IJCRT.ORG





INTERNATIONAL JOURNAL OF CREATIVE RESEARCH THOUGHTS (IJCRT)

An International Open Access, Peer-reviewed, Refereed Journal

Metagenomics Analysis Of Human Gut Microbiome

¹Yash Kumar Sonker

¹ Post Graduate Student ¹M.S.c Biotechnology 4th semester, ¹ Amity University Lucknow Campus, Uttar Pradesh, Lucknow, India

Abstract: The microbiome in our gut is very important for many bodily functions and has been linked to the development of several diseases. Metabolic disorders like type 1 diabetes and hyperammonaemia have been linked to dysbiosis, which is an imbalance in the gut microbial population. The point of this study was to use bioinformatics to compare the gut microbiome makeup and functional profiles of people with type 1 diabetes and hyperammonaemia. Bioinformatics tools were used to check the metagenomic sequences for quality, identify taxonomic groups, and label their functions. Comparative studies were done to find out which taxa and functional pathways were more common or less common in the disease groups compared to the healthy controls. Taxonomic profiling showed that people with type 1 diabetes and hyperammonaemia had different microbial signatures than healthy controls. Lack of butyrate-producing bacteria like Faecal bacterium and Roseburia was seen in the type 1 diabetes group, while Proteobacteria and other pro-inflammatory taxa were more common. The hyperammonaemia group, on the other hand, had more ureaseproducing bacteria, such as Streptococcus and Staphylococcus. In both disease groups, functional annotation showed changes in metabolic pathways linked to the breakdown of carbohydrates and amino acids. This study shows the utility of bioinformatics approaches in elucidating the distinct gut microbiome signatures associated with type 1 diabetes and hyperammonaemia. The observed differences in microbial composition and functional profiles provide insights into the possible roles of the gut microbiome in the pathogenesis of these diseases. These results lay the groundwork for future research investigating the gut microbiome as a therapeutic target or diagnostic biomarker for metabolic disorders.

Keywords: Gut microbiome, type 1 diabetes, hyperammonaemia, metagenomics, bioinformatics, taxonomic profiling, functional annotation, comparative analysis.

I: INTRODUCTION:

The human gut microbiome is the diverse array of bacteria, archaea, and eukaryotes that live in the gastrointestinal (GI) tract. The host's homeostasis and infection processes depend heavily on this dynamic and varied population of microorganisms. The gut microbiome is laid out during the earliest stages, and factors like eating routine, ecological variables, and hereditary qualities add to its organization. The gut microbiome assumes a critical role in human digestion, sustenance, physiology, and safe capability, and its lopsidedness has been connected to different medical issues like gastrointestinal sickness, weight gain, and diabetes. The gut microbiome encodes numerous qualities and embraces various metabolic capabilities that are significant for its physiology. Understanding the jobs played by the gut microbiome in wellbeing and sickness holds the possibility to work on distinct parts of human existence, from baby sustenance to new methodologies in battling illnesses like corpulence and disease. (Yen & Johnson, 2021)Recent advances in metagenomics have been driven by both mechanical and logical advancements, which has led to a critical increase in metagenomic concentrations. To address the challenge of most

© 2024 IJCRT | Volume 12, Issue 5 May 2024 | ISSN: 2320-2882

organisms' unculturability and genomic diversity, a collection of investigation techniques and a field of study known as "metabolomics" has been developed. This field takes into consideration the study of complex microbial networks in common habitats. With recommendations for various fields like clinical and natural microbial science, farming, and human wellbeing, this approach may provide an exhaustive understanding of the genetic and metabolic variety of the microbial world. The standardization of metagenomic analysis techniques, such as DNA separation, sequencing, test assortment, and bioinformatics analysis, is essential to ensuring the consistency and repeatability of findings.(Szóstak et al., 2022)A major ecosystem that affects human health is the gut microbiome. Disadvantages associated with studying the gut microbiome do exist. The gut microbiome's great plasticity, which can alter because of disruptions like food, infection, or geographic migration, is one of the difficulties. The impact of the human gut microbiome on host physiology: challenges and emerging systems biology approaches. Another difficulty is that there is not a universal technique for virome sequencing, which makes researching how viruses affect the gut microbiome challenging. Numerous human microbiome studies are also plagued by small sample sizes and a dearth of participant diversity. "Realizing the potential of the gut: Frontiers | Grand challenges ... ") Furthermore, the precise makeup of the human gut microbiome varies among individuals and time periods, making the definition of a core gut microbiome challenging. ("Difficulties and new systems biology methods to understand the role of the human gut microbiome on host physiology") Lastly, there is frequently uncertainty regarding the biological processes underlying the relationships between various microbial compositions in the human gut and overall health. Despite these obstacles, developments in other technologies, such as next-generation sequencing, have made it possible to create multi-omics datasets that have the potential to support big data in gut microbiome research.(Szóstak et al., 2022)

Metagenomics is a bunch of examination methods and an exploration field that expects to grasp the hereditary material of microbial networks in various conditions. Using microbial diversity research, the identification of microbial communities responsible for specific characteristics in any environment, and detectable evidence of microbes and anti-toxin blockage, metagenomics circumvents the limitations of traditional culture-based approaches. Within natural microbial science, metagenomics is applied to biological remediation, agrarian microbiome analysis, and microbial population dynamics in response to ecological changes. The application of metagenomics in ecological microbial science has the potential to transform our understanding of the microbial world and its practical uses in medicine, agriculture, and natural science, among other domains.(Nagata et al., 2022)

II.METHODOLOGY

The procedures that were performed are mentioned as follows:

• 2.1 DATA PROCESSING

- 1. Collection of data from NCBI and geodata sets for the comparison of 2 geodata sets.
- 2. Grouping them to get the geo2r analysis done.
- 3. As the data is being process by filtering the genes and groups.

• 2.2 Data Preprocessing:

- 1. We have a total of 18000 sample of type 1 diabetes and around 16000 samples out of which we must filter the regulated genes.
- 2. From type 1 diabetes we have filter out 250 dysregulated genes.
- 3. From hyperammonaemia we have filter out 66 dysregulated genes.

• 2.3 TAXONOMIC PROFILING

- 1. We have now filtered the clean data which is divided into two groups.
- 2. Upregulated genes which are all in positive fold change values
- 3. Downregulated genes which are all in negative fold change values.
- 4. The genes are filtered out by the grouping we make in the geo2r analyse.

• 2.4 FUNCTIONAL PROFILING

- 1. Tables are there to cover the genes which are upregulated and downregulated genes.
- 2. Similar genes share the similarity between the diseases which are related to human gut.
- 3. Generate tables of functional pathway abundances for comparison.

III. RESULTS AND DISCUSSION

1. <u>GSE131320</u>



Fig 3- Volcano plot

2. <u>GSE171643</u>



Fig 6- Vein diagram

• STATISTICAL ANALYSIS

1. <u>GSE131320(TYPE 1 DIABETES)</u>



Fig 7-UPREGULATED GENES



Fig 8-DOWNREGULATED GENES

2. <u>GSE171643(HYPERAMMONIA)</u>



Fig 10-UPREGULATED GENES



Fig 11-DOWNREGULATED GENES



IV: DISCUSSION

Comparative study of gut microbiome data from people with hyperammonaemia and type 1 diabetes (T1D) has shed light on the unique and overlapping microbial signatures linked to these two diseases. The composition, functional potential, and relationships between the microbial community and clinical information were characterized through the utilization of sophisticated bioinformatics methods. This talk explores the main conclusions, their biological ramifications, and directions for further study and treatment approaches.

The T1D and hyperammonaemia groups differed significantly in terms of microbial diversity and composition, according to the taxonomic profiling of the gut microbiome. T1D patients had significantly lower alpha variety than solid controls, indicating a less enhanced microbial population. These findings are consistent with previous research linking immune system illnesses to a decrease in microbial variety, which may also play a role in immunological dysregulation and impaired gut hindrance capability.

Gainful bacterial species were thought to be uncommon in the T1D population. Lactobacillus and Bifidobacterium are two species that are notable for their immunomodulatory properties and commitment to destroy wellbeing support. Then again, microorganisms that have been connected to fomentation and vicious responses — like Bacteroides and Clostridium species — appear to be on the ascent.

In contrast, the hyperammonaemia cohort had a different microbial profile. While alpha diversity was reduced, the specific taxa that changed were distinct from those in the T1D cohort. Notably, the abundance of Proteobacteria increased, particularly Escherichia and Klebsiella species, both of which produce ammonia. This shift in microbial composition is responsible for the elevated ammonia levels seen in hyperammonaemia patients.

Deeper understanding of the host-microbiome interactions in type 1 diabetes and hyperammonaemia was achieved through the integration of microbiome data with clinical information. We found strong relationships in T1D patients between particular microbial taxa and glycaemic control indicators such HbA1c levels. For example, lower glycaemic control was linked to higher levels of Bacteroides, suggesting that these bacteria may play a role in regulating insulin resistance and glucose metabolism.

Blood ammonia levels were strongly correlated with the number of bacteria that produce ammonia in patients with hyperammonaemia. This result emphasises how the gut bacteria directly affect the metabolic dysregulation linked to hyperammonaemia. Additionally, associations with tests of liver function indicated

that the composition of the gut microbiome may possibly represent the degree of liver damage, offering prospective indicators for disease surveillance.

The microbial profiles associated with type 1 diabetes and hyperammonaemia show how complexly the gut flora controls pathways associated with certain diseases. Both the loss of helpful bacteria and the proliferating pro-inflammatory bacteria in T1D point to a role for gut dysbiosis in the initiation and maintenance of the autoimmune response. This fits with the idea of the "leaky gut," according to which systemic immune responses may be triggered by microbial antigens that disseminate via increased intestinal permeability.

Ammonia-producing bacteria and altered routes of amino acid metabolism in hyperammonaemia. The presence of ammonia-producing bacteria and altered amino acid metabolism pathways in hyperammonaemia underscores the microbiome's role in metabolic disturbance. These results imply that reducing ammonia generation by targeting the gut flora may be a successful treatment approach. Probiotics and prebiotics made to encourage the development of bacteria that consume ammonia may be able to control ammonia levels and lessen the severity of sickness. The integration of microbiome analysis into clinical practice provides a guarantee for improved infection detection, prediction, and treatment. Microbial biomarkers, for example, could be developed to distinguish people at high risk for T1D or hyperammonaemia, allowing for early intervention and potentially changing the course of infection. Furthermore, microbiome-based treatments such as probiotics, prebiotics, and microbiome transfers could be improved to restore microbial equilibrium and improve clinical outcomes.

Nonetheless, incorporating microbiome analysis into clinical practice necessitates overcoming a few challenges, including administrative approval, normalization of restorative items, and demonstrating clinical viability through thorough preliminaries. Analysts, clinicians, and industry partners will need to work together to move microbiome-based arrangements from the lab to the bedside.

The gut microbiome is an essential and dynamic component of human health, influencing a wide range of physiological cycles and infection states. Our comparative analysis of gut microbiome data from people with type 1 diabetes and hyperammonaemia emphasizes the importance of understanding microbial commitments to disease instruments. We can uncover new bits of knowledge about the microbiome's role in health and disease by using bioinformatics and coordinating multi-omics data, paving the way for creative, demonstrative, and useful systems.

As the field of microbiome research advances, it has the potential to change our approach to medication, shifting away from side effects for executives and toward all-encompassing, personalized care that considers the multifaceted exchange between microorganisms and their human hosts. Through continued research and collaboration, we can unlock the microbiome's full potential for further development of wellbeing and prosperity.

V. ACKNOWLEDGMENT

I extend my profound gratitude to my mentor Dr. Sonia Chadda and research guide Ms. Nootan Singh, whose unwavering guidance, insightful critiques, and continuous encouragement have been throughout this research journey. I am deeply indebted to the Department of Biotechnology at AMITY UNIVERSITY LUCKNOW CAMPUS, Uttar Pradesh for providing the indispensable infrastructure and resources that facilitated this study on the metagenomics analysis of human gut microbiome.

Additionally, I appreciate the collaborative spirit of the Experiome Biotech PVT. LTD Lucknow, Uttar Pradesh for their provision of critical experimental materials and data analysis tools.

VI. REFERENCES

- 1. Council NR. The New Science of Metagenomics: Revealing the secrets of our Microbial planet. ISBN 978-0-309-10676-4, Washington, DC: The National Academies Press; 2007, 12-31.
- 2. Handelsman J, Rondon MR, Brady SF, Clardy J, Goodman RM. Molecular biological access to the chemistry of unknown soil microbes: a new frontier for natural products. Chem Biol. 1998;5:R245-9
- Laudadio I, Fulci V, Stronati L, Carissimi C. Next-generation metagenomics methodological challenges and opportunities. Omi A J Integr Biol. 2019;23:327–33. <u>https://doi.org/10.1089/omi.2019.0073</u> Mary Ann Liebert Inc, publishers.
- 4. Acinas SG, Sánchez P, Salazar G, Cornejo-Castillo FM, Sebastián M, Logares R, et al. Deep ocean metagenomes provide insight into the metabolic architecture of bathypelagic microbial communities. Commun Biol. 2021;4:604. <u>https://doi.org/10.1038/s42003-021-02112-2</u>.

- 5. Molefe RR, Amoo AE, Babalola OO. Metagenomic insights into the bacterial community structure and functional potentials in the rhizosphere soil of maize plants. J Plant Interact 2021;16:258–69. <u>https://doi.org/10.1080/17429145.2021.1936228</u> Taylor Francis.
- 6. Franzén O, Hu J, Bao X, Itzkowitz SH, Peter I, Bashir A. Improved OTU-picking using long-read 16S rRNA gene amplicon sequencing and generic hierarchical clustering. Microbiome. 2015;3:43. <u>https://doi.org/10.1186/s40168-015-0105-6</u>.
- 7. Edgar RC. Accuracy of taxonomy prediction for 16S rRNA and fungal ITS sequences. PeerJ. 2018;6:e4652.
- Piombo E, Abdelfattah A, Droby S, Wisniewski M, Spadaro D, Schena L. Metagenomics approaches for the detection and surveillance of emerging and recurrent plant pathogens. Microorganisms. 2021;9:188 <u>https://www.mdpi.com/2076-2607/9/1/188</u>.
- Kumar Awasthi M, Ravindran B, Sarsaiya S, Chen H, Wainaina S, Singh E, et al. Metagenomics for taxonomy profiling: tools and approaches. Bioengineered. 2020;11:356– 74. <u>https://doi.org/10.1080/21655979.2020.1736238</u> Taylor & Francis.
- 10. Bush A, Compson ZG, Monk WA, Porter TM, Steeves R, Emilson E, et al. Studying ecosystems with DNA metabarcoding: lessons from biomonitoring of aquatic macroinvertebrates. Front Ecol Evol. 2019;7:434.
- Sharon I, Bercovici S, Pinter RY, Shlomi T. Pathway-based functional analysis of metagenomes. J Comput Biol. 2011;18:495–505. <u>https://doi.org/10.1089/cmb.2010.0260</u> Mary Ann Liebert Inc, publishers.
- Bercovici S, Sharon I, Pinter RY, Shlomi T. Pathway-based functional analysis of Metagenomes BT

 research in computational molecular biology. In: Berger B, editor. Berlin. Heidelberg: Springer, Berlin Heidelberg; 2010. p. 50–64.
- 13. Brown SM, Chen H, Hao Y, Laungani BP, Ali TA, Dong C, et al. MGS-Fast: Metagenomic shotgun data fast annotation using microbial gene catalogs. Gigascience. 2019;8:giz020. https://doi.org/10.1093/gigascience/giz020.
- 14. Nayfach S, Bradley PH, Wyman SK, Laurent TJ, Williams A, Eisen JA, et al. Automated and accurate estimation of gene family abundance from shotgun metagenomes PLOS Comput Biol Public Libr Sci. 2015;11:e1004573. <u>https://doi.org/10.1371/journal.pcbi.1004573</u>.
- 15. Gill SR, Pop M, DeBoy RT, Eckburg PB, Turnbaugh PJ, Samuel BS, et al. Metagenomic analysis of the human Distal Gut Microbiome Sci. 2006;312:1355– 9. https://doi.org/10.1126/science.1124234 Am Association Advancement Sci.
- 16. Lam KN, Cheng J, Engel K, Neufeld JD, Charles TC. Current and future resources for functional metagenomics. Front Microbiol. 2015;6:1196. https://doi.org/10.3389/fmicb.2015.01196, https://www.frontiersin.org/articles/.
- 17. Coughlan LM, Cotter PD, Hill C, Alvarez-Ordóñez A. Biotechnological applications of functional metagenomics in the food and pharmaceutical industries. Front Microbiol. 2015;6:1–22.
- 18. Soni R, Goel R. Triphasic approach to assessment of bacterial population in different soil systems. Ekologija. 2010;6(3-4):99-104.
- 19. Vavourakis CD, Andrei A-S, Mehrshad M, Ghai R, Sorokin DY, Muyzer G. A metagenomics roadmap to the uncultured genome diversity in hypersaline soda lake sediments. Microbiome. 2018;6:168. <u>https://doi.org/10.1186/s40168-018-0548-7</u>.
- 20. Hinsu A, Dumadiya A, Joshi A, Kotadiya R, Andharia K, Koringa P, et al. To culture or not to culture: a snapshot of culture-dependent and culture-independent bacterial diversity from peanut rhizosphere. PeerJ. 2021;9:e12035.