



FORMULATION AND EVALUATION OF POLYHERBAL FACIAL CREAM CONTAINING *CURCUMA LONGA* AND *ALOE BARBADENSIS*

Dr. Revan Karodi¹, Mr. Rushikesh G. Tagad², Mr. Sushant Ahire³, Ms. Sonam Bendre⁴
HOD, Pharmacognosy Department¹
P.G. Scholar, Pharmaceutical Quality Assurance^{2,3,4}
Dr. D. Y. Patil College of Pharmacy, Akurdi, Pune

ABSTRACT

This study focuses on the development of a polyherbal facial cream containing curcuma longa (turmeric) and aloe barbadensis (aloe vera) extracts and evaluates its physicochemical properties, stability, and potential benefits for the skin. The introduction highlights the therapeutic properties and traditional use of these botanical ingredients in skincare. Curcuma longa is known for its antioxidant, anti-inflammatory, and antimicrobial properties, while aloe barbadensis offers moisturizing, soothing, and wound-healing effects, along with anti-inflammatory properties and collagen synthesis enhancement. The method of preparation involves two phases: the oil phase and the aqueous phase. The oil phase consists of ingredients such as stearic acid, white bees wax, stearyl alcohol, and cetyl alcohol, while the aqueous phase includes propylene glycol, triethanolamine, methyl paraben, propylparaben, and water. These phases are prepared separately and then combined to form the cream formulation. Overall, this study aims to contribute to the development of natural, effective, and safe skincare products by formulating a polyherbal cream containing Curcuma longa and Aloe barbadensis extracts and evaluating its properties and benefits for the skin.

KEYWORDS

Polyherbal, Cream, *Curcuma longa*, *aloe barbadensis*

INTRODUCTION

Polyherbal formulations, combining multiple botanical ingredients, have gained significant attention in the field of skincare due to their potential synergistic effects and holistic approach to addressing various skin concerns. Among the numerous botanical extracts utilized in cosmetic formulations, Curcuma longa (turmeric) and Aloe barbadensis (aloe vera) have emerged as prominent ingredients, valued for their therapeutic properties and longstanding traditional use in skincare.

Curcuma longa, a perennial plant native to South Asia, has been extensively studied for its bioactive components, notably curcuminoids. Curcumin, the principal curcuminoid found in turmeric, possesses antioxidant, anti-inflammatory, and antimicrobial properties, making it a promising ingredient in skincare formulations. Its ability to scavenge free radicals and inhibit inflammatory pathways can help protect the skin from oxidative stress and reduce the signs of aging, including wrinkles and pigmentation disorders.

The succulent shrub Aloe barbadensis, also known as aloe vera, has been used for millennia in traditional medicine and cosmetics. The gel extracted from aloe vera leaves contains a variety of bioactive compounds,

including polysaccharides, vitamins, minerals, and enzymes. These components contribute to aloe vera's moisturizing, soothing, and wound-healing properties. Aloe vera also exhibits anti-inflammatory effects, enhances collagen synthesis, and improves the skin's elasticity, making it a valuable ingredient for skincare products.

In this study, we aim to develop a polyherbal facial cream containing *Curcuma longa* and *Aloe barbadensis* and evaluate its physicochemical properties, stability, and potential benefits for the skin. By exploring the formulation of this polyherbal cream, we seek to contribute to the development of natural, effective, and safe skincare products. (1-4)

Method of Preparation (5,6,7)

Oil phase preparation (part a): The ingredients used in the oil phase are Stearic Acid, White Bees Wax, Stearyl Alcohol, and cetyl Alcohol. White Bees Wax, stearic acid, stearyl alcohol, and cetyl alcohol were added to a beaker and allowed to melt. The temperature of the oil phase is maintained between 65 –70°C.

Aqueous phase preparation (part b): Ingredients used in the aqueous phase are Propylene Glycol, Triethanolamine, Methyl Paraben, Propyl paraben, and Distilled Water. The water was heated to a temperature of 65-70 °C. Propylene Glycol, Triethanolamine, Methyl paraben, and Propyl paraben were weighed out and added to the phase, which was kept at a temperature of 65 to 70 °C.

Development of cream formulation (part a + part b): The oil phase and aqueous phase are brought at the same temperature, then the oil phase is slowly incorporated into the aqueous phase at 65-70°C and mixed for 10 to 15 Minutes in a mortar pestle with moderate agitation in a clockwise direction until the temperature is dropped to 40°C and a clicking sound is heard.

Table no.1 Formulation of Herbal Facial Cream

Ingredient	F1	F2	F3	F4	F5	F6	F7	F8	F9
<i>Curcuma longa</i>	1	1	1	1	1	1	1	1	1
<i>Aloe barbadensis</i>	1	1	1	1	1	1	1	1	1
Honey	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5
Coconut oil	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5
Stearic Acid	0.4	0.1	0.25	0.0378	0.25	0.1	0.25	0.4621	0.4
White Bees Wax	0.3	0.3	0.3	0.3	0.3	0.3	0.3	0.3	0.3
Stearyl Alcohol	1	1	1	1	1	1	1	1	1
Cetyl Alcohol	0.1	1.2	0.65	0.65	-0.1278	0.1	0.4278	0.65	1.2
Propylene glycol	1	1	1	1	1	1	1	1	1
Triethanolamine	0.4	0.4	0.4	0.4	0.4	0.4	0.4	0.4	0.4
Methyl Paraben	0.002	0.002	0.002	0.002	0.002	0.002	0.002	0.002	0.002
Propyl paraben	0.008	0.008	0.008	0.008	0.008	0.008	0.008	0.008	0.008
Distilled Water	Q.s.	Q.s.	Q.s.	Q.s.	Q.s.	Q.s.	Q.s.	Q.s.	Q.s.

Result and Discussion

Determination of extract using UV spectroscopic method:

Determination of extract of *Curcuma longa* using UV spectroscopic method

UV Spectrum of *Curcuma longa* extract (maximum wavelength was found to be 425nm).

Table no.2 UV analysis of *Curcuma longa* extract

Sr.no.	Concentration	Absorbance (λ max at 246nm)
1.	0.2 μ g	0.047
2.	0.4 μ g	0.061
3.	0.6 μ g	0.080
4.	0.8 μ g	0.107
5.	1.0 μ g	0.125
6.	R²	0.9969

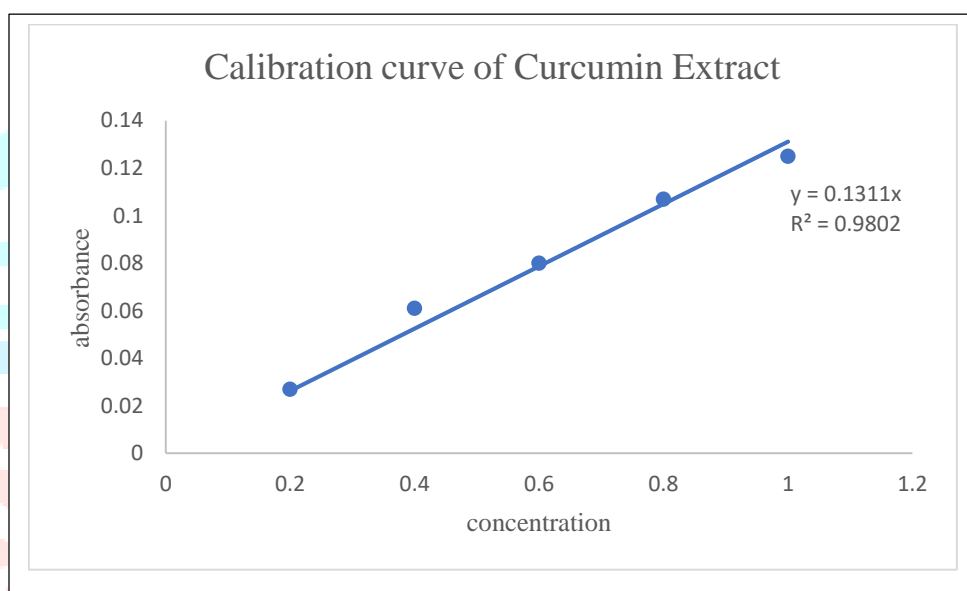


Fig no.1 *Curcuma longa* extract UV Calibration curve

Determination of the extract of *Aloe barbadensis* Miller by UV spectroscopic method:

UV Spectrum of *Aloe barbadensis miller* extract (maximum wavelength was found to be 263nm).

Table no. 3 UV analysis of the extract of *Aloe barbadensis* Miller

Sr.no.	Concentration	Absorbance (λ max at 246nm)
1.	0.2 μ g	0.011
2.	0.4 μ g	0.018
3.	0.6 μ g	0.023
4.	0.8 μ g	0.03
5.	1.0 μ g	0.036
6.	R²	0.9979

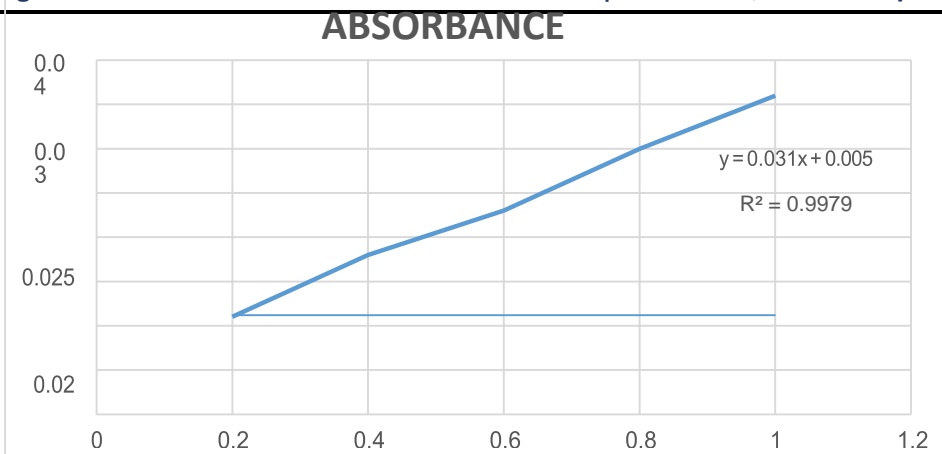


Fig no.2 Aloe barbadensis miller extract UV calibration curve

FTIR Study

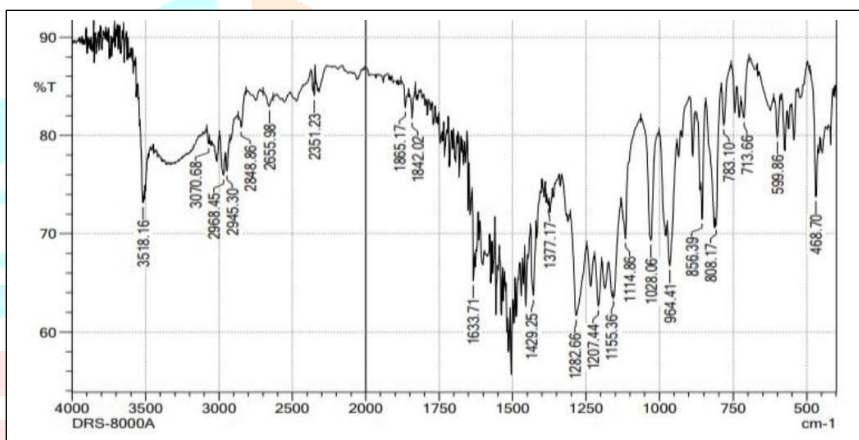


Fig no. 3 FTIR of *curcuma longa*

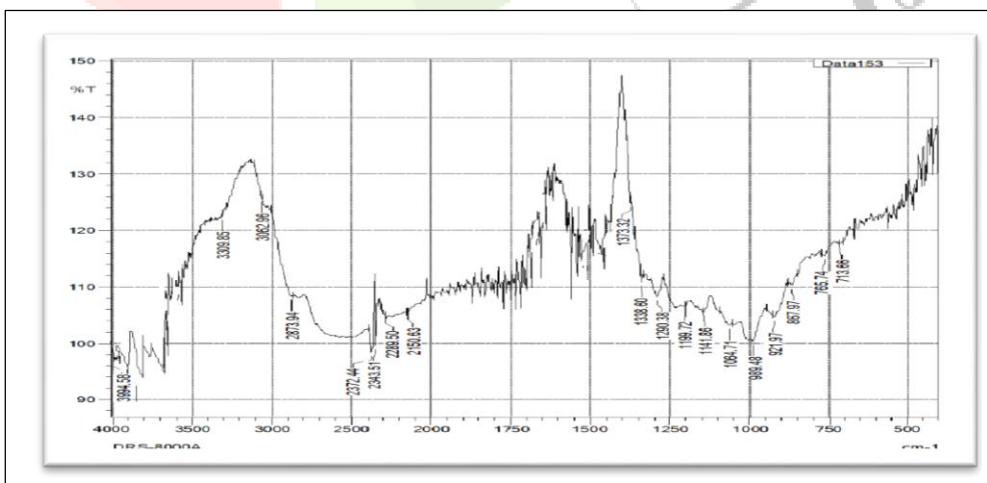


Fig no. 4 FTIR of *Aloe barbadensis*

HPTLC Analysis of Extracts:

HPTLC analysis of *Curcuma longa* extract:

Photo-documentation of *Curcuma longa* extract:

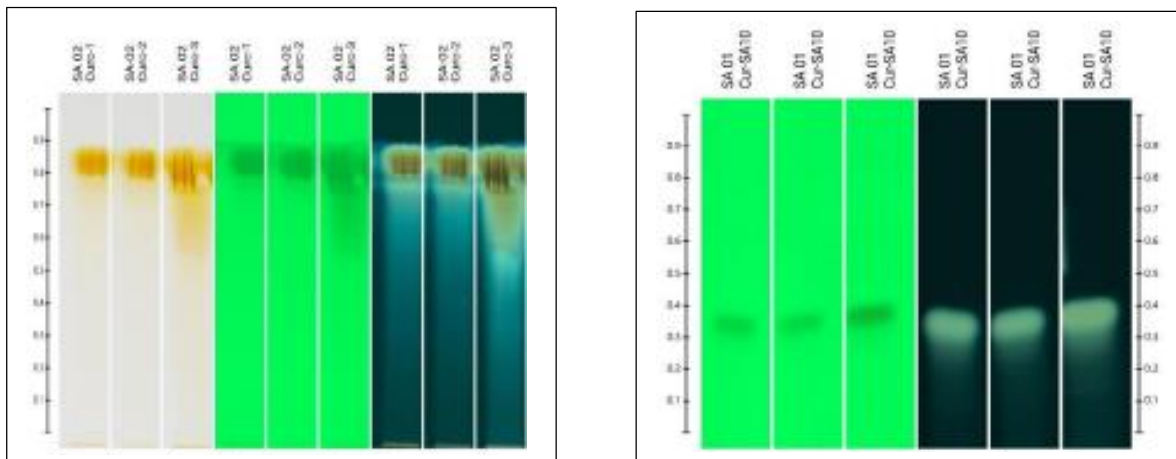


Fig. Photo documentation at 1) Visible 2) 254 nm and 3) 366 nm

Fig no. 5 Photo-Documentation of *Curcuma longa* extract

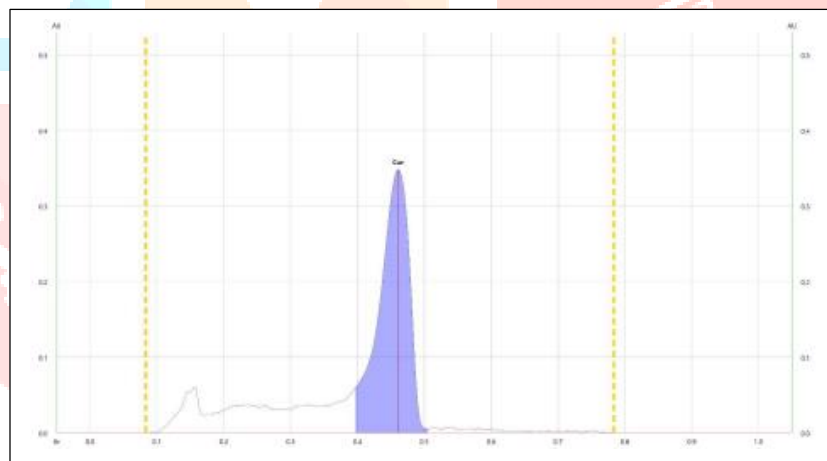


Fig no. 6 Densitogram of *Curcuma longa*

Observation:

- The peak can be seen at the solvent front and also there is merging of bands.
- Both UV 254 and UV 366 nm show a peak. The peak was once more scanned at 422 nm, the maximum wavelength for curcumin, and the densitogram was obtained.
- Scan results revealed an Rf of 0.47.

HPTLC analysis of *Aloe barbadensis miller* extract:

Photo-documentation of *Aloe barbadensis miller* extract

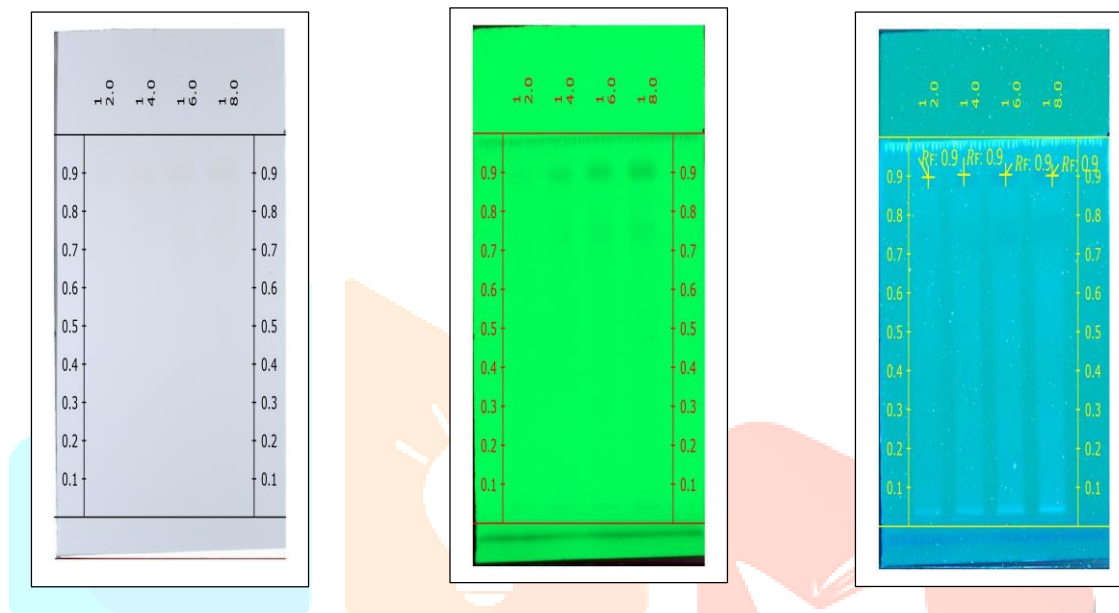


Fig no. 7 Photo-Documentation of *aloe barbadensis miller* extract

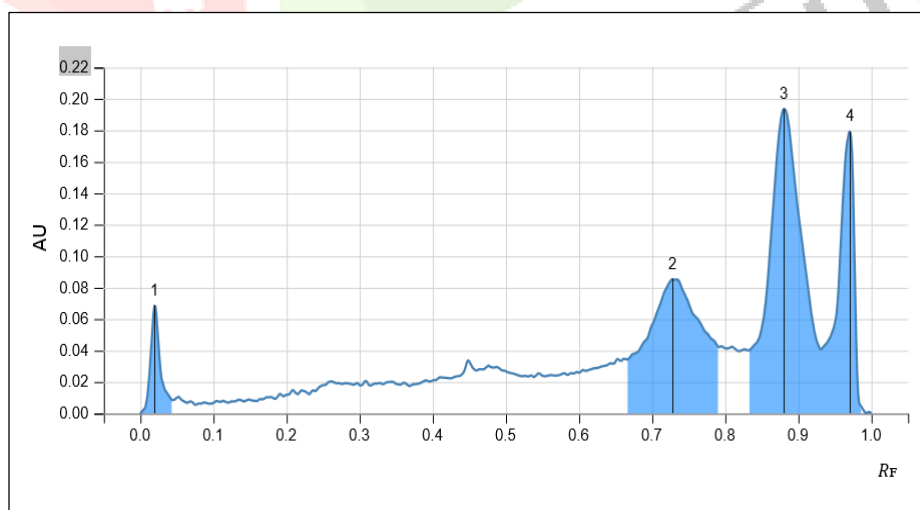


Fig no. 8 Representative Densitogram of *aloe barbadensis miller*

Evaluation of Cream:**Organoleptic Evaluation:**

Organoleptic evaluation of all 9 batches was performed. The color, Texture, homogeneity, and foreign particles of formulated batches were observed in table.

Table no. 4 Organoleptic Evaluation

Batch	Color	Texture	Homogeneity	Foreign particles
Batch no. F1	Faint Yellowish	Smooth	Excellent	Free from foreign particles
Batch no. F2	Yellowish	Smooth	Excellent	Free from foreign particles
Batch no. F3	Yellowish	Smooth	Excellent	Free from foreign particles
Batch no. F4	Yellowish	Smooth	Excellent	Free from foreign particles
Batch no. F5	Dark Yellowish	Smooth	Excellent	Free from foreign particles
Batch no. F6	Dark Yellowish	Smooth	Excellent	Free from foreign particles
Batch no. F7	Dark Yellowish	Smooth	Excellent	Free from foreign particles
Batch no. F8	Yellowish	Smooth	Excellent	Free from foreign particles
Batch no. F9	Yellowish	Smooth	Excellent	Free from foreign particles

Measurement of PH

Digital pH meter was used to measure the pH of Herbal Cream. The evaluation of all 9 batches was performed. The PH of all batches shows the pH range that is suitable for the skin pH and the result is shown in table.

Table no. 5 Observed pH

Formulation no.	Observed PH
Batch no. F1	6.3
Batch no. F2	6.9
Batch no. F3	5.9
Batch no. F4	6.0
Batch no. F5	5.7
Batch no. F6	5.1
Batch no. F7	6.2
Batch no. F8	5.6
Batch no. F9	5.8

Spreadability:

By placing a sample between two slides, then compressing it to uniform thickness and by setting the definite weight for a specified period, the spreadability of herbal cream has been measured which is shown in the table. (8)

Table no. 6 Observed Spreadability

Formulation no	Observed Spreadability [CM]
Batch no. F1	3.9cm
Batch no. F2	4.2cm
Batch no. F3	2.26cm
Batch no. F4	1.8cm
Batch no. F5	3.2cm
Batch no. F6	3.6cm
Batch no. F7	3.9cm
Batch no. F8	3.5cm
Batch no. F9	3.7cm

Viscosity

The Brooke field viscometer was used to perform the viscosity of herbal cream. The cream was dipped into a viscometer and readings of the cream were observed at room temperature and the result is given in table. (8,9)

Table no.7 Observed Viscosity

Formulation no.	Observed viscosity
Batch no. F1	14162cp
Batch no. F2	15870cp
Batch no. F3	15283cp
Batch no. F4	15120cp
Batch no. F5	14000cp
Batch no. F6	13976cp
Batch no. F7	13988cp
Batch no. F8	13957cp
Batch no. F9	12999cp

Drug Diffusion Test:

The in vitro drug release from cream was carried out using Franz diffusion cell at 50 RPM and $37^{\circ}\text{C} \pm 0.5^{\circ}\text{C}$. All Phosphate buffer was used as the dissolution medium. 1 ml of dissolution medium was withdrawn at predetermined time intervals and fresh dissolution was replaced. The sample was withdrawn at regular intervals and analyzed by UV-spectrophotometer at 246 nm and 263 nm for the presence of the drug and the result is shown in table.

Table no.8 Observed Drug Release

Formulation no.	Observed drug release
Batch no. F1	62.785%
Batch no. F2	58.063%
Batch no. F3	91.365%
Batch no. F4	52.458%
Batch no. F5	29.53125%
Batch no. F6	45.235%
Batch no. F7	72.975%
Batch no. F8	83.125%
Batch no. F9	64.658%

Wash Ability

A small amount of formulated cream was applied to the skin and the ease of washing of cream with water was checked.

Table no. 9 Observed Washability

Formulation no.	Observed Washability
Batch no. F1	Easily washable
Batch no. F2	Easily washable
Batch no. F3	Easily washable
Batch no. F4	Easily washable
Batch no. F5	Easily washable
Batch no. F6	Easily washable
Batch no. F7	Easily washable
Batch no. F8	Easily washable
Batch no. F9	Easily washable

Centrifugation Test

The centrifugation testing was carried out for all 9 herbal cream formulations when the creams are placed in a centrifuge machine and then they don't show any separation of two phases then all 9 formulations pass centrifugation testing. (10)

Table no. 10 Centrifugation Test

Formulation no.	Centrifugation test
Batch no. F1	No phase separation
Batch no. F2	No phase separation
Batch no. F3	No phase separation
Batch no. F4	No phase separation
Batch no. F5	No phase separation
Batch no. F6	No phase separation
Batch no. F7	No phase separation
Batch no. F8	No phase separation
Batch no. F9	No phase separation

Freeze and Thaw Test

The cream was exposed to low temperature in a refrigerator and then brought to room temperature. Results were observed for all 9 batches visually such as changes in the color and results are shown in table (11).

Table no. 11 Freeze and Thaw Test

Formulation no.	Freeze and Thaw Test
Batch no. F1	Passes
Batch no. F2	Passes
Batch no. F3	Passes
Batch no. F4	Passes
Batch no. F5	Passes
Batch no. F6	Passes
Batch no. F7	Passes
Batch no. F8	Passes
Batch no. F9	Passes

Particle Size of Cream:

Globule size and its distribution in Cream-

The globules in the formulation were found to have an average size of 100 nm and a zeta potential of -2.1 mv. In photomicrograph, appropriately diluted emulsions of the optimized batches were observed under a light microscope at 40x. Almost spherical globules of emulsion were seen in the photomicrograph. Though this study does not give any exact estimate of size however it gives a general idea about formation of cream and success of the method used. (12,13)

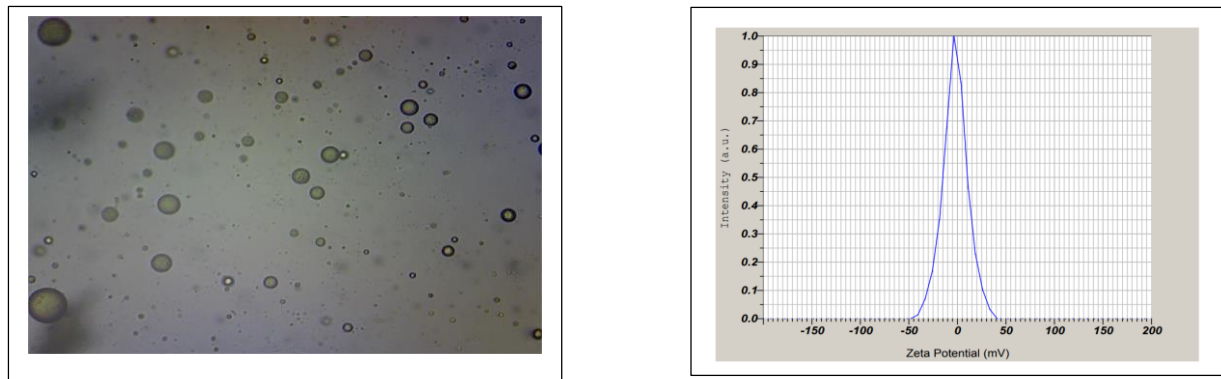


Fig no. 9 Particle Size of Formulated Batch

After Feel Test

All 9 batches of the herbal cream formulation underwent an after-feel test. The sample of cream has been tested for its smoothness, emollient characteristics and the amount of residue that remained after it was applied. (14,15)

Table no. 12 after Feel Test

Formulation no.	After feel test
Batch no. F1	Emollient
Batch no. F2	Emollient
Batch no. F3	Emollient
Batch no. F4	Emollient
Batch no. F5	Emollient
Batch no. F6	Emollient
Batch no. F7	Emollient
Batch no. F8	Emollient
Batch no. F9	Emollient

Accelerated Stability Study:

The formulated herbal cream was tested for accelerated stability by in vitro evaluation of herbal cream it was placed in a stability chamber at different temperatures ($20^{\circ}\text{C} \pm 1^{\circ}\text{C}$ and $40^{\circ}\text{C} \pm 1^{\circ}\text{C}$) for 30 days. All the formulations were found to be stable upon storage for 30 days. No change was observed in their appearance, pH, homogeneity, spreadability, viscosity, after-feel, Washability, and irritation result are shown in table. (16)

Table no. 13 Accelerated Stability Study

Formulation no.	Accelerated Stability Study
Batch no. F1	No change was observed.
Batch no. F2	No change was observed.
Batch no. F3	No change was observed.
Batch no. F4	No change was observed.
Batch no. F5	No change was observed.
Batch no. F6	No change was observed.
Batch no. F7	No change was observed.
Batch no. F8	No change was observed.
Batch no. F9	No change was observed.

Antimicrobial Activity Study:

The facial activity of the optimized batch (F3) of the formulation was carried out. The culture was used as Staphylococcus aureus and the antimicrobial test was performed using the agar well diffusion. The method used was the Cup plate method and the agar media used is sabouraud dextrose agar result is shown in table.

Table no. 14 Zone of Inhibition Study

Formulation sample.	Observed Zone of inhibition
Marketed formulation	11 ± 1
Optimized batch F3	12.46 ± 1.50



Fig no. 10 zone of inhibition of batch F3

The zone of inhibition for marketed formulation was found to be 11 ± 1 cm and facial cream prepared was found to be 12.46 ± 1.50 cm.

CONCLUSION

The main objective of this study was to formulate an herbal cream. The efficient selection of medicinal plants and their extract with correct concentration with perfect formulation can show a good medicinal effect on the body and it may increase the potency of drugs and formulation. The use of *Curcuma longa* (turmeric) and *Aloe barbadensis miller* (aloe vera) herbal cream shows a facial effect and all these herbal ingredients showed various activities such as antimicrobial, antibacterial, antiviral, Antiprotozoal, Antihelmintic, Anti-diabetic. The herbal cream formulation is stable at room temperature and can be effectively used against various diseases on the skin. The cream containing turmeric and aloe vera was tested for various physiochemical tests and the results were found according to the standard value. Herbal creams are a better substitute than synthetic cream formulations.

REFERENCES

1. Aggarwal BB, Yuan W, Li S, Gupta SC. Curcumin-free turmeric exhibits anti-inflammatory and anticancer activities: Identification of novel components of turmeric. *Mol Nutr Food Res*. 2013 Sep;57(9):1529-42. doi: 10.1002/mnfr.201200838. PMID: 23847103.
2. Surjushe A, Vasani R, Saple DG. Aloe vera: a short review. *Indian J Dermatol*. 2008;53(4):163-166. doi:10.4103/0019-5154.44785.
3. Kaur R, Kapoor S. Aloe vera: A valuable wonder plant for health and beauty. *J Med Plants Stud*. 2013;1(4):32-37.
4. Vaughn AR, Branum A, Sivamani RK. Effects of turmeric (*Curcuma longa*) on skin health: A systematic review of the clinical evidence. *Phytother Res*. 2016 Aug;30(8):1243-64. doi: 10.1002/ptr.5640. Epub 2016 May 23. PMID: 27213821
5. Kohen R. Skin antioxidants their role in aging and insidative stress new approaches
6. Mishra A, Mishra AK, Chattopadhyay P. Herbal Cosmeceuticals for Photoprotection from Ultraviolet B Radiation: A Review. *Tropical Journal of Pharmaceutical Research*. 2018; 17(5):1051-1060.
7. Gupta M, Mazumder UK, Gomathi P, Selvan VT. Anti-oxidative and anti-inflammatory effects of topical curcumin gel in experimental burn wounds in rats. *Pharmaceutical Biology*. 2017; 55(1):2066-2072.
8. Ahmad N, Bhatia S, Ali SS, et al. Topical anti-inflammatory and analgesic activities of standardized pomegranate rind extract in a model of UVB-induced oxidative stress. *Pharmaceutical Biology*. 2018; 56(1):68-77.
9. Khan IU, Akhtar N, Murtaza G, et al. Development and characterization of novel sandalwood oil-loaded nanocarrier-based gel for potential anti-aging application. *Drug Development and Industrial Pharmacy*. 2019; 45(9):1487-1496.
10. Li X, Ma L, Gu Y, et al. Evaluation of the antioxidant and anti-aging activities of emulsions containing flavonoids-loaded silk fibroin nanoparticles. *International Journal of Nanomedicine*. 2019; 14:4121-4132.
11. Adukwu EC, Bowles MH, Edwards-Jones V, Bone H. Antimicrobial activity, cytotoxicity and chemical analysis of lemongrass essential oil (*Cymbopogon flexuosus*) and pure citral. *Applied Microbiology and Biotechnology*. 2016; 100(16):7549-7557.
12. Sharma S, Sharma R, Jain A. Evaluation of anti-inflammatory and anti-aging potential of selected medicinal plants. *International Journal of Cosmetic Science*. 2017; 39(6):620-624.
13. Bano F, Anjum S, Riaz N, et al. In vitro and in vivo antioxidant activities of *Nigella sativa* L. oil and its nanoemulsion. *Journal of Food Science and Technology*. 2019; 56(7):3084-3093.
14. Saeed N, Khan MR, Shabbir M. Antioxidant activity, total phenolic and total flavonoid contents of whole plant extracts *Torilis leptophylla* L. *BMC Complementary and Alternative*

Medicine. 2012; 12:221.

15. Koleva II, van Beek TA, Linssen JP, et al. Screening of plant extracts for antioxidant activity: a comparative study on three testing methods. *Phytochemical Analysis*. 2002; 13(1):8-17.
16. Miguel Lopez I. Anticancer and carcinogenic properties of curcumin: Considerations for its clinical development as a cancer chemopreventive and chemotherapeutic agent.

