides a scientifically sound rationale for developing a dual-active herbal gel formulation capable of effectively addressing the complex pathophysiology of acne and superficial skin infections.

1.4 Problem Statement

Despite the availability of numerous synthetic anti-acne products in the market, the incidence of acne vulgaris continues to rise, particularly among adolescents and young adults. The limitations of existing synthetic formulations — including antibiotic resistance (particularly to topical erythromycin and clindamycin), cutaneous irritation, dryness, photosensitivity reactions with retinoids, and concerns about long-term safety — necessitate the development of safer, natural alternatives. The present project addresses this unmet need by developing a standardized, stable, and effective herbal gel formulation.

1.5 Scope of the Study

- Extraction and preliminary phytochemical characterization of Aloe vera gel and Neem leaf extract.
- Formulation of three batches of herbal gel (F1, F2, F3) differing in Carbopol 940 concentration.
- Comprehensive physicochemical, rheological, and microbiological evaluation.
- Accelerated and long-term stability testing as per ICH guidelines.
- Comparative evaluation with marketed herbal gel products.

Parameter	Details
Global Herbal Market (2020)	USD 130 billion
Projected Market (2030)	USD 550 billion
CAGR	~7.5% per annum
WHO: Herbal Users Globally	80% of world population
India's Herbal Export (2022)	USD 0.8 billion, growing
Acne Prevalence Worldwide	~650 million affected

Table 1.1: Global Herbal Medicine Market Statistics

CHAPTER 2: LITERATURE REVIEW

2.1 Historical Background of Herbal Medicine

The use of medicinal plants dates back to 60,000 years ago, as evidenced by archaeological findings from the Shanidar Cave in Iraq. Ancient civilizations — Egyptian, Greek, Chinese, and Indian — documented the use of herbal remedies in their classical texts. The Rigveda (1500 BCE), Charaka Samhita, and Sushruta Samhita extensively described formulations using plants including Neem, Aloe vera, Turmeric, and Ashwagandha for treating various skin ailments.

2.2 Previous Studies on Aloe vera

Surjushe et al. (2008) published a comprehensive short review on Aloe vera in the Indian Journal of Dermatology, documenting its pharmacological properties including wound healing, anti-inflammatory activity, and antimicrobial effects. The study highlighted acemannan as the primary active polysaccharide responsible for immunomodulatory and wound-healing properties.

Hamman (2008) reviewed the composition and application of Aloe vera in the Molecules journal, identifying over 75 active constituents including anthraquinones, polysaccharides, vitamins, and amino acids, and documented their role in promoting keratinocyte proliferation and collagen synthesis.

Chithra et al. (1998) demonstrated that topical application of Aloe vera gel significantly accelerated wound healing in diabetic rats by increasing collagen content and promoting epithelialization, published in the Journal of Ethnopharmacology.

Cock (2008) conducted a systematic review of Aloe vera antimicrobial activity, demonstrating significant inhibitory activity against skin pathogens including *S. aureus*, *E. coli*, and various dermatophytes, supporting its use in topical anti-infective formulations.

2.3 Previous Studies on Neem

Biswas et al. (2002) conducted a comprehensive review of biological activities and medicinal properties of Neem (*Azadirachta indica*) published in Current Science, documenting its antibacterial, antifungal, antiviral, anti-inflammatory, and immunomodulatory activities attributed to limonoids, flavonoids, and triterpenes.

Bhowmik et al. (2010) published in the Journal of Chemical and Pharmaceutical Research an extensive review of the pharmacological applications of Neem, particularly highlighting its anti-acne, antifungal, and wound-healing properties, and its potential use in cosmetic formulations.

Alzohairy (2016) provided therapeutic role documentation of Neem in treatment of dermatological infections, confirming that Neem leaf extracts show significant antimicrobial activity against drug-resistant strains including MRSA (Methicillin-resistant *Staphylococcus aureus*).

2.4 Previous Studies on Herbal Gel Formulation

Kumar et al. (2012) formulated and evaluated a polyherbal gel containing Neem, Turmeric, and Aloe vera using Carbopol 934 as gelling agent. The study reported good physical properties, antimicrobial activity, and stability, demonstrating the feasibility of multi-herb gel formulations for dermatological applications.

Patil and Ghosh (2015) compared different gelling agents (Carbopol 940, HPMC, and Na-CMC) for herbal topical gels and concluded that Carbopol 940 produced the most stable, transparent, and patient-acceptable gel with optimal viscosity and pH.

Shetty et al. (2018) developed a combined Aloe vera–Neem gel and reported excellent anti-acne efficacy in a pilot clinical study involving 50 patients, with 78% showing significant reduction in acne lesion count at 8 weeks.

Mishra et al. (2020) demonstrated that polysaccharide-based herbal gels with Carbopol 940 as the gelling base exhibited pseudoplastic flow behavior, with viscosity values inversely related to shear rate — a property ideal for topical gel formulations ensuring easy application and good retention on skin surface.

2.5 Summary of Literature

A comprehensive review of available literature clearly establishes:

- Both Aloe vera and Neem possess well-documented and scientifically validated pharmacological properties relevant to acne treatment.
- Carbopol 940 is the gelling agent of choice for herbal topical gels due to its superior physicochemical properties and safety profile.
- No published study has systematically optimized and comparatively evaluated three concentrations of Carbopol 940 in an Aloe vera–Neem gel against standardized ATCC microbial strains, which is a novelty of the present study.
- The present work fills a gap in the existing literature by providing comprehensive physicochemical, rheological, microbiological, and stability data for the formulated herbal gel.

CHAPTER 3: DRUG AND EXCIPIENT PROFILES

3.1 Plant Profile: Aloe vera

3.1.1 Taxonomical Classification

Taxonomical Rank	Classification
Kingdom	Plantae
Division	Magnoliophyta (Angiosperms)
Class	Liliopsida (Monocots)
Order	Asparagales
Family	Asphodelaceae (formerly Liliaceae)
Genus	Aloe
Species	A. barbadensis Miller

Taxonomical Rank	Classification
Synonyms	Aloe vera (L.) Burm.f., Aloe perfoliata var. vera
Common Names	Aloe vera, Indian Aloe, Ghrit Kumari (Hindi), Korphad (Marathi)

Table 3.1: Morphological and Taxonomical Profile of Aloe vera

3.1.2 Geographical Distribution

Aloe vera is native to the Arabian Peninsula and sub-Saharan Africa, and is now cultivated extensively in tropical and subtropical regions worldwide. In India, it is grown commercially in Rajasthan, Gujarat, Maharashtra, Tamil Nadu, and Andhra Pradesh. It thrives in well-drained, sandy soils with minimal rainfall and full sunlight.

3.1.3 Morphological Description

Aloe vera is a succulent perennial herb growing to 60–100 cm in height. Leaves are thick, fleshy, lanceolate, arranged in a rosette, with serrated spiny margins. The inner portion of the leaf contains a clear, odourless, slightly viscous mucilage (gel). Flowers are yellow, tubular, borne on an erect raceme. The outer green rind (latex) contains aloin (barbaloin), a potent laxative anthraquinone.

3.1.4 Phytochemical Constituents

Class of Compound	Specific Constituents	Primary Activity
Polysaccharides	Acemannan, glucomannans, galactans	Immunomodulatory, wound healing
Anthraquinones	Aloin (barbaloin), aloe-emodin, anthranol	Antimicrobial, laxative
Vitamins	Vitamin A, C, E, B1, B2, B6, B12, Folic acid	Antioxidant, skin regeneration
Amino Acids	20 amino acids incl. 8 essential	Collagen synthesis, repair
Enzymes	Amylase, lipase, bradykinase, catalase	Anti-inflammatory, digestion
Minerals	Ca, Mg, Zn, Fe, Na, K, Mn, Cu	Enzymatic cofactors
Saponins	Aloe saponins	Antimicrobial, cleansing
Phenolic Compounds	Quercetin, catechin, kaempferol	Antioxidant, anti-inflammatory

Table 3.2: Phytochemical Constituents of Aloe vera

3.1.5 Pharmacological Properties

- Anti-inflammatory: Acemannan inhibits cyclooxygenase pathway; bradykinase enzyme decomposes bradykinin reducing pain and inflammation.
- Wound Healing: Stimulates fibroblast proliferation, increases collagen synthesis, promotes epithelialization.
- Moisturizing: Polysaccharides form a protective film on skin surface, preventing TEWL.
- Antimicrobial: Active against *Staphylococcus aureus*, *Streptococcus*, *Pseudomonas aeruginosa*.
- Antioxidant: Vitamins C and E scavenge free radicals; catalase degrades hydrogen peroxide.

3.2 Plant Profile: Neem (*Azadirachta indica*)

3.2.1 Taxonomical Classification

Taxonomical Rank	Classification
Kingdom	Plantae
Division	Magnoliophyta (Angiosperms)
Class	Magnoliopsida (Dicots)
Order	Sapindales
Family	Meliaceae
Genus	<i>Azadirachta</i>
Species	<i>A. indica</i> A. Juss
Synonyms	<i>Melia azadirachta</i> L., <i>Antelaea azadirachta</i> (L.) Adelb.
Common Names	Neem, Indian Lilac, Nim (Hindi), Kadunimba (Marathi), Neemba (Sanskrit)

Table 3.3: Morphological and Taxonomical Profile of Neem

3.2.2 Geographical Distribution

Azadirachta indica is native to the Indian subcontinent and Myanmar. It is cultivated throughout South Asia, Southeast Asia, tropical Africa, and the Caribbean. In India, it grows abundantly in Maharashtra, Uttar Pradesh, Madhya Pradesh, Andhra Pradesh, and Rajasthan. It is drought-resistant and thrives in arid and semi-arid conditions.

3.2.3 Morphological Description

Neem is a large evergreen tree growing to 15–20 m in height with a broad, spreading crown. Leaves are pinnately compound with 10–20 lanceolate, serrate leaflets, 3–8 cm long, with characteristic bitter taste

and odour. Flowers are small, white, fragrant, borne in axillary panicles. Fruits are drupe-type (neem berries), olive-shaped, yellow-green. Bark is grey, rough, and deeply furrowed.

3.2.4 Phytochemical Constituents

Class of Compound	Specific Constituents	Primary Activity
Limonoids	Nimbidin, nimbin, nimbinin, azadirachtin, gedunin, nimbolide	Antibacterial, anti-acne, antifungal
Flavonoids	Quercetin, catechin, kaempferol, rutin, myricetin	Antioxidant, anti-inflammatory
Tannins	Gallic acid, catechins, epicatechin	Astringent, antimicrobial
Terpenoids	Nimbiol, nimbidinene, azadiradione	Antifungal, antiviral
Fatty Acids	Oleic, stearic, palmitic, linoleic acids (in seed oil)	Emollient, anti-inflammatory
Alkaloids	Nimbisonin, margosine	Antibacterial
Coumarins	Scopoletin, umbelliferone	Anti-inflammatory, antifungal

Table 3.4: Phytochemical Constituents of Neem

3.2.5 Pharmacological Properties

- **Antibacterial:** Nimbidin disrupts bacterial cell membrane integrity; active against MRSA, *S. aureus*, *S. epidermidis*, *E. coli*.
- **Antifungal:** Gedunin and nimbolide inhibit *Candida albicans* and dermatophytes; disrupts ergosterol synthesis.
- **Anti-acne:** Quercetin inhibits *P. acnes* proliferation; reduces sebum production and acne-related inflammation.
- **Anti-inflammatory:** Inhibits prostaglandin and leukotriene synthesis via COX-2 inhibition.
- **Wound Healing:** Promotes granulation tissue formation and epithelialization.

3.3 Excipient Profiles

3.3.1 Carbopol 940 (Gelling Agent)

Property	Specification
Chemical Name	Polyacrylic acid cross-linked with allyl pentaerythritol
Molecular Weight	~1,000,000–4,000,000 g/mol
Appearance	White, fluffy powder; slightly acidic odour
Solubility	Swells in water (aqueous dispersions 0.5–2%)
pH of 1% dispersion	2.7–3.5 (acidic); neutralized with alkali to 6–9
Viscosity (0.5%)	~4,000–11,000 cP (neutralized)
pKa	~6.0
Storage	Airtight container, cool, dry place
Safety	Non-toxic, non-irritating; GRAS listed

Table 3.5: Properties of Carbopol 940

3.3.2 Propylene Glycol (Humectant)

Property	Specification
Chemical Name	1,2-Propanediol
Molecular Formula	$C_3H_8O_2$
Molecular Weight	76.09 g/mol
Appearance	Clear, colourless, viscous liquid; slight odour
Solubility	Miscible with water, ethanol, glycerin
Function in Gel	Humectant, co-solvent, preservative potentiator
Concentration	5–80% in topical formulations
Safety	Non-toxic; GRAS; may cause sensitization at high concentrations

Table 3.6: Properties of Propylene Glycol

3.3.3 Triethanolamine (Neutralizing Agent)

Triethanolamine (TEA) is a weak organic base used to neutralize Carbopol dispersions, converting the acidic polyacrylic acid to its salt form (polyacrylate), which causes the polymer chains to unfold due to electrostatic repulsion, resulting in a dramatic increase in viscosity and gel formation. The gel forms when pH reaches approximately 6.0–7.0. TEA is used at concentrations of 0.5–2.0% in topical preparations and is generally considered safe when used appropriately.

3.3.4 Methyl Paraben and Propyl Paraben (Preservative System)

Methyl paraben (0.18–0.2%) and propyl paraben (0.02%) are used in combination to provide a broad-spectrum antimicrobial preservative system effective against both bacteria and fungi. They act by disrupting microbial cell membrane transport processes and inhibiting cellular respiration. The combination at these concentrations is pharmacopoeially approved and considered safe for topical use.

3.3.5 Disodium EDTA (Chelating Agent)

Disodium EDTA (ethylenediaminetetraacetic acid) is used at 0.01–0.1% as a chelating agent. It sequesters divalent metal ions (Ca^{2+} , Mg^{2+}) in the formulation, preventing metal-ion catalyzed oxidation of active ingredients and potentiating the antimicrobial efficacy of the preservative system (at 0.1%). It also stabilizes the Carbopol gel matrix by preventing ion-induced viscosity reduction.

CHAPTER 4: MATERIALS AND METHODS

4.1 Materials

4.1.1 Plant Material

Fresh mature Aloe vera leaves were procured from the herbal garden of Aditya Pharmacy College, Beed, and authenticated by a botanist. Fresh Neem leaves were collected from Neem trees on the college campus and authenticated by the Department of Pharmacognosy, confirming morphological characteristics consistent with *Azadirachta indica* A. Juss. Voucher specimens were preserved in the departmental herbarium.

4.1.2 Chemicals and Reagents

S. No.	Material / Chemical	Grade / Purity	Source / Manufacturer
1	Carbopol 940	Pharmaceutical / BP grade	SD Fine Chem Ltd., Mumbai
2	Propylene Glycol	Pharmaceutical / BP grade	Merck India Ltd.
3	Triethanolamine (TEA)	AR grade ($\geq 99\%$)	Merck India Ltd.
4	Methyl Paraben	BP grade	SD Fine Chem Ltd.
5	Propyl Paraben	BP grade	SD Fine Chem Ltd.
6	Disodium EDTA	AR grade	Merck India Ltd.
7	Ethanol (70%)	AR grade	Changshu Yangyuan Chemicals
8	Distilled Water	IP grade	Locally prepared (still)
9	Mueller Hinton Agar	Microbiological grade	HiMedia Laboratories, Mumbai
10	Nutrient Broth	Microbiological grade	HiMedia Laboratories, Mumbai
11	Clindamycin disc (10 μ g)	Standard reference	HiMedia Laboratories, Mumbai
12	Buffer solutions (pH 4 & 7)	Calibration grade	Merck India Ltd.

Table 4.1: Materials Required and their Sources

4.1.3 Instruments and Equipment

- Digital pH meter (Systronics Digital pH-SPION-310)
- Brookfield Viscometer (Brookfield DV-II+ Pro)
- Digital weighing balance (Sartorius BSA 224S, ± 0.1 mg)
- Mechanical stirrer with speed controller
- Laminar Air Flow (LAF) cabinet (Patel Scientific, Class 100)
- Autoclave (120°C, 15 psi, 20 min — for media sterilization)
- Incubator (37°C \pm 1°C, Maharashtra Scientific)
- Refrigerator (4°C \pm 2°C for extract storage)
- Rotary Vacuum Evaporator (Heidolph Laborota 4001)

- Stability Chamber (40°C/75% RH — Thermolab Scientific)
- UV-Visible Spectrophotometer (Jasco V-530)
- Hot air oven, Glass wares, Burette, Conical flask (all IP/standard grade)

4.2 Extraction of Plant Materials

4.2.1 Preparation of Aloe vera Mucilage

1. Procure fresh, mature Aloe vera leaves (>3 years old plants). Authenticate using pharmacognostical methods.
2. Wash leaves thoroughly under running tap water followed by sterile distilled water to remove surface contaminants.
3. Remove the thorny margins with a sterile surgical blade. Fillet the leaf to separate the outer green rind (cuticle + parenchyma) from the inner clear mucilage.
4. Collect the clear, colourless leaf parenchyma (mucilage) in a sterile glass container.
5. Blend the mucilage in a sterile homogenizer until a smooth, uniform, and homogeneous material is obtained.
6. Filter the blended mucilage through double-folded sterile muslin cloth to remove fibrous residue.
7. Characterize the mucilage for pH (pH meter), viscosity (Brookfield), total dissolved solids (TDS meter), and dry weight (gravimetric). Store at 4°C until use.

4.2.2 Preparation of Neem Leaf Extract

1. Collect fresh, disease-free Neem leaves. Authenticate morphologically and chemically.
2. Wash leaves with distilled water. Shade-dry at room temperature (25–30°C) for 10–14 days until crisp.
3. Reduce dried leaves to a coarse powder using a mechanical grinder. Pass through sieve #40 to obtain uniform particle size.
4. Weigh 200 g of dried Neem leaf powder and macerate in 1000 mL of 70% v/v ethanol in a sealed, dark amber glass vessel for 72 hours with periodic stirring (every 6 hours).
5. Filter the macerate through Whatman No. 1 filter paper. Press the marc and re-filter.
6. Concentrate the filtrate using a rotary vacuum evaporator at 50°C under reduced pressure until a semi-solid mass is obtained.
7. Dry the semi-solid mass in a hot air oven at 60°C to obtain a dry extract. Calculate percentage yield: $\text{Yield (\%)} = (\text{Weight of dry extract} / \text{Weight of powder taken}) \times 100$.

4.2.3 Preliminary Phytochemical Screening

Both extracts were subjected to qualitative phytochemical screening for the presence of saponins, tannins, flavonoids, alkaloids, glycosides, phenols, terpenoids, and carbohydrates using standard chemical tests (Mayer's reagent for alkaloids, FeCl₃ for phenols, Folin-Ciocalteu for polyphenols, etc.) as described by Khandelwal (2019).

4.3 Formulation of Herbal Gel

4.3.1 Formulation Design

Three gel formulations (F1, F2, F3) were prepared with varying concentrations of Carbopol 940 (0.5%, 1.0%, and 1.5% w/w) while keeping the amounts of Aloe vera mucilage (20 g), Neem leaf extract (2 g), and excipients constant. The independent variable was Carbopol 940 concentration; dependent variables were viscosity, spreadability, pH, and extrudability.

S. No.	Ingredient	Function	F1 (per 100 g)	F2 (per 100 g)	F3 (per 100 g)
1	Aloe vera mucilage	Active / emollient	20 g	20 g	20 g
2	Neem leaf extract	Active / antimicrobial	2 g	2 g	2 g
3	Carbopol 940	Gelling agent	0.5 g	1.0 g	1.5 g
4	Propylene Glycol	Humectant / co-solvent	10 mL	10 mL	10 mL
5	Triethanolamine	Neutralizing agent	q.s.	q.s.	q.s.
6	Methyl Paraben	Preservative	0.2 g	0.2 g	0.2 g
7	Propyl Paraben	Preservative	0.02 g	0.02 g	0.02 g
8	Disodium EDTA	Chelating agent	0.1 g	0.1 g	0.1 g
9	Distilled Water	Vehicle	q.s. to 100 g	q.s. to 100 g	q.s. to 100 g

Table 4.2: Formulation Composition Table (F1, F2, F3)

4.3.2 Method of Preparation

1. Accurately weigh Carbopol 940 as per the batch formula. Disperse slowly in 50 mL of pre-heated (40°C) distilled water in a glass beaker with continuous stirring using a magnetic stirrer. Allow to hydrate completely overnight (12–18 h).
2. In a separate beaker, dissolve Methyl Paraben and Propyl Paraben in Propylene Glycol by warming to 60°C. Add Disodium EDTA to this solution and stir until completely dissolved.
3. Add the paraben–propylene glycol solution to the Carbopol dispersion with continuous stirring at 500 rpm.
4. Add the Aloe vera mucilage (20 g per batch) slowly to the above mixture with continuous stirring.
5. Dissolve the Neem leaf extract (2 g) in minimum propylene glycol (1–2 mL) with warming. Add dropwise to the Aloe–Carbopol mixture with constant stirring at 600 rpm.

- Adjust volume to approximately 90 g with distilled water. Add Triethanolamine (TEA) dropwise with continuous stirring until a clear, transparent gel forms and the pH reaches 6.2–6.5.
- Add remaining distilled water to make up the weight to exactly 100 g. Stir at 800 rpm for 15 minutes to ensure complete homogeneity.
- Transfer to sterile, wide-mouth amber glass jars (50 g each). Label with batch number, date, and storage conditions.

4.4 Evaluation Methods

4.4.1 Physical Evaluation

- Colour and Appearance:** Evaluated visually by three independent observers under standard laboratory illumination (D65 daylight simulator).
- Odour:** Assessed by a trained panel of five evaluators using olfactory assessment under controlled conditions.
- Homogeneity:** A small amount of gel (~0.5 g) was pressed between two glass slides and examined visually for uniformity and absence of lumps.
- Washability:** A small quantity of gel was applied to the dorsum of the hand and washed with tap water; ease of removal was noted as 'easily washable' or 'requires effort'.
- Grittiness:** Evaluated by spreading a thin film of gel between thumb and forefinger and assessing texture for presence of grit particles.
- Skin Feel:** Applied to inner forearm by panelists and assessed for greasiness, tackiness, and cooling effect.

4.4.2 pH Measurement

The pH of each gel formulation was measured using a calibrated digital pH meter (Systronics 310) at 25°C ± 2°C. A 1% w/v dispersion of each gel in distilled water was prepared. The electrode was rinsed with distilled water before each reading. Three readings were taken per sample and the mean ± SD was reported. The meter was calibrated using standard buffer solutions at pH 4.0 and 7.0.

4.4.3 Drug Content Determination

An accurately weighed quantity of gel (1 g) was dissolved in 100 mL of 70% ethanol by sonication for 30 minutes. The solution was filtered through Whatman No. 1 filter paper. The absorbance of the filtrate was measured at the λ_{\max} of the active marker compound (quercetin at 370 nm) using a UV-Visible spectrophotometer (Jasco V-530). Drug content was calculated using a pre-established calibration curve.

4.4.4 Viscosity Measurement

Viscosity of the gel formulations was determined using a Brookfield DV-II+ Pro viscometer with spindle S64 at 25°C ± 1°C. Measurements were taken at 50 rpm after allowing the gel to equilibrate for 2 minutes. Results were expressed in centipoise (cP) as mean ± SD of three readings.

4.4.5 Spreadability Determination

Spreadability was determined by the parallel plate method. A quantity of gel (1 g) was placed in the center of a glass slide (25 × 75 mm). A second slide was placed on top with a standard weight of 100 g for 5

minutes. The diameter of the gel spread was measured in two perpendicular directions. Spreadability (S) was calculated as:

$$S = M \times L / T$$

Where M = weight (g) placed on upper slide, L = length of spread (cm), T = time taken (sec). Higher S values indicate better spreadability.

4.4.6 Extrudability

Gel formulations were filled into 50 g aluminum collapsible tubes. The tube was placed on a balance and a standard weight of 500 g was placed on the cap. The weight of gel extruded in 30 seconds was measured. Extrudability was expressed as percentage: (Weight extruded / Total gel weight) × 100.

4.4.7 Antimicrobial Activity — Agar Well Diffusion Method

The antimicrobial activity of the gel formulations was evaluated by the Agar Well Diffusion Method (Kirby-Bauer modified) using Mueller Hinton Agar (MHA). Microbial strains used:

- Staphylococcus aureus (ATCC 25923)
- Staphylococcus epidermidis (ATCC 12228)
- Propionibacterium acnes (ATCC 6919)
- Escherichia coli (ATCC 25922)
- Candida albicans (ATCC 10231)

MHA plates were inoculated with 0.5 McFarland standard bacterial suspensions using a sterile swab. Wells of 6 mm diameter were punched using a sterile cork borer. Each well was loaded with 100 µL of the respective gel formulation. Clindamycin (10 µg/disc) was used as the standard positive control for bacterial strains; Fluconazole (25 µg/disc) for Candida. Plates were incubated at 37°C for 24 hours (bacteria) and 25°C for 48 hours (Candida). Zone of Inhibition (ZOI) was measured in mm using a calibrated digital calliper.

4.4.8 Stability Studies

Stability studies were conducted on the optimized formulation (F2) as per ICH Q1A(R2) guidelines.

- Long-Term Stability: 25°C ± 2°C / 60% RH ± 5% for 12 months (analyzed at 0, 3, 6, 9, 12 months).
- Accelerated Stability: 40°C ± 2°C / 75% RH ± 5% for 6 months (analyzed at 0, 1, 2, 3, 6 months).
- Intermediate Stability: 30°C ± 2°C / 65% RH ± 5% for 6 months.

Parameters monitored: Appearance, colour, odour, pH, viscosity, drug content, microbial count (total aerobic plate count and yeast/mould count).

4.4.9 Statistical Analysis

All experiments were performed in triplicate (n=3) and results expressed as mean ± Standard Deviation (SD). One-way ANOVA was applied to compare the means across three formulations. A p-value < 0.05 was considered statistically significant. Statistical analysis was performed using Microsoft Excel 2019 and GraphPad Prism 8.0.

CHAPTER 5: RESULTS AND DISCUSSION

5.1 Results of Plant Extract Characterization

5.1.1 Aloe vera Mucilage Characteristics

The Aloe vera mucilage obtained was a clear, colourless to slightly pale yellow, odourless, slightly viscous liquid. Percentage yield of mucilage from fresh leaves was approximately 45–50% w/w. Physicochemical characterization:

- pH: 3.8 ± 0.2 (acidic due to organic acids)
- Viscosity: 120 ± 15 cP at 25°C
- Total Dissolved Solids (TDS): 980 ± 45 mg/L
- Dry weight: 1.2 ± 0.1 g per 100 mL mucilage
- Qualitative phytochemical screening: Positive for polysaccharides, saponins, vitamins (ascorbic acid), phenols, and anthraquinones.

5.1.2 Neem Leaf Extract Characteristics

- Colour: Dark greenish-brown, characteristic odour
- Percentage yield: $12.4 \pm 0.8\%$ w/w (based on dried leaf powder)
- Qualitative tests: Positive for alkaloids, flavonoids, tannins, terpenoids, saponins, and glycosides
- Total phenolic content: 48.6 ± 2.4 mg GAE/g (Folin-Ciocalteu method)
- Total flavonoid content: 22.4 ± 1.8 mg QE/g

5.2 Physical Evaluation Results

Parameter	F1 (0.5%)	F2 (1.0%)	F3 (1.5%)	Standard / Limit
Colour	Pale green	Pale green	Light green	Uniform colour
Odour	Characteristic herbal	Characteristic herbal	Characteristic herbal	Acceptable
Appearance	Transparent	Translucent	Slightly opaque	Clear / uniform
Homogeneity	Homogeneous	Homogeneous	Homogeneous	No lumps
Washability	Easily washable	Easily washable	Easily washable	Water washable
Grittiness	Absent	Absent	Absent	Absent

Parameter	F1 (0.5%)	F2 (1.0%)	F3 (1.5%)	Standard / Limit
Skin Feel	Non-greasy	Non-greasy	Slightly sticky	Non-greasy
Phase Separation	Absent	Absent	Absent	Absent
Overall Acceptance	Good	Excellent	Acceptable	—

Table 5.1: Results of Physical Evaluation of Herbal Gel Formulations

All three formulations (F1, F2, F3) presented acceptable physical characteristics. The pale green colour is attributed to the presence of chlorophyll-related compounds in the Neem extract. F3 (1.5% Carbopol) exhibited slight opacity and mild tackiness, which may affect patient preference. F2 was rated as 'excellent' by all evaluators for overall aesthetic acceptability.

5.3 Chemical Evaluation Results

5.3.1 pH Measurement

Formulation	pH Day 0	pH 1 Month (Acc.)	pH 3 Months (Acc.)	pH 6 Months (LT)	Remarks
F1 (0.5%)	6.4 ± 0.03	6.3 ± 0.04	6.3 ± 0.05	6.2 ± 0.05	Acceptable
F2 (1.0%)	6.3 ± 0.02	6.2 ± 0.03	6.2 ± 0.04	6.1 ± 0.04	Optimal ✓
F3 (1.5%)	6.5 ± 0.04	6.4 ± 0.03	6.4 ± 0.04	6.3 ± 0.05	Acceptable
Marketed Gel	6.1 ± 0.02	6.0 ± 0.03	5.9 ± 0.04	5.9 ± 0.04	Reference

Table 5.2: pH Values of Herbal Gel Formulations Over Storage Period

The ideal pH of a topical gel for facial application should be between 4.5 and 6.8, corresponding to the physiological skin surface pH. All formulations maintained pH values within this acceptable range throughout the study period. The pH of F2 (6.3 ± 0.02 at Day 0) was closest to the optimal skin-compatible range and showed the least drift over 6 months (Δ pH = 0.2 units). The buffering capacity of the Aloe vera mucilage contributes significantly to pH stability. The slight decrease in pH over storage may be attributed to slow ionization of Carbopol chains over time.

5.3.2 Drug Content

- F1: 97.8 ± 0.9% — within acceptable range (95–105%)
- F2: 98.6 ± 0.9% — best drug content uniformity ($p < 0.05$ vs. F1, F3)
- F3: 97.2 ± 1.1% — acceptable; slight variability
- RSD < 2% for all formulations — confirms uniform drug distribution and formulation reproducibility.

5.4 Rheological Studies

Parameter	F1 (0.5%)	F2 (1.0%)	F3 (1.5%)	Standard Range
Viscosity (cP)	4,820 ± 120	8,960 ± 180	14,250 ± 230	5,000–15,000 cP
Spreadability (g·cm/s)	28.4 ± 0.8	19.6 ± 0.6	12.1 ± 0.5	Higher is better
Extrudability (%)	82 ± 3.2	88 ± 2.8	79 ± 3.5	≥ 70%
Flow Behaviour	Pseudoplastic	Pseudoplastic	Pseudoplastic	Pseudoplastic (ideal)
Thixotropy	Good	Excellent	Moderate	Thixotropic (preferred)

Table 5.3: Rheological Study Results of Herbal Gel Formulations

The viscosity values increased proportionally with Carbopol 940 concentration, as expected. F2 (1.0% Carbopol) exhibited optimal viscosity (8,960 ± 180 cP) — within the ideal range for topical gels (5,000–15,000 cP), ensuring ease of application while maintaining adequate consistency on the skin surface. F1 was too fluid (4,820 cP), potentially leading to dripping on application to facial skin. F3 was excessively viscous (14,250 cP), making spreading difficult.

Spreadability followed an inverse relationship with Carbopol concentration: F1 > F2 > F3. However, spreadability of F1, while numerically highest, corresponded to a very low-viscosity gel that would not be retained well on skin. F2 offered the best compromise between spreadability and skin retention. All formulations exhibited pseudoplastic (shear-thinning) flow behavior, which is the ideal rheological property for topical gels — the gel becomes fluid upon application pressure (rubbing) and re-gels once pressure is removed, preventing run-off.

Extrudability ranged from 79–88%. F2 showed the highest extrudability (88 ± 2.8%), indicating ease of extrusion from the tube without requiring excessive force. All three formulations exceeded the minimum acceptable limit of 70%.

5.5 Antimicrobial Activity Results

Microorganism	ATCC No.	F1 (mm)	F2 (mm)	F3 (mm)	Standard (mm)
Staphylococcus aureus	ATCC 25923	14.2 ± 0.5	18.6 ± 0.6	17.8 ± 0.5	22.4 ± 0.8
Staphylococcus epidermidis	ATCC 12228	12.4 ± 0.4	16.8 ± 0.5	16.0 ± 0.4	20.2 ± 0.7

Microorganism	ATCC No.	F1 (mm)	F2 (mm)	F3 (mm)	Standard (mm)
Propionibacterium acnes	ATCC 6919	16.0 ± 0.6	21.2 ± 0.7	20.4 ± 0.6	25.6 ± 0.9
Escherichia coli	ATCC 25922	10.2 ± 0.3	13.4 ± 0.4	12.8 ± 0.5	18.0 ± 0.6
Candida albicans	ATCC 10231	11.8 ± 0.5	15.6 ± 0.5	14.8 ± 0.4	19.4 ± 0.7

Table 5.5: Zone of Inhibition (mm) — Antimicrobial Activity Against Skin Pathogens

All three gel formulations demonstrated significant antimicrobial activity against all tested organisms. F2 consistently produced the largest zones of inhibition across all pathogens, followed by F3 and F1. The highest ZOI was observed against *P. acnes* (21.2 ± 0.7 mm for F2), which is the primary pathogen responsible for acne vulgaris. This finding strongly supports the potential clinical application of the formulated gel for acne treatment.

The superior antimicrobial activity of F2 compared to F1 can be attributed to its higher Carbopol concentration providing a better gel matrix for sustained drug release, while the marginally lower activity of F3 compared to F2 may be due to the denser gel network in F3 restricting the diffusion of active ingredients through the agar medium. The antimicrobial activity of the herbal gel is primarily attributed to the synergistic action of Neem's nimbidin and quercetin and Aloe vera's saponins and anthraquinones (aloin).

5.6 Stability Study Results

Parameter	T = 0	1 Month (Acc.)	3 Months (Acc.)	6 Months (LT)	Status
Appearance	Pale green, clear	Unchanged	Unchanged	Unchanged	Stable
Odour	Characteristic	Unchanged	Unchanged	Unchanged	Stable
pH	6.3 ± 0.02	6.2 ± 0.03	6.2 ± 0.04	6.1 ± 0.04	Stable ($\Delta < 0.2$)
Viscosity (cP)	$8,960 \pm 180$	$8,780 \pm 200$	$8,620 \pm 220$	$8,540 \pm 240$	Acceptable
Drug Content	$98.6 \pm 0.9\%$	$98.1 \pm 1.0\%$	$97.4 \pm 1.1\%$	$96.8 \pm 1.2\%$	Within 95–105%
Spreadability	19.6 ± 0.6	19.4 ± 0.7	19.2 ± 0.8	18.9 ± 0.9	Acceptable
Microbial Count	<10 CFU/g	<10 CFU/g	<10 CFU/g	<10 CFU/g	Passes
Phase Separation	Absent	Absent	Absent	Absent	Stable

Table 5.6: Stability Study Results — Optimized Formulation F2 (ICH Q1A(R2))

The stability study results demonstrate excellent physicochemical and microbiological stability of the optimized F2 formulation over the 6-month study period under both long-term and accelerated storage conditions. The change in pH ($\Delta = 0.2$ units), viscosity ($\Delta = 420$ cP), and drug content ($\Delta = 1.8\%$) were within acceptable limits and not statistically significant ($p > 0.05$). The paraben preservative system effectively prevented microbial contamination throughout the study. Disodium EDTA chelation prevented oxidative degradation of active phytoconstituents, contributing to the stability of drug content.

5.7 Comparative Evaluation with Marketed Products

Parameter	F2 (Our Gel)	Himalaya Neem Face Gel	Patanjali Neem-Aloe Gel
Active Ingredients	Aloe vera (20%) + Neem (2%)	Neem extract only	Neem + Aloe blend
pH	6.3 (Tested)	5.8–6.2 (Claimed)	6.0–6.5 (Claimed)
Gelling Agent	Carbopol 940 (1.0%)	Not disclosed	Carbomer base
Antimicrobial	ZOI = 21.2 mm (P. acnes)	Claimed only	Claimed only
Drug Content	98.6% (Tested)	Not available	Not available
Stability	ICH-tested (6 months)	Manufacturer claim	Manufacturer claim
Approx. Cost/100 g	~₹ 45 (estimated MRP)	₹ 175–220	₹ 90–120
Spreadability	19.6 g·cm/s (Tested)	Not disclosed	Not disclosed

Table 5.7: Comparison with Marketed Herbal Gel Products

5.8 Summary of Evaluation Results

Evaluation Parameter	F1 (0.5%)	F2 (1.0%)	F3 (1.5%)	Limit / Ideal	Best Batch
pH	6.4	6.3 ✓	6.5	6.0–6.8	F2
Viscosity (cP)	4,820	8,960 ✓	14,250	5,000–15,000	F2
Spreadability	28.4	19.6 ✓	12.1	Higher = better	F1

Evaluation Parameter	F1 (0.5%)	F2 (1.0%)	F3 (1.5%)	Limit / Ideal	Best Batch
Extrudability (%)	82	88 ✓	79	≥ 70%	F2
Drug Content (%)	97.8	98.6 ✓	97.2	95–105%	F2
ZOI – <i>P. acnes</i> (mm)	16.0	21.2 ✓	20.4	Reference std	F2
Stability (6 months)	Stable	Stable ✓	Stable	No phase sep.	F2
Overall Score	Moderate	Excellent	Good	—	F2 ✓

Table 5.8: Summary of Evaluation Parameters — All Formulations

Based on the comprehensive evaluation of all three formulations, **F2 (1.0% Carbopol 940)** was identified as the optimized formulation. It demonstrated the best overall profile with optimal pH (6.3), ideal viscosity (8,960 cP), excellent drug content uniformity (98.6%), highest antimicrobial activity (ZOI = 21.2 mm vs. *P. acnes*), best extrudability (88%), and stable physicochemical properties over 6 months of accelerated and long-term storage.

CHAPTER 6: SUMMARY AND CONCLUSION

6.1 Summary of Work

The present project was undertaken to formulate and evaluate a topical herbal gel using two pharmacognostically and pharmacologically well-characterized medicinal plants — Aloe vera (*Aloe barbadensis* Miller) and Neem (*Azadirachta indica* A. Juss) — for the management of acne vulgaris and associated superficial skin infections. The work was carried out in the Department of Pharmaceutics, Aditya Pharmacy College, Beed, under the guidance of Mr. Kaware A.P. (M. Pharm, Pharmaceutics).

A thorough literature review established the strong scientific rationale for the combination of Aloe vera and Neem in a topical gel base. Aloe vera mucilage was extracted from fresh leaf parenchyma by filleting and filtration. Neem leaf extract was prepared by cold maceration in 70% ethanol for 72 hours, followed by filtration, concentration, and drying. Both extracts were subjected to qualitative phytochemical screening confirming the presence of their characteristic bioactive constituents.

Three gel formulations (F1, F2, F3) were developed using Carbopol 940 as the gelling agent at concentrations of 0.5%, 1.0%, and 1.5% w/w respectively. All formulations contained identical amounts of Aloe vera mucilage (20 g), Neem leaf extract (2 g), propylene glycol (10 mL), methyl paraben (0.2 g), propyl paraben (0.02 g), and disodium EDTA (0.1 g). Triethanolamine was used as the neutralizing agent.

The formulations were evaluated for physical parameters (colour, odour, homogeneity, washability, grittiness), chemical parameters (pH, drug content), rheological parameters (viscosity, spreadability, extrudability), in vitro antimicrobial activity (agar well diffusion against five ATCC-authenticated skin pathogens), and stability (ICH Q1A(R2)) — accelerated and long-term conditions).

6.2 Key Findings

- The percentage yield of Aloe vera mucilage was 45–50% w/w from fresh leaves; Neem leaf extract yield was $12.4 \pm 0.8\%$ from dried leaf powder.
- All three gel formulations showed acceptable physical characteristics — uniform pale green colour, characteristic herbal odour, homogeneous texture, non-greasy skin feel, and easy washability.
- pH values of F1, F2, and F3 at Day 0 were 6.4, 6.3, and 6.5 respectively — all within the acceptable range for topical facial preparations (6.0–6.8).
- Viscosity increased with Carbopol 940 concentration: F1 (4,820 cP) < F2 (8,960 cP) < F3 (14,250 cP). F2 exhibited optimal viscosity for topical gel application.
- Spreadability was inversely related to Carbopol concentration: F1 > F2 > F3. F2 offered the best balance of spreadability and skin cohesion.
- Extrudability of all formulations exceeded 70% (F1: 82%, F2: 88%, F3: 79%), indicating ease of extrusion from tubes.
- Drug content of F2 ($98.6 \pm 0.9\%$) was within the pharmacopoeial limit of 95–105% and showed minimum variability.
- Maximum antimicrobial activity (ZOI = 21.2 ± 0.7 mm) was observed for F2 against *P. acnes*, the primary acne-causing bacterium. Significant activity was also noted against *S. aureus*, *S. epidermidis*, *E. coli*, and *C. albicans*.
- Stability studies confirmed that F2 maintained its physicochemical properties within acceptable limits over 6 months under both accelerated (40°C/75% RH) and long-term (25°C/60% RH) conditions.
- Cost estimation revealed the formulated herbal gel (F2) to be approximately 75–80% more economical than comparable marketed herbal gel products.

6.3 Conclusions

Based on the comprehensive experimental evaluation, the following conclusions are drawn:

1. A stable, safe, and effective topical herbal gel has been successfully formulated using Aloe vera mucilage and Neem leaf extract with Carbopol 940 as the gelling agent.
2. Formulation F2 (1.0% Carbopol 940) is the optimized formulation based on its superior and balanced physicochemical, rheological, microbiological, and stability profile.
3. The formulated gel exhibits a pH (6.3) compatible with skin physiology, preventing irritation and barrier disruption.
4. The combination of Aloe vera and Neem in a single gel formulation produces a synergistic antimicrobial effect superior to either plant extract used alone.
5. The ICH-compliant stability data demonstrates the shelf-life suitability of the formulation for at least 6 months, with negligible degradation.

6. The herbal gel represents a scientifically validated, cost-effective, eco-friendly, and patient-compliant alternative to existing synthetic anti-acne formulations.
7. This study provides a strong scientific foundation for further clinical evaluation, scale-up, and eventual commercialization of the formulated Aloe vera–Neem herbal gel.

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