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The Complexity Of Non-Coding Rnas: Implications For Cancer Diagnosis And Therapy

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Abstract:

Non-coding RNAs (ncRNAs) constitute a diverse family of RNA molecules that regulate various cellular processes, with implications spanning from basic molecular biology to clinical applications. This review explores the multifaceted roles of ncRNAs in gene regulation, focusing on their involvement in cancer biology and potential applications as diagnostic and therapeutic targets. NcRNAs encompass a wide array of molecules, including microRNAs (miRNAs), long non-coding RNAs (lncRNAs), and others, each exerting regulatory effects on gene expression through various mechanisms. MiRNAs, for instance, regulate mRNA transcripts via sequence-specific interactions, while lncRNAs modulate gene expression at transcriptional and posttranscriptional levels. In cancer, dysregulated expression of ncRNAs contributes to tumorigenesis, metastasis, and drug resistance, highlighting their potential as biomarkers for diagnosis and prognosis. Liquid biopsies, utilizing circulating tumor-derived ncRNAs, offer less invasive alternatives to traditional tissue biopsies for cancer detection and monitoring. Additionally, advancements in RNA sequencing technologies enable comprehensive profiling of ncRNAs, aiding in the identification of cancer-specific signatures. Therapeutically, ncRNAs hold promise as targets for innovative treatments, such as RNA-based gene silencing therapies. Clinical trials investigating ncRNA-targeted therapies have shown encouraging results in diverse cancer types, although challenges remain in ensuring efficacy and safety. Overall, the burgeoning field of ncRNA research offers insights into the complex regulatory networks governing cancer biology and presents opportunities for the development of novel diagnostic tools and therapeutic interventions.

Keywords: Non-coding RNAs (ncRNAs), Cancer biology, Gene regulation, Diagnostic biomarkers, Therapeutic targets, Liquid biopsies, RNA-based therapies

Introduction to non-coding RNAs

"Frontiers in Non-Coding RNA: Regulation and Therapy"

Non-coding RNAs (ncRNAs) are a family of RNA molecules that play a critical or regulatory role in the process of protein synthesis, in contrast to messenger RNAs, which function as templates for the process. ncRNAs are essential for translation since they are the building blocks of ribosomes (rRNA) or transfer amino acids (tRNA) to the formed peptide. It was just discovered thirty years ago that additional non-coding RNAs (ncRNAs) regulate the expression of proteins or other metabolic processes. [1]. These later ones, which are commonly known as the regulatory ncRNA, attracted attention right away and continue to do so now as fresh information demonstrating their participation in processes like cell proliferation, apoptosis, and differentiation is being shown practically daily. [2]. Cytoplasmic regulatory non-coding RNAs (ncRNAs) are classified into two types based on their nucleotide count: microRNAs (miRNA) and long non-coding RNAs (lncRNAs)[3]. Through the identification of complementary sites at the UTR region of the mRNA, miRNAs interact with mRNA transcripts in a sequencespecific manner that results in transcriptional suppression or the destruction of the mRNA. [4]. LncRNAs are RNA molecules that have a length of more than 200 nucleotides. The primary way that these particles control expression is through their interactions with transcriptional regulatory elements. By competing with the enhancer or creating chromatin loops with target genes, lncRNAs can either positively or adversely impact enhancer activity. Furthermore, it has been demonstrated that lncRNAs interact with the transcript and prevent splicing.[3]. It's interesting to note that lncRNAs have the ability to draw in and bind miRNAs, thereby decreasing their activity. These layered relationships between various RNA types show how intricately these regulatory ncRNAs interact. [3]. ncRNAs influence gene expression, which has a myriad of impacts on different biological mechanisms.[5]. Many diseases in humans, including cancer, metabolic syndrome, heart disease, autoimmune disorders, and infectious diseases, can be brought on by dysregulated expression of non-coding RNAs. Many diseases in humans, including cancer, metabolic syndrome, heart disease, autoimmune disorders, and infectious diseases, can be brought on by dysregulated expression of non-coding RNAs. [6–9]. Because non-coding regulatory transcripts are highly stable and resistant to enzymatic degradation, they can be released into the extracellular space and bloodstream within exosomes. Consequently, ncRNAs can function as prognostic or diagnostic biomarkers.[10]. Lastly, regulatory non-coding RNAs are thought to be therapeutic agents for the management of several illnesses.[11]. A new class of medicines has been developed as a result of ncRNA-based gene silencing, which targets and inhibits genes linked to particular diseases, including cancer. Some of these therapies have received FDA approval. [12]. To create the best possible diagnostic techniques and therapeutic approaches, it is essential to comprehend the mechanisms behind the interactions between regulatory ncRNAs and their targets. Because non-coding RNA is involved in the pathophysiology of many different human diseases, such as inflammatory diseases, genetic abnormalities, and malignancies, it has become the focus of translational research. Because of its special ability to be produced quickly and energetically, RNA is an attractive target for therapeutic development. Based on the length of the transcript, non-coding RNAs (ncRNAs) are classified into two main classes in addition to the traditional functional subtypes such ribosomal RNA (rRNA), small nuclear RNA, small nucleolar RNA, and tRNA. MicroRNA (miRNA), small interfering RNA (siRNA), and PIWIinteracting RNA are examples of tiny ncRNAs (<200 nucleotides), whereas RNAs longer than 200 nucleotides are referred to as long ncRNAs (lncRNAs). [13-14]

Clinical Advances in Short Non-Coding RNAs

Unconjugated sncRNAs

1. Age-related macular degeneration and diabetic macular edema

Age-Related Macular Degeneration with Diabetic Macular Edema The first human clinical trials using siRNA targeting vascular endothelial growth factor (VEGF) were retinal degeneration patients. [16]. The most common cause of significant vision impairment in Americans over 65 is exudative, or "wet," age-related macular degeneration (AMD). [15]. Dry AMD causes drusen to build up on the retina. The pressure this puts on the retinal pigment epithelium triggers an inflammatory reaction that increases VEGF, which in turn causes choroidal neovascularization. [15] The most common cause of blindness in people between the ages of 20 and 74 is diabetic macular edema (DME), which can develop when elevated VEGF increases blood-retinal barrier permeability, causing an excess of fluid to accumulate in the eye and edema. [17] Previous clinical trials have

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demonstrated that VEGF is a useful target for treatments, most often antibodies, to reduce AMD or DME-related visual loss. Anti-VEGF antibodies, such ranibizumab, are the standard of care; however, because they must be administered intravitreally every 4 to 6 weeks, there is a risk of lens damage, intravitreal hemorrhage, endophthalmitis, and retinal tears. [15] A siRNA that targets VEGF mRNA is called bevasiranib (also called Cand5) [17]. The overexpression of VEGF is caused by mRNA stability rather than enhanced translation, which makes siRNA-based therapies superior to antibody-based therapies. Additionally, the use of siRNA theoretically permits downregulation rather than inhibiting the activity. [15] Because the VEGF mRNA that is already there is not completely destroyed, bevasiranib does exhibit a clear anti-angiogenic impact, which takes around 6 weeks to develop from the commencement of treatment. In this instance, anti-VEGF antibody combo therapy may be the most beneficial course of action. [15] NCT00557791 was a phase 3 clinical trial that was intended to investigate the advantages of this combination medication, however it was never initiated. Numerous investigations have revealed that bevasiranib primarily acts on the cell surface toll-like receptor 3 (TLR3) through RNA-mediated activation, which decreases CNV through intracellular signaling, rather than by inducing an RNAi response. [16] Since these siRNAs were not created with cell penetration in mind, it's possible that fewer of them than expected reach their intended target. [16] A phase 3 clinical trial for bevasiranib was stopped in 2009 due to early results indicating a very low chance of achieving the primary aim (NCT00499590).

Respiratory Syncytial Infection

The primary cause of hospitalization for infants in the United States is respiratory syncytial virus (RSV), in part due to the lack of a vaccination and the scarcity of treatment options for this infection.[18]. The most frequent community-acquired respiratory virus in lung transplant patients is RSV infection, which is linked to bronchiolitis obliterans syndrome, a major barrier to patient and graft survival [42]. Alnylam Pharmaceuticals created the siRNA ALN-RSV01, which targets the mRNA encoding the nucleocapsid protein, which is essential for RSV replication [18,43]. Delivery without a carrier is effective, as it can be given directly to the mucosa and destroyed by the nucleases if it enters the systemic circulation, as is the case with lung-targeted siRNAs. [21] Intranasal injection of 150 mg dosages given once or five times per day was shown to be safe and well-tolerated in safety and tolerability studies involving 101 healthy people [20]. Following an experimental RSV challenge in 88 healthy adults, 71.4% of the placebo group and 44.2% of the ALN-RSV01 group contracted the virus [18]. In Phase 2a trials, ALN-RSV01 was demonstrated to lower the risk of new or progressive bronchiolitis obliterans syndrome (BOS) when combined with standard of care in transplant patients who were naturally infected with RSV. It did not, however, advance to a phase 3 study and did not reach the primary goal of reduced day 180 BOS [19].

Pachyonychia Congenita

The dominant hereditary disorder known as pachyonychia congenita (PC) is characterized by thicker nails, keratoderma, leukokeratosis, and excruciating blisters that are mostly on the soles of the feet [42]. Without the assistance of an ambulatory device, more than 50% of patients are unable to walk [22]. Oral retinoids, topical keratolytics, and mechanical callus removal are the only effective symptom control options available for PC at this time [22]. Mutations in keratins K6a, K6b, K16, or K17 cause this syndrome. The most frequently altered gene, K6a mRNA, is the target of the siRNA treatment TD101 [42, 22]. The efficacy of TD101 intradermal injection in suppressing mutant K6a expression was confirmed by measuring in vivo mRNA levels using quantitative reverse transcription PCR (qRT-PCR). The same levels of mutant and wild-type K6a were expressed by PC-10 cells and patient callus samples that were obtained. Nevertheless, the administration of TD101 resulted in a 98% reduction in the expression of mutant K6a [42].

Hepatitis C

The most prevalent hepatic miRNA, miR-122, facilitates the spread of the hepatitis C virus (HCV). miR-122 binds to the 50 end of HCV RNA, shielding it from nuclease assault and hiding an RNA motif that could trigger an innate immune reaction [23]. Cirrhosis and ultimately hepatocellular cancer can result with chronic HCV [24]. Currently undergoing clinical trials is miravirsen, an anti-miR-122 ASO made of locked nucleic acid (LNA) ribonucleotides that hybridize to mature miR-122 and prevent its association with HCV RNA [28]. The second

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oxygen molecule in LNAs is connected to the ribonucleotide's 40 carbons. This alteration can raise target affinity while shielding the oligonucleotide from nuclease degradation [24, 25].

Clinical investigations have shown that treatment with miravirsen reduces the viral load in individuals with chronic HCV in a dose-dependent manner while having no discernible impact on other miRNAs' plasma levels [23]. One week after the first dose in the experimental group, a placebo-controlled study of five weekly doses of miravirsen decreased plasma levels of miR-122 from $3.9 \times 103/4$ to $3.1 \times 101/4$ µL. In the highest-dose group, these values were sustained for the duration of the study. [23] By contrast, after one week of treatment, the mean plasma levels in the placebo group were $1.1 \times 104/4$ µL, compared to $1.3 \times 104/4$ µL at baseline. Following treatment, all dosed patients showed improvement, and some even had undetectable levels of miR-122. The HCV viral load did not correlate with the drop in miR-122 plasma levels, despite the fact that the viral load did in dosed patients. It is believed that a C3U nucleotide alteration in the 50 UTR region of the HCV RNA makes this process miR-122-independent and hence resistant to miravirsen in a large number of patients who experienced virological recurrence after taking the medication [26,27]. It has been suggested that miR-122 may also function as a tumor suppressor [35], which has sparked worries that receiving anti-miR-122 therapy may elevate the risk of hepatocellular carcinoma. In preclinical research, mice given miravirsen for five weeks did not grow tumors, nevertheless. Even yet, considering that hepatocellular carcinoma does arise in mir-122-knockout animal models [26, 35], this worry calls for additional safety research to assess the danger [28].

5. Acute Kidney Injury

The complex illness known as acute kidney injury (AKI) is marked by an abrupt drop in glomerular filtration rate, which is then followed by an increase in blood creatinine concentration or oliguria. AKI typically happens in the context of a recent or ongoing medical condition. About 20% of hospitalized patients are affected by it. The pooled incidence rate of AKI in clinical trials was 21.6%, and 10% of patients needed kidney replacement therapy [29, 30]. A p53-targeting siRNA called QPI-1002 (Teprasiran, Quark Pharmaceuticals) is used to prevent AKI and post-kidney replacement delayed graft function [31]. 10 mg/kg of QPI-1002 decreased the incidence, severity, and length of AKI following heart surgery in high-risk patients in a phase 2 clinical trial [31]. Nevertheless, a phase 3 clinical trial (NCT03510897) was abruptly stopped since the patients' results did not reach the efficacy objectives at day 90.

6. Alport's Disease

A multifaceted miRNA, miR-21 is involved in inflammation, fibrosis, immunological response, and carcinogenesis [41,35,32,33]. Mutations in the genes encoding several α chains of collagen 4 result in the hereditary condition known as Alport syndrome. The kidney's and other organs' capillary membranes are jeopardized by altered collagen 4 function. Alport syndrome patients and genetic mice models exhibit elevated expression of miR-21 [34,36]. Subcutaneous administration of 25 mg/kg antimiR-21 ASO twice a week increased animal survival by 46% in the Col4a3–/– mouse model [34]. The development of glomerular crescents, periglomerular fibrosis, and glomerulosclerosis—all linked to the advancement of Alport syndrome—were markedly postponed by the anti-miR-21 ASO treatment. [34] Mechanistically, anti-miR-21 ASO therapy prolongs kidney function by preventing TGF- β -induced fibrosis and inflammation and by safeguarding PPAR α /retinoid X receptor (PPAR α /RXR)-dependent mitochondrial activity. Anti-miR-21 ASO (RG-021, now known as lademirsen) was administered subcutaneously to individuals with Alport syndrome in phase 1 clinical trials at a dose of 1.5 mg/kg, either as a single dose or as four doses spaced 14 days apart (NCT03373786). Following a well-tolerated course of treatment, individuals with Alport syndrome are currently being actively recruited for a phase 2 clinical trial to assess the therapeutic efficacy of lademirsen in maintaining kidney function (NCT02855268).

7. cardiovascular disease

Targeting miR-92a-3p, MRG-110 is an LNA-modified ASO that is used to treat wound healing and cardiovascular disease [25]. Inhibiting miR-920 has been shown to improve wound healing, circulation following hind limb ischemia, and vascularization following heart attacks. It also mitigates the negative effects of miR-920's antiangiogenic effects on wound healing, which are partly due to the downregulation of pro-angiogenic

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integrin alpha 5 [25, 37]. Whole blood miR-92a-3p is reduced in a dose-dependent manner by MRG-110 therapy. In experimental models of acute and chronic excision wounds in pigs and db/db mice, it also enhances the development of granulation tissue and stimulates angiogenesis [37]. Significant clinical potential was indicated by the fact that these effects were higher in the MRG-110 group compared to the positive control groups treated with rhVEGF-165 and rhPDGF-BB. There were no noteworthy safety issues. In human trials, the treatment's considerable effectiveness was observed at half-maximum dosages of 0.27 to 0.31 mg/kg [25]. After 24-72 hours of therapy, there was over 95% inhibition in the high dose groups, and this inhibition persisted for two weeks.

8. Leukemias and Lymphomas

As a treatment for many hematologic malignancies, such as cutaneous T-cell lymphoma (CTCL), diffuse large B-cell lymphoma (DLBCL), and chronic lymphocytic leukemia (CLL), MRG-106 (cobomarsen), an LNAmodified ASO, targets miR-155. Mycosis fungoides (MF), the most prevalent subtype of CTCL, has a significant etiology, which is supported by functional investigations and clinical data [38]. Cobomarsen's formulation promoted CD4+ T-cell and MF cell uptake [38]. Treatment with cobomarsen increased the expression of BACH1, PICALM, and JARID2, which are direct targets of miR-1550, and disrupted the pro-survival function of miR-155. [38] JARID2 (jumonji and AT-rich interaction domain containing 2) is a negative regulator of leukemia cell proliferation, PICALM (phosphatidylinositol binding clathrin assembly protein) is an endocytosis adaptor, and BACH1 (BTB and CNC homology 1, basic leucine zipper transcription factor 1) is a mediator of the oxidative stress response.[38] In patients with hematological malignancies, a phase 1 clinical trial (NCT02580552) showed that cobomarsen was safe and had minimal toxicity. In order to compare the safety and effectiveness of cobomarsen medication to vorinostat, a histone deacetylase (HDAC) inhibitor, in patients with CTCL of the MF subtype, a phase 2 clinical trial (NCT03713320) was started in 2018. One of the attractive things about cobomarsen treatment is that it can be given once a week instead of vorinostat's daily dosage; nevertheless, cobomarsen is given intravenously, whereas vorinostat is taken orally. Despite recruiting 37 patients, this research trial was stopped for business reasons without any particular concerns about the efficacy of cobomarsen. [39] Because there were few eligible participants, an expected crossover phase 2 clinical trial (NCT03837457) had to be canceled. The further clinical evaluation of cobomarsen is supported by genetic investigations in Mir-155-knockout mice models, successful treatment with anti-miR-155 ASO or comparable inhibitors in in vivo animal models. [41], and an unusual response in a single patient diagnosed with an aggressive subtype of DLBCL [40].

Non-Coding RNAs: Cancer Biomarkers and Diagnosis

Because of their distinct expression profiles, high relative stability, and ease of PCR characterisation, non-coding RNAs (ncRNAs) are a great class of prospective biomarkers [43]. Thus, in the last ten years, a number of clinical trials have been carried out to find ncRNA biomarkers in cancer patients in order to create screening instruments. Since this may affect patient cohort composition and specimen selection, it is crucial that the intended application of the proposed biomarker(s) be clearly specified as predictive, prognostic, or diagnostic. [44,45] Early studies of identifying tumor ncRNAs implemented the strategy of comparative profiling between both healthy and malignant tissues [46]. Tissue biopsies have a long history of clinical use and are a useful tool, but they are invasive and impractical for patients who are fragile or have inaccessible malignancies [47, 48]. Moreover, tissue biopsies yield information that is reliant on both space and time, which means that it may give an erroneous picture of tumor heterogeneity and ongoing tumor processes such drug resistance [47-50]. Specialized signals of non-coding ncRNAs produced from cancer have been detected in bodily fluids such as blood, saliva, and urine [51–53], prompting investigators to investigate the feasibility of liquid biopsies [48]. Liquid biopsies are less invasive than tissue biopsies, making them ideal for therapy monitoring [202] and screening [54]. Nevertheless, the frequency of circulating tumor cells (CTCs) is comparatively low, and while free-traveling ncRNAs are vulnerable to destruction by circulating RNAses, ncRNAs can travel through physiological fluids without the assistance of cells [45,49]. Examining ncRNAs contained in extracellular vesicles (EVs), which are secreted by tumor cells, is an other strategy [55] Studies have revealed that tumor cells secrete more vesicles than normal cells do, which may contribute to the development of pre-metastatic niches and the advancement of cancer. [45] It is imperative to ascertain if the observed changes in ncRNA levels are

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obscured by variations in vesicle concentrations between individuals with cancer and healthy individuals. The whole genome expression profiles of patient and control samples can be compared to identify dysregulated noncoding RNAs (ncRNAs) since high-throughput sequencing technologies, such as next-generation sequencing, have been available [58,44]. Notably, the heterogeneity of human malignancies and the widespread expression of most ncRNAs make it unlikely that a single biomarker will be adequate for disease characterisation [56,57]. Instead, in order to achieve high sensitivity and specificity, bio classifier systems made up of a panel of biomarkers might be required. [56] Large-scale ncRNA expression profiles have previously shown promise in the classification of poorly differentiated tumors, and they may also add to our knowledge of the dynamics underlying cancers [59, 45]. Making sure these bio classifiers are repeatable, however, is a significant hurdle. As a result, standardizing bioinformatic analyses, normalization techniques, and protocols for RNA extraction and sample processing is crucial [44,53,57]. PCA3 is the only ncRNA to date to be approved as a biomarker by the FDA, despite the identification of many potential ncRNA biomarker candidates [68]. Prostate cancer has a distinct upregulation of the lncRNA PCA3 [50]. The upregulation of PCA3 in prostate cancer patient urine has been reported by Hessels et al. [61], which has led to the development of non-invasive PCA3 urine tests for the clinical identification of early prostate cancer [62]. Currently, this test is utilized in combination with other recognized assays (e.g., TMPRSS2:ERG urine test, PSA blood test) [68,63,64].

ncRNAs as Lung Cancer Biomarkers

Lung cancer has one of the lowest five-year survival rates of any malignancy (19%) [69]. There are few curative treatments available for the more than half of patients who arrive with advanced-stage disease [65,66]. As a result, a significant percentage of these patients receive no treatment [67]. Crucially, screening for lung cancer can raise high-risk individuals' survival rates. Eighty percent of lung cancers detected in the early stages are detected by screening programs; in the absence of screening, a staggering seventy percent of patients receive a late-stage diagnosis [70]. A low-dose CT scan is currently the gold standard for screening for lung cancer [71].

ncRNAs: Cancer Biology

Calin and Croce's seminal study [22,72] from 2002 revealed a connection between dysregulated miR-15a and miR-16-1 and chronic lymphocytic leukemia (CLL). Since then, numerous ncRNAs have been connected to well-known cancer pathways [79, 80]. Even if there aren't many well-studied ncRNAs included, it's important to remember that focusing on a single biomolecule or route in isolation oversimplifies the biological reality of multiple cancer pathways interacting with one another [76]. Additionally, a single ncRNA can interact with a wide range of proteins, mRNAs, DNA, and other ncRNAs [76,82,83] and be involved in the regulation of several biological processes [81].

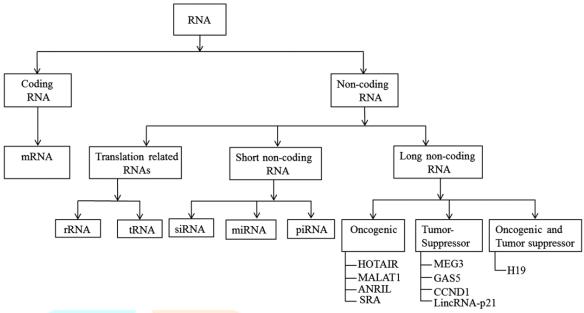
Overview of ncRNAs in Cancer

The transcriptome of cancer cells and tissues may be examined thanks to developments in RNA sequencing (RNA-seq) technologies [74]. This approach makes it possible to determine the frequency and sequences of dysregulated non-coding RNAs in malignancies [74, 75]. miRNAs have been explored the most in relation to the roles played by ncRNAs in human cancers [84, 85]. Numerous in vitro and in vivo research employ tactics of over- and under-expressing the miRNA(s) of interest in order to unveil the roles of miRNAs related to cancer. Examining the generated biological activity using a variety of functional tests comes next [73,78]. The function of miRNAs can also be ascertained by elucidating their mRNA targets using high-throughput sequencing or in silico methods (e.g., Targets can, miRanda) [73,86]. Remarkably, recent research has demonstrated that secreted miRNAs can function as ligands to initiate premetastatic inflammatory responses in the tumor microenvironment in addition to causing RNAi [87,88]. Less is known about piRNAs' roles in cancer. Although more recent research has looked at the PIWI/piRNA relationship in malignancies, the majority of studies to date have focused on the PIWI clade of Argonaut proteins independently of piRNAs [89, 90]. These complexes are generally overexpressed in malignancies, and this overexpression has been connected to aggressive cancer characteristics.[77] Numerous well-established lncRNAs (e.g., HOTAIR, H19, MEG3, MALAT1) have been associated with malignancies. They play a variety of roles in the development of malignancies, particularly in the areas of drug response, angiogenesis, metastasis, cell proliferation, and post-transcriptional gene regulation. The effect of non-coding RNAs (ncRNAs) can be broadly classified as either tumorigenic or tumor suppressive based on the understanding gained from functional investigations. However, certain ncRNAs may exhibit both activities depending on the context [91,92].

Diverse Functions of ncRNAs

Non-translated or non-coding RNA (ncRNA) molecules are transcripts of genomic sequences that are not intended for translation. [93] The human genome encodes a large number of non-coding RNAs. The majority of these non-coding RNAs have been extensively linked to the regulation of cellular homeostasis. [94] Certain ncRNAs have a direct bearing on modifications and/or alterations in cells' epigenetic makeup. As shown in Figure 1, total cellular RNAs are categorized according to their functions. Long non-coding RNAs (lncRNAs) and small nuclear RNAs (snoRNAs), microRNAs, small interfering RNAs (siRNAs), PIWI-interacting RNAs (piRNAs), transfer RNA (tRNA), ribosomal RNA (rRNA), and other functionally significant RNAs are produced from a subset of ncRNAs. X inactivation specific transcript (exist) and HOX antisense intergenic RNA (HOTAIR) are two extensively researched long noncoding RNAs. [103] Although the entire number of noncoding RNAs (ncRNAs) encoded in the human genome is unknown, thousands of ncRNAs with potential functions may exist, according to recent transcriptomic and bioinformatic research. It's probable that a large number of the recently discovered ncRNAs are non-functional because their functions haven't been confirmed. Examples of ncRNAs' roles and functions are still being discovered, though. ncRNAs are classified as either long non-coding RNA ($\ln cRNAs > 200$ nt) or short non-coding RNA (sncRNAs < 30 nt), depending on the length of RNA generated post-transcriptionally. Longer than 200 nt, non-protein coding transcripts are referred to as lncRNAs. Practical factors, such as the separation of RNAs in standard experimental methods, are the reason for this limit. Furthermore, Table 1 shows that this cutoff separates lncRNAs from small regulatory RNAs including siRNAs, piRNAs, miRNAs, and snoRNAs. Short noncoding RNAs, or miRNAs, range in length from 18 to 24 nt and play a role in skin fibrosis.[95] LncRNAs have a wide range of functions, including roles in telomere biology, higher order chromosomal dynamics, and subcellular structural organization. [96,97] T-UCR, a novel subclass of ncRNAs, is produced from an ultra-conserved region.[98] T-UCRs are a subset of DNA segments larger than 200 bp that are fully conserved across species (genomes of rats, mice, and humans).[100, 99] Furthermore, the type of the genes involved in oncogenesis and/or tumour suppression determines subsequent divisions of ncRNAs. Steroid receptor RNA activator (SRA), for instance, is an oncogenic noncoding RNA that, in response to insulin, increases adipogenesis and suppresses the expression of inflammatory genes linked to adipocytes.[100] Maternally expressed gene 3 (MEG3) is a ncRNA encoding gene that is found on human chromosome 14q32.3 on the DLK1-MEG3 locus. When this gene is inactivated, the brain develops micro vessels and genes that promote angiogenesis are expressed. [101] By competing with the DNAglucocorticoid response element (DNA-GRE) at the glucocorticoid receptor's DNA binding domain, GAS5 ncRNA functions as a ruse and modifies growth arrest under hunger. [102]

NON-CODING RNAS: POWER AND PROMISES



Feature Prospective

It provides an in-depth exploration of the multifaceted roles of non-coding RNAs (ncRNAs) in gene regulation, with a focus on their involvement in cancer biology and potential applications as diagnostic and therapeutic targets. It discusses how ncRNAs, including microRNAs (miRNAs) and long non-coding RNAs (lncRNAs), exert regulatory effects on gene expression through various mechanisms. In cancer, dysregulated expression of ncRNAs contributes to tumorigenesis, metastasis, and drug resistance, making them potential biomarkers for diagnosis and prognosis. Liquid biopsies, which utilize circulating tumor-derived ncRNAs, offer less invasive alternatives for cancer detection and monitoring. Additionally, advancements in RNA sequencing technologies enable comprehensive profiling of ncRNAs, aiding in the identification of cancer-specific signatures. Therapeutically, ncRNAs hold promise as targets for innovative treatments, such as RNA-based gene silencing therapies, with clinical trials showing encouraging results across diverse cancer types. However, challenges remain in ensuring efficacy and safety in ncRNA-targeted therapies. Overall, the burgeoning field of ncRNA research offers insights into the complex regulatory networks governing cancer biology and presents opportunities for the development of novel diagnostic tools and therapeutic interventions.

Conclusion

Non-coding RNAs (ncRNAs) represent a burgeoning field of research with profound implications for cancer biology and clinical practice. These molecules, including microRNAs (miRNAs) and long non-coding RNAs (lncRNAs), exert intricate regulatory effects on gene expression, influencing key processes involved in tumorigenesis and cancer progression. Their dysregulation in cancer underscores their potential as diagnostic biomarkers and therapeutic targets. Advancements in RNA sequencing technologies have facilitated the identification of cancer-specific ncRNA signatures, offering promise for improved cancer diagnosis and prognosis. Liquid biopsies, harnessing circulating tumor-derived ncRNAs, provide less invasive alternatives to traditional tissue biopsies for monitoring disease progression and treatment response. Moreover, therapeutic targeting of dysregulated ncRNAs holds great potential for innovative cancer treatments. RNA-based gene silencing therapies, including those targeting miRNAs and lncRNAs, have shown encouraging results in preclinical and clinical studies, highlighting their promise for personalized medicine approaches. Overall, the study of ncRNAs in cancer represents a frontier in cancer research, offering insights into the complex molecular mechanisms driving malignancy and paving the way for the development of novel diagnostic tools and therapeutic interventions aimed at improving patient outcomes in the fight against cancer.

References

- 1. Patil, V.S.; Zhou, R.; Rana, T.M. Gene regulation by non-coding RNAs. *Crit. Rev. Biochem. Mol. Biol.* **2014**, *49*, 16–32. [CrossRef] [PubMed]
- 2. Ambros, V. The functions of animal microRNAs. *Nature* 2004, 431, 350–355. [CrossRef] [PubMed]
- 3. Park, Y.B.; Kim, J.M. Identification of long non-coding RNA-mRNA interactions and genome-wide lncRNA annotation in animal transcriptome profiling. *J. Anim. Sci. Technol.* **2023**, *65*, 293–310. [PubMed]
- 4. Al-Mahayni, S.; Ali, M.; Khan, M.; Jamsheer, F.; Moin, A.S.M.; Butler, A.E. Glycemia-Induced miRNA Changes: A Review. *Int. J. Mol. Sci.* 2023, *24*, 7488. [CrossRef] [PubMed]
- 5. O'Brien, J.; Hayder, H.; Zayed, Y.; Peng, C. Overview of MicroRNA Biogenesis, Mechanisms of Actions, and Circulation. *Front. Endocrinol.* **2018**, *9*, 402. [CrossRef] [PubMed]
- Dopytalska, K.; Czaplicka, A.; Szymanska, E.; Walecka, I. The Essential Role of microRNAs in Inflammatory and Autoimmune Skin Diseases-A Review. Int. J. Mol. Sci. 2023, 24, 9130. [CrossRef] [PubMed]
- 7. SiouNing, A.S.; Seong, T.S.; Kondo, H.; Bhassu, S. MicroRNA Regulation in Infectious Diseases and Its Potential as a Biosensor in Future Aquaculture Industry: A Review. *Molecules* 2023, 28, 4357. [CrossRef] [PubMed]
- 8. Otsuka, K.; Nishiyama, H.; Kuriki, D.; Kawada, N.; Ochiya, T. Connecting the dots in the associations between diet, obesity, cancer, and microRNAs. *Semin. Cancer Biol.* **2023**, *93*, 52–69. [CrossRef] [PubMed]
- 9. Arunima, A.; van Schaik, E.J.; Samuel, J.E. The emerging roles of long non-coding RNA in host immune response and intracellular bacterial infections. *Front. Cell Infect. Microbiol.* **2023**, *13*, 1160198. [CrossRef] [PubMed]
- Bravo-Vazquez, L.A.; Frias-Reid, N.; Ramos-Delgado, A.G.; Osorio-Perez, S.M.; Zlotnik-Chavez, H.R.; Pathak, S.; Banerjee, A.; Bandyopadhyay, A.; Duttaroy, A.K.; Paul, S. MicroRNAs and long non-coding RNAs in pancreatic cancer: From epigenetics to potential clinical applications. *Transl. Oncol.* 2023, 27, 101579. [CrossRef] [PubMed]
- Bajan, S.; Hutvagner, G. RNA-Based Therapeutics: From Antisense Oligonucleotides to miRNAs. *Cells* 2020, 9, 137. [CrossRef] [PubMed]
- 12. Kara, G.; Calin, G.A.; Ozpolat, B. RNAi-based therapeutics and tumor targeted delivery in cancer. *Adv. Drug Deliv. Rev.* **2022**, *182*, 114113. [CrossRef] [PubMed]
- Gomes, A. Q., Nolasco, S., and Soares, H. (2013). Non-coding RNAs: multitasking molecules in the cell. Int. J. Mol. Sci. 14, 16010–16039. doi: 10.3390/ijms140816010
- Adams, B. D., Parsons, C., Walker, L., Zhang, W. C., and Slack, F. J.(2017). Targeting noncoding RNAs in disease. J. Clin. Invest. 127, 761–771.doi: 10.1172/JCI84424

15. Garba, A.O.; Mousa, S.A. Bevasiranib for the treatment of wet, age-related macular degeneration. *Ophthalmol. Eye Dis.* **2010**, *2*, 75–83. [CrossRef]

16. Ambati, J. Age-related macular degeneration and the other double helix the cogan lecture. *Investig. Ophthalmol. Vis. Sci.* **2011**, *52*, 2166–2169. [CrossRef]

17. du Castel, C. Safety and Effificacy Study of Small Interfering RNA Molecule (Cand5) to Treat Diabetic Macular Edema. Available online: https://ClinicalTrials.gov/show/NCT00306904 (accessed on 5 January 2022)

18. DeVincenzo, J.; Lambkin-Williams, R.; Wilkinson, T.; Cehelsky, J.; Nochur, S.; Walsh, E.; Meyers, R.; Gollob, J.; Vaishnaw, A. A randomized, double-blind, placebo-controlled study of an RNAi-based therapy directed against respiratory syncytial virus. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 8800–8805. [CrossRef] [PubMed]*Cancers* **2022**, *14*, 1588

19. Gottlieb, J.; Zamora, M.R.; Hodges, T.; Musk, A.W.; Sommerwerk, U.; Dilling, D.; Arcasoy, S.; DeVincenzo, J.; Karsten, V.; Shah, S.; et al. ALN-RSV01 for prevention of bronchiolitis obliterans syndrome after respiratory syncytial virus infection in lung transplant recipients. *J. Heart Lung Transplant. Off. Publ. Int. Soc. Heart Transplant.* **2016**, *35*, 213–221. [CrossRef] [PubMed]

20. DeVincenzo, J.; Cehelsky, J.E.; Alvarez, R.; Elbashir, S.; Harborth, J.; Toudjarska, I.; Nechev, L.; Murugaiah, V.; Van Vliet, A.; Vaishnaw, A.K.; et al. Evaluation of the safety, tolerability and pharmacokinetics of ALN-RSV01, a novel RNAi antiviral

therapeutic directed against respiratory syncytial virus (RSV). Antivir. Res. 2008, 77, 225–231. [CrossRef]

21. Astor, T.L. RNA interference, RSV, and lung transplantation: A promising future for siRNA therapeutics. *Am. J. Respir. Crit. Care Med.* **2011**, *183*, 427–428. [CrossRef]

22. Leachman, S.A. Study of TD101, a Small Interfering RNA (siRNA) Designed for Treatment of Pachyonychia Congenita. Available online: https://ClinicalTrials.gov/show/NCT00716014 (accessed on 6 January 2022).

23. van der Ree, M.H.; van der Meer, A.J.; van Nuenen, A.C.; de Bruijne, J.; Ottosen, S.; Janssen, H.L.; Kootstra, N.A.; Reesink, H.W.

Miravirsen dosing in chronic hepatitis C patients results in decreased microRNA-122 levels without affecting other microRNAs

in plasma. Aliment. Pharmacol. Ther. 2016, 43, 102–113. [CrossRef]

24. Gebert, L.F.; Rebhan, M.A.; Crivelli, S.E.; Denzler, R.; Stoffel, M.; Hall, J. Miravirsen (SPC3649) can inhibit the biogenesis of miR-122. *Nucleic Acids Res.* **2014**, *42*, 609–621. [CrossRef] [PubMed]

25. Abplanalp, W.T.; Fischer, A.; John, D.; Zeiher, A.M.; Gosgnach, W.; Darville, H.; Montgomery, R.; Pestano, L.; Allée, G.; Paty, I.;

et al. Effificiency and Target Derepression of Anti-miR-92a: Results of a First in Human Study. *Nucleic Acid Ther.* **2020**, *30*, 335–345. [CrossRef] [PubMed]

26. Li, Y.P.; Van Pham, L.; Uzcategui, N.; Bukh, J. Functional analysis of microRNA-122 binding sequences of hepatitis C virus and identifification of variants with high resistance against a specifific antagomir. *J. Gen. Virol.* **2016**, *97*, 1381–1394. [CrossRef] [PubMed]

27. Ottosen, S.; Parsley, T.B.; Yang, L.; Zeh, K.; van Doorn, L.J.; van der Veer, E.; Raney, A.K.; Hodges, M.R.; Patick, A.K. In vitro

antiviral activity and preclinical and clinical resistance profifile of miravirsen, a novel anti-hepatitis C virus therapeutic targeting

the human factor miR-122. Antimicrob. Agents Chemother. 2015, 59, 599-608. [CrossRef] [PubMed]

28.Lindow, M.; Kauppinen, S. Discovering the fifirst microRNA-targeted drug. *J. Cell Biol.* **2012**, *199*, 407–412. [CrossRef] [PubMed]

29. Wonnacott, A.; Meran, S.; Amphlett, B.; Talabani, B.; Phillips, A. Epidemiology and outcomes in community-acquired versus hospital-acquired AKI. *Clin. J. Am. Soc. Nephrol.* **2014**, *9*, 1007–1014. [CrossRef]

30. James, M.T.; Hemmelgarn, B.R.; Wiebe, N.; Pannu, N.; Manns, B.J.; Klarenbach, S.W.; Tonelli, M.; Alberta Kidney Disease Network. Glomerular fifiltration rate, proteinuria, and the incidence and consequences of acute kidney injury: A cohort study. *Lancet* **2010**, *376*, 2096–2103. [CrossRef]

31. Thielmann, M.; Corteville, D.; Szabo, G.; Swaminathan, M.; Lamy, A.; Lehner, L.J.; Brown, C.D.; Noiseux, N.; Atta, M.G.; Squiers, E.C.; et al. Teprasiran, a Small Interfering RNA, for the Prevention of Acute Kidney Injury in High-Risk Patients Undergoing Cardiac Surgery: A Randomized Clinical Study. *Circulation* **2021**, *144*, 1133–1144. [CrossRef]

32. Bautista-Sanchez, D.; Arriaga-Canon, C.; Pedroza-Torres, A.; De La Rosa-Velazquez, I.A.; Gonzalez-Barrios, R.; ContrerasEspinosa, L.; Montiel-Manriquez, R.; Castro-Hernandez, C.; Fragoso-Ontiveros, V.; Alvarez-Gomez, R.M.; et al. The Promising Role of miR-21 as a Cancer Biomarker and Its Importance in RNA-Based Therapeutics. *Mol. Ther. Nucleic Acids* **2020**, *20*, 409–420. [CrossRef]

33. Sheedy, F.J. Turning 21: Induction of miR-21 as a Key Switch in the Inflflammatory Response. *Front. Immunol.* 2015, 6, 19. [CrossRef]

34. Gomez, I.G.; MacKenna, D.A.; Johnson, B.G.; Kaimal, V.; Roach, A.M.; Ren, S.; Nakagawa, N.; Xin, C.; Newitt, R.; Pandya, S.; et al. Anti—microRNA-21 oligonucleotides prevent Alport nephropathy progression by stimulating metabolic pathways. *J. Clin. Investig.* **2015**, *125*, 141–156. [CrossRef] [PubMed]

35. Sempere, L.F.; Powell, K.; Rana, J.; Brock, A.A.; Schmittgen, T.D. Role of non-coding RNAs in tumor progression and metastasis in pancreatic cancer. *Cancer Metastasis Rev.* **2021**, *40*, 761–776. [CrossRef]

36. Guo, J.; Song, W.; Boulanger, J.; Xu, E.Y.; Wang, F.; Zhang, Y.; He, Q.; Wang, S.; Yang, L.; Pryce, C.; et al. Dysregulated Expression of microRNA-21 and Disease-Related Genes in Human Patients and in a Mouse Model of Alport Syndrome. *Hum. Gene Ther.* **2019**, *30*, 865–881. [CrossRef] [PubMed]*Cancers* **2022**, *14*, 1588

37. Gallant-Behm, C.L.; Piper, J.; Dickinson, B.A.; Dalby, C.M.; Pestano, L.A.; Jackson, A.L. A synthetic microRNA-92a inhibitor (MRG-110) accelerates angiogenesis and wound healing in diabetic and nondiabetic wounds. *Wound Repair Regen.* **2018**, *26*, 311–323. [CrossRef] [PubMed]

38. Seto, A.G.; Beatty, X.; Lynch, J.M.; Hermreck, M.; Tetzlaff, M.; Duvic, M.; Jackson, A.L. Cobomarsen, an oligonucleotide inhibitor of miR-155, co-ordinately regulates multiple survival pathways to reduce cellular proliferation and survival in cutaneous T-cell lymphoma. *Br. J. Haematol.* **2018**, *183*, 428–444. [CrossRef]

39. miRagen. Miragen Announces Internal Review of Preliminary Topline Data for the Phase 2 Solar Clinical Trial of Cobomarsen in Patients with Cutaneous T-Cell Lymphoma (CTCL). Available online: http://investors.miragen.com/press-releases/pressrelease/2020/miRagen-Announces-Internal-Review-of-Preliminary-Topline-Data-for-the-Phase-2-SOLAR-Clinical-Trial-ofCobomarsen-in-Patients-with-Cutaneous-T-Cell-Lymphoma-CTCL/default.aspx (accessed on 14 March 2022).

40. Anastasiadou, E.; Seto, A.; Beatty, X.; Hermreck, M.; Gilles, M.E.; Stroopinsky, D.; Pinter-Brown, L.C.; Pestano, L.; Marchese, C.; Avigan, D.; et al. Cobomarsen, an oligonucleotide inhibitor of miR-155, slows DLBCL tumor cell growth in vitro and in vivo. *Clin. Cancer Res.* **2020**, *27*, 1139–1149. [CrossRef]

41. Sempere, L.F.; Azmi, A.S.; Moore, A. microRNA-based diagnostic and therapeutic applications in cancer medicine. *Wiley Interdiscip. Rev. RNA* **2021**, *12*, e1662. [CrossRef]

42. Hickerson, R.P.; Leachman, S.A.; Pho, L.N.; Gonzalez-Gonzalez, E.; Smith, F.J.; McLean, W.I.; Contag, C.H.; Leake, D.; Milstone, L.M.; Kaspar, R.L. Development of quantitative molecular clinical end points for siRNA clinical trials. *J. Investig. Dermatol.* **2011**, *131*, 1029–1036. [CrossRef]

43. Kashi, K.; Henderson, L.; Bonetti, A.; Carninci, P. Discovery and functional analysis of lncRNAs: Methodologies to investigate an uncharacterized transcriptome. *Biochim. Biophys. Acta* **2016**, *1859*, 3–15. [CrossRef] [PubMed]

44. Goossens, N.; Nakagawa, S.; Sun, X.; Hoshida, Y. Cancer biomarker discovery and validation. *Transl Cancer Res.* **2015**, *4*, 256–269. [CrossRef]

45. Qi, P.; Du, X. The long non-coding RNAs, a new cancer diagnostic and therapeutic gold mine. *Mod. Pathol.* **2013**, *26*, 155–165. [CrossRef]

46. Calin, G.A.; Croce, C.M. MicroRNA signatures in human cancers. *Nat. Rev. Cancer* **2006**, *6*, 857–866. [CrossRef] [PubMed]

47. Voigt, W.; Manegold, C.; Pilz, L.; Wu, Y.L.; Mullauer, L.; Pirker, R.; Filipits, M.; Niklinski, J.; Petruzelka, L.; Prosch, H. Beyond tissue biopsy: A diagnostic framework to address tumor heterogeneity in lung cancer. *Curr. Opin. Oncol.* **2020**, *32*, 68–77. [CrossRef] [PubMed]

48. Ilie, M.; Hofman, P. Pros: Can tissue biopsy be replaced by liquid biopsy? *Transl. Lung Cancer Res.* 2016, 5, 420–423. [CrossRef] [PubMed]

49. Mino-Kenudson, M. Cons: Can liquid biopsy replace tissue biopsy?-the US experience. *Transl. Lung Cancer Res.* **2016**, *5*, 424–427. [CrossRef] [PubMed]

50. Gerlinger, M.; Rowan, A.J.; Horswell, S.; Math, M.; Larkin, J.; Endesfelder, D.; Gronroos, E.; Martinez, P.; Matthews, N.; Stewart, A.; et al. Intratumor heterogeneity and branched evolution revealed by multiregion sequencing. *N. Engl. J. Med.* **2012**, *366*, 883–892. [CrossRef]

51. Mitchell, P.S.; Parkin, R.K.; Kroh, E.M.; Fritz, B.R.; Wyman, S.K.; Pogosova-Agadjanyan, E.L.; Peterson, A.; Noteboom, J.; O'Briant, K.C.; Allen, A.; et al. Circulating microRNAs as stable blood-based markers for cancer detection. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 10513–10518. [CrossRef]

52. Chen, X.; Ba, Y.; Ma, L.; Cai, X.; Yin, Y.; Wang, K.; Guo, J.; Zhang, Y.; Chen, J.; Guo, X.; et al. Characterization of microRNAs in serum: A novel class of biomarkers for diagnosis of cancer and other diseases. *Cell Res.* **2008**, *18*, 997–1006. [CrossRef] [PubMed]

53. Fernandez-Mercado, M.; Manterola, L.; Larrea, E.; Goicoechea, I.; Arestin, M.; Armesto, M.; Otaegui, D.; Lawrie, C.H. The circulating transcriptome as a source of non-invasive cancer biomarkers: Concepts and controversies of non-coding and coding RNA in body fluids. *J. Cell. Mol. Med.* **2015**, *19*, 2307–2323. [CrossRef]

54. Wan, J.C.M.; Massie, C.; Garcia-Corbacho, J.; Mouliere, F.; Brenton, J.D.; Caldas, C.; Pacey, S.; Baird, R.; Rosenfeld, N. Liquid biopsies come of age: Towards implementation of circulating tumour DNA. *Nat. Rev. Cancer* **2017**, *17*, 223–238. [CrossRef] [PubMed]

55. Fatima, F.; Nawaz, M. Vesiculated Long Non-Coding RNAs: Offshore Packages Deciphering Trans-Regulation between Cells, Cancer Progression and Resistance to Therapies. *Noncoding RNA* **2017**, *3*, 10. [CrossRef] [PubMed] 56. Hanna, J.; Hossain, G.S.; Kocerha, J. The Potential for microRNA Therapeutics and Clinical Research. *Front. Genet.* **2019**, *10*, 478. [CrossRef] [PubMed]

57. Gustafson, D.; Tyryshkin, K.; Renwick, N. microRNA-guided diagnostics in clinical samples. *Best Pract. Res. Clin. Endocrinol. Metab.* **2016**, *30*, 563–575. [CrossRef]

58. Sun, M.; Kraus, W.L. From discovery to function: The expanding roles of long noncoding RNAs in physiology and disease. *Endocr. Rev.* **2015**, *36*, 25–64. [CrossRef] [PubMed]

59. Lu, J.; Getz, G.; Miska, E.A.; Alvarez-Saavedra, E.; Lamb, J.; Peck, D.; Sweet-Cordero, A.; Ebert, B.L.; Mak, R.H.; Ferrando, A.A.; et al. MicroRNA expression profiles classify human cancers. *Nature* **2005**, *435*, 834–838. [CrossRef] [PubMed]

60. Bussemakers, M.J.; van Bokhoven, A.; Verhaegh, G.W.; Smit, F.P.; Karthaus, H.F.; Schalken, J.A.; Debruyne, F.M.; Ru, N.; Isaacs, W.B. DD3: A new prostate-specific gene, highly overexpressed in prostate cancer. *Cancer Res.* **1999**, *59*, 5975–5979.

61. Hessels, D.; Klein Gunnewiek, J.M.; van Oort, I.; Karthaus, H.F.; van Leenders, G.J.; van Balken, B.; Kiemeney, L.A.; Witjes, J.A.; Schalken, J.A. DD3(PCA3)-based molecular urine analysis for the diagnosis of prostate cancer. *Eur. Urol.* **2003**, *44*, 8–15; discussion 15–16. [CrossRef]

62. Groskopf, J.; Aubin, S.M.; Deras, I.L.; Blase, A.; Bodrug, S.; Clark, C.; Brentano, S.; Mathis, J.; Pham, J.; Meyer, T.; et al. APTIMA PCA3 molecular urine test: Development of a method to aid in the diagnosis of prostate cancer. *Clin. Chem.* **2006**, *52*, 1089–1095. [CrossRef] [PubMed]

63. Marks, L.S.; Bostwick, D.G. Prostate Cancer Specificity of PCA3 Gene Testing: Examples from Clinical Practice. *Rev. Urol.* **2008**, *10*, 175–181.

64. Lee, G.L.; Dobi, A.; Srivastava, S. Prostate cancer: Diagnostic performance of the PCA3 urine test. *Nat. Rev. Urol.* **2011**, *8*, 123–124. [CrossRef]

65. Hou, J.; Meng, F.; Chan, L.W.; Cho, W.C.; Wong, S.C. Circulating Plasma MicroRNAs As Diagnostic Markers for NSCLC. *Front. Genet.* **2016**, *7*, 193. [CrossRef] [PubMed]

66. Pine, S.R.; Ryan, B.M. Identifying therapeutic vulnerabilities in lung cancer: Application of a chemistry-first approach. *Transl. Lung Cancer Res.* **2018**, *7*, S265–S269. [CrossRef]

67. David, E.A.; Daly, M.E.; Li, C.S.; Chiu, C.L.; Cooke, D.T.; Brown, L.M.; Melnikow, J.; Kelly, K.; Canter, R.J. Increasing Rates of No Treatment in Advanced-Stage Non-Small Cell Lung Cancer Patients: A Propensity-Matched Analysis. *J. Thorac. Oncol.* **2017**, *12*, 437–445. [CrossRef]

68. Slack, F.J.; Chinnaiyan, A.M. The Role of Non-coding RNAs in Oncology. *Cell* **2019**, *179*, 1033–1055. [CrossRef] [PubMed]

69. Siegel, R.L.; Miller, K.D.; Jemal, A. Cancer statistics, 2020. CA Cancer J. Clin. 2020, 70, 7–30. [CrossRef]

70 Borondy Kitts, A.K. The Patient Perspective on Lung Cancer Screening and Health Disparities. *J. Am. Coll. Radiol.* **2019**, *16*, 601–606. [CrossRef]

71. Patop, I.L.; Kadener, S. circRNAs in Cancer. Curr. Opin. Genet. Dev. 2018, 48, 121–127. [CrossRef] [PubMed]

72. Calin, G.A.; Dumitru, C.D.; Shimizu, M.; Bichi, R.; Zupo, S.; Noch, E.; Aldler, H.; Rattan, S.; Keating, M.; Rai, K.; et al. Frequent deletions and down-regulation of micro- RNA genes miR15 and miR16 at 13q14 in chronic lymphocytic leukemia. *Proc. Natl. Acad. Sci. USA* **2002**, *99*, 15524–15529. [CrossRef] [PubMed]

73. Sun, M.; Kraus, W.L. From discovery to function: The expanding roles of long noncoding RNAs in physiology and disease. *Endocr. Rev.* **2015**, *36*, 25–64. [CrossRef] [PubMed]

74. Luo, M.L. Methods to Study Long Noncoding RNA Biology in Cancer. *Adv. Exp. Med. Biol.* **2016**, *927*, 69–107. [CrossRef]

75. Choudhari, R.; Sedano, M.J.; Harrison, A.L.; Subramani, R.; Lin, K.Y.; Ramos, E.I.; Lakshmanaswamy, R.; Gadad, S.S. Long noncoding RNAs in cancer: From discovery to therapeutic targets. *Adv. Clin. Chem.* **2020**, *95*, 105–147. [CrossRef] [PubMed]

76. Zhang, P.; Wu, W.; Chen, Q.; Chen, M. Non-Coding RNAs and their Integrated Networks. J. Integr. Bioinform. 2019, 16. [CrossRef]

77. Moyano, M.; Stefani, G. piRNA involvement in genome stability and human cancer. *J. Hematol. Oncol.* **2015**, *8*, 38. [CrossRef]

78. Slack, F.J.; Chinnaiyan, A.M. The Role of Non-coding RNAs in Oncology. *Cell* **2019**, *179*, 1033–1055. [CrossRef] [PubMed]

79 Hanahan, D.; Weinberg, R.A. Hallmarks of cancer: The next generation. *Cell* **2011**, *144*, 646–674. [CrossRef] [PubMed]

80. Hanahan, D.; Weinberg, R.A. The hallmarks of cancer. Cell 2000, 100, 57–70. [CrossRef]

81. Ling, H.; Girnita, L.; Buda, O.; Calin, G.A. Non-coding RNAs: The cancer genome dark matter that matters! *Clin. Chem. Lab. Med.* **2017**, *55*, 705–714. [CrossRef] [PubMed]

82. Calin, G.A.; Croce, C.M. MicroRNA signatures in human cancers. *Nat. Rev. Cancer* 2006, *6*, 857–866. [CrossRef] [PubMed]

83. Salmena, L.; Poliseno, L.; Tay, Y.; Kats, L.; Pandolfifi, P.P. A ceRNA hypothesis: The Rosetta Stone of a hidden RNA language? *Cell* **2011**, *146*, 353–358. [CrossRef]

84. Wang, Y.; Lee, C.G. MicroRNA and cancer—Focus on apoptosis. J. Cell. Mol. Med. 2009, 13, 12–23. [CrossRef] [PubMed]

85. Hayes, J.; Peruzzi, P.P.; Lawler, S. MicroRNAs in cancer: Biomarkers, functions and therapy. *Trends Mol. Med.* **2014**, *20*, 460–469. [CrossRef]

86. Krek, A.; Grun, D.; Poy, M.N.; Wolf, R.; Rosenberg, L.; Epstein, E.J.; MacMenamin, P.; da Piedade, I.; Gunsalus, K.C.; Stoffel, M.; et al. Combinatorial microRNA target predictions. *Nat. Genet.* **2005**, *37*, 495–500. [CrossRef] [PubMed]

87. Fabbri, M.; Paone, A.; Calore, F.; Galli, R.; Gaudio, E.; Santhanam, R.; Lovat, F.; Fadda, P.; Mao, C.; Nuovo, G.J.; et al. MicroRNAs bind to Toll-like receptors to induce prometastatic inflflammatory response. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, E2110–E2116. [CrossRef] [PubMed]

88. Fabbri, M.; Paone, A.; Calore, F.; Galli, R.; Croce, C.M. A new role for microRNAs, as ligands of Toll-like receptors. *RNA Biol.* **2013**, *10*, 169–174. [CrossRef] [PubMed]

89. Qiao, D.; Zeeman, A.M.; Deng, W.; Looijenga, L.H.; Lin, H. Molecular characterization of hiwi, a human member of the piwi gene family whose overexpression is correlated to seminomas. *Oncogene* **2002**, *21*, 3988–3999. [CrossRef]

90. Zhao, Y.M.; Zhou, J.M.; Wang, L.R.; He, H.W.; Wang, X.L.; Tao, Z.H.; Sun, H.C.; Wu, W.Z.; Fan, J.; Tang, Z.Y.; et al. HIWI is associated with prognosis in patients with hepatocellular carcinoma after curative resection. *Cancer* **2012**, *118*, 2708–2717. [CrossRef]

91. Esquela-Kerscher, A.; Slack, F.J. Oncomirs—Micrornas with a role in cancer. *Nat. Rev. Cancer* **2006**, *6*, 259–269. [CrossRef]

92. Svoronos, A.A.; Engelman, D.M.; Slack, F.J. OncomiR or Tumor Suppressor? The Duplicity of MicroRNAs in Cancer. *Cancer Res.* **2016**, *76*, 3666–3670. [CrossRef]

93. Wilusz JE, Sunwoo H, Spector DL. Long noncoding RNAs: functional surprises from the RNA world. Genes Dev 2009; 23: 1494–1504.

94. Huttenhofer A, Schattner P, Polacek N. Non-coding RNAs: hope or hype? Trends Genet: TIG 2005; 21: 289–297.

95. Babalola O, Mamalis A, Lev-Tov H, Jagdeo J. The role of microRNAs in skin fifibrosis. Arch Dermatol Res 2013; 305: 763–776.

96. Bergmann JH, Spector DL. Long non-coding RNAs: modulators of nuclear structure and function. Curr Opin Cell Biol 2014; 26: 10–18.

97. Cusanelli E, Chartrand P. Telomeric noncoding RNA: telomeric repeat-containing RNA in telomere biology. Wiley Interdiscip Rev RNA 2014;5: 407–419.

98. Lujambio A, Portela A, Liz J, Melo SA, Rossi S, Spizzo R, et al. CpG island hypermethylation-associated silencing of non-coding RNAs transcribed from ultraconserved regions in human cancer. Oncogene 2010; 29: 6390–6401.

99. Esteller M. Non-coding RNAs in human disease. Nat Rev Genet 2011; 12: 861–874.

100. Xu B, Gerin I, Miao H, Vu-Phan D, Johnson CN, Xu R, et al. Multiple roles for the non-coding RNA SRA in regulation of adipogenesis and insulin sensitivity. PLoS One 2010; 5: e14199.

101. Zhou Y, Zhang X, Klibanski A. MEG3 noncoding RNA: a tumor suppressor. J Mol Endocrinol 2012; 48: R45–R53.

102. Kino T, Hurt DE, Ichijo T, Nader N, Chrousos GP. Noncoding RNA gas5 is a growth arrest- and starvation-associated repressor of the glucocorticoid receptor. Sci Signal 2010; 3: ra8.

103. Swanton C, Caldas C. Molecular classifification of solid tumours: towards pathway-driven therapeutics. Br J Cancer 2009; 100: 1517–1522.