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STUDTY OF AEROMYCOFLORA IN INDOOR ENVIRONMENT OF COLLEGE BUILDING, GOVT.NAVEEN COLLEGE, SONAKHAN

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Abstract:-Fungi are cosmopolitan in distribution and occur in any habitat where life is possible. Aeromycoflora which is refers to the airborne fungal spores in the atmosphere. A large number of aeromycoflora were found in indoor and also outdoor environment. The indoor environment is much more important as a major part of the day is spent indoor. The indoor fungi are not only responsible for causing disease but also responsible for the spoilage of food, clothes, books etc. Indoor environments play important roles in human health. The health hazards posed by polluted indoor environment. The primary objective of this study was to identify the aeromycoflora survey of government college building of Sonakhan, dist. Balodabazar-Bhatapara(C.G.). Study carried out between June and October 2023. The indoor aeromycoflora species was represented by Aspergillus, Penicillium, Rhizopus, Cladosporium, and Alternaria.

KEYWORDS:-Cosmoplitan,aeromycoflora,sonakhan, Aspergillus.

1.INTRODUCTION:- Fungi are ubiquitous and versatile. The enormous types of spores ,fungal propagules are found in large numbers in indoor and outdoor environment. Some of them have the potentiality to cause spoilage of food, allergies and many other adverse health effects namely allergie, bronchial, as thma and rhinitis. Fungi are eukeryotic and spore bearing organism, which exist as saprophyte or as parasite of animal and plants. Fungi spores are generally considerd to be significant cause of both allergic rhinitis and allergic as thma. This paper is deals with the aeromycoflora survey of Govt. Naveen College, Sonakhan . Aeromycoflora of college was studied by petriplate method . Enviornmental factor play an important role for the distribution of the fungal spore .

Fungal flora of the air was determined from different parts of the indoor environment of the college campus of Govt.Naveen College ,Sonakhan every month during investigation period from June to October 2023.

2.METHODS AND MATERIALS: for study of aeromycoflora we required :

i.Study site :Endoor environment of Govt.Naveen College ,Sonakhan Dist.-Balodabazar-Bhatapara.

ii.Culture Media :for cultur of aeromycoflra ,PDA(Potato Dextrose Agar) media was used .

Petriplats exposur method was used for the isolation of aeromycoflora from the environment every month. Five petriplats containing PDA media were exposed for 10 to 20 minutes in the endoor environment of college. After, these exposed petriplats were brought to laboratory and incubated at 28+1c for 3 to 5 days. At the end of the incubation period the fungal colonies were counted, isolated and identified with the help of available literature.

Microphotography was performed by digital microscope in the laboratory of department of botany.



Fig:Fungal colonies growing on the Petriplate of different sampling site in the indoor environment at the Govt.Naveen College ,Sonakhan.

3.ECOLOGICAL STUDIES:-Percentage frequency and percentage contribution of fungal flora is calculated (Sharma K.2001) by the following formula:

Number of observation in which a species appeared Percentage frequency = ------ x 100 Total no.of observations

Total no.of colonies of a species in all observations Percentage contribution = ------ x 100 Total no.of colonies of all the species

4.RESULT AND DISCUSSION:-Indoor environment is rich in aeromycoflora,fungal species and fungal spore.

We isolated total 30 fungal flora from research site and categorized into three major group i.e.Zygomycotina,Ascomycotina and Anamorphic fungi.Environmental factor play a key role in fungal distribution. Highest fungal flora 23 out of 30 were isolated from anamorphic group .According to their distribution in the petriplates the highest percentage frequency was recorded as Aspergillus niger, Aspergillus nidulans,A.flavus,A.fumigates,A.oryzae,Rhizopus stolonifera,Rhizopus oryzae,Cladosporium sp. And Mycelia sterile white(100%) wheres lowest percentage frequency is represents by Gilmaniella humicola (20%)(Table-

2). Aspergillus ,Rhizopus ,Penicilium sp.and unidentified sterile hyphae is more common in indoor environment . In this research we observed the highest percentage contribution in Aspergillus niger (10.87) and lowest percentage contribution of Gilmaniella humicola(0.08). Distribution of fungal spore are affected by geographical location and meterological condition. In the month of October high percentage of humidity and moderate range of temperature the spore concentration was more . The higher number of aeromycoflora was reported in the month of July and October.

Table: 1 Isolated aeromycoflora of indoor environment at Govt.Naveen College,Sonakhan from June to
October 2023.

| S.No | Name of Fungal species | June | July | Aug. | Sep. | Oct. | Total |
|------|-----------------------------------|------|------|------|------|------|-------|
| 1 | A amongillug florug | 10 | 12 | 10 | 5 | 0 | 51 |
| 1 | Asperginus navus | 18 | 12 | 10 | 3 | 9 | 34 |
| 2 | Aspergillus fumigatus | 21 | 15 | 13 | 22 | 1/ | 88 |
| 3 | Aspergillus luchensis | 8 | - | 2 | 16 | 34 | 60 |
| 4 | Aspergillus nidulans | 16 | 8 | 11 | 23 | 10 | 68 |
| 5 | Aspergillus niger | 14 | 20 | 28 | 16 | 41 | 119 |
| 6 | Aspergillus oryz <mark>ae</mark> | 14 | 6 | 9 | 5 | 16 | 50 |
| 7 | Rhizopus nigricans | 3 | 5 | 8 | 1 | - | 17 |
| 8 | Rhizopus stolinife <mark>r</mark> | 4 | 1 | 4 | 6 | 5 | 19 |
| 9 | Rhizopus oryzae | 3 | 1 | 4 | 1 | 2 | 11 |
| 10 | Penicillium chrysogenum | 16 | 3 | 6 | 8 | - | 33 |
| 11 | Penicillium citrinum | 1 | | - | - | 5 | 6 |
| 12 | Penicillium expansum | 2 | 1 | - | - | 2 | 5 |
| 13 | Penicillium italicum | 1 | 3 | - | - | - | 4 |
| 14 | Cladosporium | 22 | 16 | 38 | 10 | 23 | 99 |
| 15 | cladosporioides | 20 | 16 | 0 | | | 70 |
| 15 | Cladosporium nerbarum | 30 | 15 | 8 | 14 | 0 | /3 |
| 16 | Cladosporium raphigera | 3 | - | 2 | | 1 | 6 |
| 17 | Cladosporium oxysporium | 23 | 16 | 10 | 36 | 18 | 103 |
| 18 | Curvularia clavata | 10 | - | - | 8 | 1 | 19 |
| 19 | Alternaria altarnata | 17 | 9 | 16 | 13 | 21 | 76 |
| 20 | Mucor varians | 3 | - | 1 | 6 | - | 10 |
| 21 | Mucor racemosum | 2 | 4 | - | - | 1 | 7 |
| 22 | Chaetomium globosum | 1 | 5 | 9 | 4 | 1 | 20 |
| 23 | Myrothecium roridum | 3 | 1 | - | 4 | 3 | 11 |
| 24 | Fusarium chlamydosporum | 6 | 9 | 4 | - | 13 | 32 |
| 25 | Fusarium moniliformae | 8 | 4 | 2 | 1 | 3 | 18 |
| 26 | Curvularia lunata | - | 2 | 1 | - | 5 | 8 |
| 27 | Alternaria citri | 3 | 8 | - | 12 | 4 | 27 |
| 28 | Alternaria tenuissima | 6 | 9 | 17 | 8 | 2 | 42 |
| 29 | Gilmaniella humicola | - | - | 1 | - | - | 1 |
| 30 | Mycelia sterile white | 21 | 14 | 16 | 32 | 10 | 83 |
| | Total | | | | | | 1169 |
| | | | | | | | |

| S.No. | Name of fungal species | Total | Percentage Frequency | Percentage |
|-------|----------------------------------|-------|----------------------|--------------|
| | | | | Contribution |
| 1 | Aspergillus flavus | 54 | 100 | 4.61 |
| 2 | Aspergillus fumigatus | 88 | 100 | 7.52 |
| 3 | Aspergillus luchensis | 60 | 80 | 5.13 |
| 4 | Aspergillus nidulans | 68 | 100 | 5.81 |
| 5 | Aspergillus niger | 119 | 100 | 10.17 |
| 6 | Aspergillus oryzae | 50 | 100 | 4.27 |
| 7 | Rhizopus nigricans | 17 | 80 | 1.45 |
| 8 | Rhizopus stolinifer | 19 | 100 | 1.62 |
| 9 | Rhizopus oryzae | 11 | 100 | 0.94 |
| 10 | Penicillium chrysogenum | 33 | 80 | 2.82 |
| 11 | Penicillium citrinum | 06 | 40 | 0.51 |
| 12 | Penicillium expansum | 05 | 60 | 0.42 |
| 13 | Penicillium italicum | 04 | 40 | 0.34 |
| 14 | Cladosporium | 99 | 100 | 8.46 |
| | cladosporioides | | | |
| 15 | Cladosporium herbarum | 73 | 100 | 6.24 |
| 16 | Cladosporium raphigera | 06 | 60 | 0.51 |
| 17 | Cladosporium | 103 | 100 | 8.81 |
| | oxysporium | / | | |
| 18 | Curvularia cl <mark>avata</mark> | 19 | 60 | 1.62 |
| 19 | Alternaria alternata | 76 | 100 | 6.50 |
| 20 | Mucor varians | 10 | 60 | 0.85 |
| 21 | Mucor racemosum | 07 | 60 | 0.59 |
| 22 | Chaetomium globosum | 20 | 100 | 1.71 |
| 23 | Myrothecium roridum | 11 | 80 | 0.94 |
| 24 | Fusarium | 32 | 80 | 2.73 |
| | chlamydosporum | | | |
| 25 | Fusarium moniliformae | 18 | 100 | 1.53 |
| 26 | Curvularia lunata | 08 | 60 | 0.68 |
| 27 | Alternaria citri | 27 | 80 | 2.30 |
| 28 | Alternaria tenuissima | 42 | 100 | 3.59 |
| 29 | Gilmaniella humicola | 01 | 20 | 0.08 |
| 30 | Mycelia sterile white | 83 | 100 | 7.10 |

| Table -2 | Ecological | fluctuation | of Aeromycoflora | |
|----------|------------|-------------|------------------|--|
| 14010 -2 | Leological | Inactuation | of Actomyconora | |

5.CONCLUSION: The primary objective of this study was to identify the aeromycoflora, their concentration and diversity in the indoor environment. A large number of aeromycoflora were found in indoor and also outdoor environment .The indoor environment is much more important as a major part of the day is spent indoor. The indoor fungi are not only responsible for causing disease but also responsible for the spoilage of food, clothes, books etc. Indoor environments play important roles in human health. The detail study based on the atmospheric parameter could provide data for more understanding the correlation between abiotic and biotic factor. The effect of aeromycoflora , airborne fungal spore on human health is also point of concern which could be studied by the use of preliminary data provided in this research article.

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