ISSN: 2320-2882

IJCRT.ORG



INTERNATIONAL JOURNAL OF CREATIVE RESEARCH THOUGHTS (IJCRT)

An International Open Access, Peer-reviewed, Refereed Journal

VILDAGLIPTIN: A COMPREHENSIVE REVIEW ON ITS PHARMACOLOGICAL, PHARMACEUTICAL AND ANALYTICAL PROFILE

P. Siva Krishna^{1*}, M.M. Eswarudu¹, T. Likhitha¹, N. Venkatesh¹, Ch. Poojitha¹, K. Sujana¹, B. Gopaiah¹, P. Srinivasa Babu²

1. Department of Pharmaceutical Analysis, Vignan Pharmacy College, Vadlamudi, Guntur, 522213, Andhra Pradesh, India.

2. Department of Pharmaceutics, Vignan Pharmacy College, Vadlamudi, Guntur, 522213, Andhra Pradesh,

India.

Corresponding author:

Pusuluri Siva Krishna, Assistant Professor, Department of Pharmaceutical Analysis Vignan Pharmacy College, Vadlamudi, 522213, Andhra Pradesh, India.

ABSTRACT:

This review article presents a comprehensive and in-depth analysis of vildagliptin, a widely prescribed dipeptidyl peptidase-4 (DPP-4) inhibitor for the management of type II diabetes mellitus. The article synthesizes a wealth of information regarding the pharmacological, pharmaceutical, and analytical aspects of vildagliptin. The pharmacological profile encompasses its mechanism of action, clinical efficacy, safety, and potential interactions, offering a well-rounded understanding of its therapeutic impact. Furthermore, the pharmaceutical perspective delves into formulation strategies, dosage forms, and novel delivery methods, shedding light on the diverse approaches taken to optimize drug delivery and patient compliance. In parallel, the review addresses the analytical methods employed for Vildagliptin quantification, exploring advances in chromatographic, spectroscopic, and other techniques that ensure accurate and precise assessment of the drug in various matrices. This comprehensive synthesis of vildagliptin's multifaceted attributes serves as a valuable resource for clinicians, researchers, and pharmaceutical professionals, offering insights that can potentially inform future therapeutic advancements and research endeavours in the realm of diabetes management and drug development.

Keywords: Vildagliptin, Dipeptidyl peptidase-4 (DPP-4) inhibitor, Optimize drug delivery, Chromatography, Spectroscopy.

Introduction:

Vilidagliptin (LAF237) inhibits dipeptidyl peptidase-4 (DPP-4) selectively in the body to control blood sugar levels. A drug that inhibits GLP-1 secretion and insulinotropic effects is prescribed for the management of type II diabetes mellitus. Vildagliptin inhibits DPP-4, which leads to the destruction of glucose-dependent insulinotropic polypeptide (GIP), which is an incretin hormone that stimulates insulin secretion and regulates blood sugar levels. Consequently, GLP-1 and GIP levels are elevated, which leads to improved glycemic control. The risk of hypoglycemia associated with vildagliptin is relatively low in clinical trials. [1]

During 2008, the European Medicines Agency approved oral vildagliptin for treatment of adults with type II diabetes mellitus, either alone or in combination with metformin, sulfonylureas, or thiazolidinediones in patients who did not achieve adequate glycemic control with monotherapy. Galvus is the brand name for this drug. The fixed-dose formulation of vildagliptin, Eucreas, is also available for adults whose glycemic control is not adequate with vildagliptin alone. Vildagliptin is currently under investigation in the US.



Figure 1: Chemical Structure of Vildagliptin Table 1: Drug Profile of Vildagliptin [2]

DRUG	Vildagliptin
IUPAC Name	(2S)-1-{2-[(3-hydroxyadamantan-1-yl) amino] acetyl) pyrrolidine -2- carbonitrile
Chemical Formula	$C_{17}H_{25}N_{3}O_{2}$
Molecular Mass	303.3993 g/mole
Melting Point	153-155°C
Physical State	Solid
Solubility	Soluble in Water and Methanol
рКа	14.71 and 9.03 Strongest acidic and basic respectively
t1/2	90 minutes
Therapeutic Use	Used to reduce hyperglycaemia in type II diabetes mellitus.

Pharmacology

Pharmacodynamics

Pharmacotherapeutic group:

Diabetes medications, dipeptidyl peptidase-4 inhibitors, ATC code: A10BH02 A potent and selective DPP-4 inhibitor, vildagliptin belongs to the islet enhancer class.

Pharmacodynamic effects

In addition to increasing endogenous levels of these incretin hormones, vildagliptin improves glucosedependent insulin secretion by improving beta cell sensitivity. Treatment with vildagliptin 50-100 mg daily in patients with type 2 diabetes significantly improved markers of beta cell function including HOMA- β (Homeostasis Model Assessment– β), proinsulin to insulin ratio and measures of beta cell responsiveness from the frequently-sampled meal tolerance test. In non-diabetic (normal glycaemic) individuals, vildagliptin does not stimulate insulin secretion or reduce glucose levels.

As vildagliptin enhances endogenous GLP-1 levels, it also stimulates the secretion of glucose-sensitive glucagon by alpha cells.

A decrease in fasting and postprandial hepatic glucose production results from an enhanced increase in the insulin/glucagon ratio during hyperglycaemia due to increased incretin hormone levels. The known effect of increased GLP-1 levels delaying gastric emptying is not observed with vildagliptin treatment.

Pharmacokinetics

Absorption

Vildagliptin is rapidly absorbed following oral administration in the fasting state, with peak plasma concentrations observed at 1.7 hours. AUC does not change when food is consumed, but the time to peak plasma concentration is delayed to 2.5 hours. Food-related administration of vildagliptin reduced Cmax (19%). Galvus can be taken with or without food, since the magnitude of the change is not clinically significant. There is an absolute bioavailability of 85%.

Distribution

Vildagliptin is poorly bound to plasma proteins (9.3%), and it is equally distributed between plasma and red blood cells. Upon intravenous administration of vildagliptin, the mean volume of distribution is 71 litres, indicating extravascular distribution.

Biotransformation

Vildagliptin is mainly eliminated by metabolism in humans, accounting for 69% of its dose. A major metabolite of BQS 867, LAY 151, is inactive and is formed during the hydrolysis of the cyano moiety. It accounts for 57% of the dose. The kidney may be an important organ in the hydrolysis of vildagliptin to its major inactive metabolite, LAY151, in vitro in human kidney microsomes. A study in vivo using rats with DPP-4 deficiencies found that DPP-4 contributes to the hydrolysis of vildagliptin. No quantifiable amount of vildagliptin is metabolized by CYP 450 enzymes. Consequently, co-medications that inhibit or induce CYP 450 should not affect the metabolic clearance of vildagliptin. During in vitro studies, vildagliptin was shown

not to inhibit or induce CYP 450 enzymes. The metabolism of co-medications metabolized by CYP 1A2, CYP 2C8, CYP 2C9, CYP 2C19, CYP 2D6, CYP 2E1 or CYP 3A4/5 is unlikely to be affected by vildagliptin. Elimination

A dose of [14C] vildagliptin is excreted by approximately 85% in the urine and 15% in the feces following oral administration. Approximately 23% of the unchanged vildagliptin dose was excreted in the urine after oral administration. Vildagliptin has a total plasma clearance of 41 and a renal clearance of 13 l/h in healthy subjects after intravenous administration. A two-hour elimination half-life is typical after intravenous administration. The elimination half-life after oral administration is approximately 3 hours.

Linearity/non-linearity

As the therapeutic dose range for vildagliptin increased, the C_{max} and the area under the plasma concentration time curves (AUC) increased approximately dose proportionally.

Mechanism of action

It regulates blood glucose levels and maintains glucose homeostasis by acting on two incretin hormones, glucose-dependent insulinotropic peptide (GIP) and glucagon-like peptide-1 (GLP-1). An oral glucose challenge is estimated to stimulate insulin responses through the action of GLP-1 and GIP by more than 70%. The GIP and GLP-1 receptors are G-protein-coupled and activate insulin secretion in a glucose-dependent manner. Additionally, GLP-1 promotes islet neogenesis and differentiation, and attenuates pancreatic betacell apoptosis in addition to its effects on insulin secretion. In type II diabetes mellitus, GLP-1 secretion is impaired, and the insulinotropic effect of GIP is significantly diminished. The incretin hormones also exert extra-pancreatic effects, such as lipogenesis and myocardial function.[1]

Vildagliptin lowers blood glucose by selectively inhibiting dipeptidyl peptidase-4 (DPP-4), an enzyme responsible for truncating and inactivating GLP-1 and GIP upon their release from intestinal cells. After the second amino acid from the N-terminal end, DPP-4 cleaves oligopeptides. As a result of the inhibition of DPP-4, GLP-1 and GIP half-lives are prolonged, increasing levels of active circulating incretin hormones.3 Vildagliptin inhibits DPP-4 for a dose-dependent duration.5 Vildagliptin reduces fasting and prandial glucose and HbA1c levels. It enhances the glucose sensitivity of alpha- and beta-cells and augments glucosedependent insulin secretion. Fasting and postprandial glucose levels are decreased, and postprandial lipid and lipoprotein metabolism are also improved [3]. Mechanism of action of Vildagliptin was shown in Figure 2



Figure 2: Mechanism of action of Vildagliptin [6]

Toxicity

The oral Lowest published toxic dose (TDLO) is 0.3 mg/kg in rats and 1 mg/kg in mice. [5]

There is limited information regarding overdose with vildagliptin. In one study, patients experienced muscle pain, mild and transient paresthesia, fever, edema, and a transient increase in lipase levels at a dose of 400 mg. At 600 mg, one subject experienced edema of the feet and hands and increases in creatine phosphokinase (CPK), aspartate aminotransferase (AST), C-reactive protein (CRP) and myoglobin levels. Supportive management is recommended in case of an overdose. There is no known antidote, and vildagliptin and its major metabolite cannot be removed via hemodialysis. [4]

Description of selected adverse reactions

Hepatic impairment

Rare cases of hepatic dysfunction (including hepatitis) have been reported. In these cases, the patients were generally asymptomatic without clinical sequelae and liver function returned to normal after discontinuation of treatment. In data from controlled monotherapy and add-on therapy trials of up to 24 weeks in duration, the incidence of ALT or AST elevations \Box 3x ULN (classified as present on at least 2 consecutive measurements or at the final on-treatment visit) was 0.2%, 0.3% and 0.2% for vildagliptin 50 mg once daily, vildagliptin 50 mg twice daily and all comparators, respectively. These elevations in transaminases were generally asymptomatic, non-progressive in nature and not associated with cholestasis or jaundice.

Angioedema

Rare cases of angioedema have been reported on vildagliptin at a similar rate to controls. A greater proportion of cases were reported when vildagliptin was administered in combination with an angiotensin converting enzyme inhibitor (ACE-Inhibitor). The majority of events were mild in severity and resolved with ongoing vildagliptin treatment.

Hypoglycaemia

Hypoglycaemia was uncommon when vildagliptin (0.4%) was used as monotherapy in comparative controlled monotherapy studies with an active comparator or placebo (0.2%). No severe or serious events of hypoglycaemia were reported. When used as add-on to metformin, hypoglycaemia occurred in 1% of vildagliptin-treated patients and in 0.4% of placebo-treated patients. When pioglitazone was added, hypoglycaemia occurred in 0.6% of vildagliptin-treated patients and in 1.9% of placebo-treated patients. When sulphonylurea was added, hypoglycaemia occurred in 1.2% of vildagliptin treated patients and in 0.6% of placebo-treated patients. When sulphonylurea and metformin were added, hypoglycaemia occurred in 5.1% of vildagliptin treated patients and in 1.9% of placebo treated patients. In patients taking vildagliptin in combination with insulin, the incidence of hypoglycaemia was 14% for vildagliptin and 16% for placebo.

Overdose

Symptoms

Information on the likely symptoms of overdose was taken from a rising dose tolerability study in healthy subjects given Galvus for 10 days. At 400 mg, there were three cases of muscle pain, and individual cases of mild and transient paraesthesia, fever, oedema and a transient increase in lipase levels. At 600 mg, one subject experienced oedema of the feet and hands, and increases in creatine phosphokinase (CPK), aspartate aminotransferase (AST), C-reactive protein (CRP) and myoglobin levels. Three other subjects experienced oedema of the feet, with paraesthesia in two cases. All symptoms and laboratory abnormalities resolved without treatment after discontinuation of the study medicinal product.

Table 1: Adverse reactions reported in patients who received vildagliptin as monotherapy or as addon therapy in controlled clinical studies and in post-marketing experience. [4]

System organ class - adverse reaction	Frequency				
Infections and infestations					
Nasopharyngitis	Very common				
Upper respiratory tract infection	Common				
Metabolism and nutrition disorders					
Hypoglycaemia	Uncommon				
Nervous system disorders					
Dizziness	Common				
Headache	Common				
Tremor	Common				
Eye disorders					
Vision blurred	Common				
Gastrointestinal disorders					
Constipation	Common				
Nausea	Common				
Gastro-oesophageal reflux disease	Common				
Diarrhoea	Common				
Abdominal pain, including upper	Common				
Vomiting	Common				
Flatulence	Uncommon				
Pancreatitis	Rare				
Hepatobiliary disorders					
Hepatitis	Not known*				
Skin and subcutaneous tissue disorders					
Hyperhidrosis	Common				
Rash	Common				
Pruritis	Common				
Dermatitis	Common				
Urticaria	Uncommon				

IJCRT2308472 International Journal of Creative Research Thoughts (IJCRT) www.ijcrt.org e382

Exfoliative and bullous skin lesions, including	Not known*
bullous pemphigoid	
Cutaneous vasculitis	Not known*
Musculoskeletal and connective tissue disorders	
Arthralgia	Common
Myalgia	Common
Reproductive system and breast disorders	
Erectile dysfunction	Uncommon
General disorders and administration site conditi	ons
Asthenia	Common
Oedema peripheral	Common
Fatigue	Uncommon
Chills	Uncommon
Investigations	
Abnormal liver function tests	Uncommon
Weight increase	Uncommon
* Based on post-marketing experience.	

Management

In the event of an overdose, supportive management is recommended. Vildagliptin cannot be removed by haemodialysis. However, the major hydrolysis metabolite (LAY 151) can be removed by haemodialysis.

Table 2: Available marketed formulations of Vildagliptin

Name	Dosage form	Strength	Route	Manufacturer
Galvus	Tablet	50 mg	Oral	Novartis India Ltd.
Jalra	Tablet	50 mg	Oral	USV Ltd.
Xiliarx	Tablet	50 mg	Oral	European Medicines Agency
Gliptagreat	Tablet	50 mg	Oral	Mankind Pharma Ltd
Vildazem	Tablet	50 mg	Oral	Zeelab Pharmacy Pvt Ltd
Zomelis	Tablet	50 mg	Oral	Eris Lifesciences Ltd

Table 3: List of Analytical methods available for Vildagliptin estimation

S. No.		Parameters		Results		
	Method: UV					
			Linearity range	2-32 μg/mL		
			λ_{max}	268 nm		
		Shimeday UV 1800 double been	accuracy	97.78%		
1[7]	System	spectrophotometer	\mathbb{R}^2	0.9997		
			%RSD	1.27573%		
			LOD	1.46053 μg/mL		
			LOQ	3.46748 µg/mL		
			Linearity	$5.20 \mu g/mI$		
				range	5-50 μg/mL	
2101	System	UV-Vis double beam	\mathbb{R}^2	0.9987		
2[0]	System	spectrophotometer	LOD	2.54 μg/mL		
			LOQ	4.69 μg/mL		
			% RSD	0.85%		

www.ijcrt.org

$\begin{array}{ c c c c c c c c c c c c c c c c c c c$		System	Shimadzu 1800 spectronic UV-Visible		
$\begin{array}{ c c c c c c c c c c c c c c c c c c c$		bystem	corporation , japan)		
$[11] \begin{tabular}{ c c c c c c c c c c c c c c c c c c c$	3[9]	λ_{max}	218.25 nm		
[11] [7[13] [7[13] [7[13] [7]] [7[13] [7]] [7[13] [7]] [7[13] [7]] [7[13] [7]] [7[13] [7]] [7[13] [7]] [7[13] [7]] [7[13] [7]] [7[13] [7]] [7[13] [7]] [7[13] [7]] [7[13] [7]] [7[13] [7]] [7[13] [7]] [7[13] [7]] [7[13] [7]] [7[13] [7]] [7[13] [7]] [7]] [7[13] [7]] [7[13] [7]] [7[13] [7]] [7[13] [7]] [7]] [7[13] [7]] [7[13] [7]] [7]] [7[13] [7]] [7] [7] [7] [7] [7] [7] [7] [7] [Linearity	(0.100 / J		
$[13] \begin{tabular}{ c c c c c c c } \hline R^2 & 0.998 \\ \hline UV-Visible spectrophotometer (Shimadzu UV-1800 spectrophotometer, Shimadzu, Japan) \\ \hline UV-Vis spectrophotometer, Shimadzu, Japan) \\ \hline UV-Vis spectrophotometer 1600 (shimadzu, japan) \\ \hline UD 0.0272 µg/mL \\ \hline VO 0.023 µg/mL \\ \hline UD 0.025 µg/mL \\ \hline VItraviolet visible (UVVIS) \\ spectrophotometer (Shimadzu model 18001 (Shimadzu, Kyoto, Japan)) \\ \hline UD 0.055 µg/mL \\ \hline VItraviolet visible (UVVIS) \\ \hline VItraviolet visible (UVVIS) \\ spectrophotometer (Shimadzu model 18001 (Shimadzu, Kyoto, Japan)) \\ \hline UD 0.055 µg/mL \\ \hline VItraviolet visible (UVVIS) $		range	60-100 μg/mL		
		\mathbb{R}^2	0.998		
$\begin{array}{ c c c c c c c c c c c c c c c c c c c$				Linearity range	8-32 μg/mL
$\begin{array}{c c c c c c c c c c c c c c c c c c c $			UV-Visible spectrophotometer	Wavelength	197 nm
$[13] System = \frac{1}{2} (1) Sy$	4[10]	System	(Shimadzu UV-1800	\mathbb{R}^2	0.999
$[11] System = \frac{1}{10000000000000000000000000000000000$.[10]	System	spectrophotometer, Shimadzu, Japan)	LOD	0.247 μg/mL
[11] System UV-Vis spectrophotometer 1600 (shimadzu, japan) UDD 0.272 µg/mL 207.2-230.6 Wavelength 200.6 Wavelength 200.6 Wavelength 200.6 Wavelength 210.0 0.272 µg/mL % RSD <+ 2% UI concentration 0.7-1.0 µg/mL Wavelength 217 nm R2 0.999 LOD 0.023 µg/mL UI traviolet visible (UVVIS) spectrophotometer (Shimadzu model 18001 (Shimadzu, Kyoto, Japan)) Wavelength 200.2 mg/mL LOQ 0.225 µg/mL Wavelength 200.5 nm R2 0.999 LOD 0.0055 µg/mL UOD 0.055 µg/mL UOD 0.055 µg/mL UOD 0.0166 µg/mL Wavelength 200.5 nm R2 0.999 LOD 0.0166 µg/mL Wavelength 200.5 nm R2 0.999 LOD 0.0166 µg/mL Wavelength 200.2 mg/mL Spectrophotometer (Shimadzu, Kyoto, Japan)) How Concentration R2 0.999 LOD 0.0166 µg/mL Wavelength 200.2 mg/mL Spectrophotometer (Shimadzu, Kyoto, Japan)) How Concentration R2 0.999 LOD 0.025 µg/mL Spectrophotometer (Shimadzu, Kyoto, Japan)) How Concentration R2 0.999 LOD 0.0166 µg/mL Spectrophotometer (Shimadzu, Kyoto, Japan)) How Concentration R2 0.999 LOD 0.0166 µg/mL Spectrophotometer (Shimadzu, Kyoto, Japan)) How Concentration R2 0.999 LOD 0.0166 µg/mL Spectrophotometer (Shimadzu, Kyoto, Japan)) How Concentration R2 0.999 LOD 0.0166 µg/mL Spectrophotometer (Shimadzu, Kyoto, Japan) How Concentration R2 0.999 LOD 0.0166 µg/mL Spectrophotometer (Shimadzu, Kyoto, Japan) How Concentration R2 0.999 LOD 0.0166 µg/mL Spectrophotometer (Shimadzu, Kyoto, Japan) How Concentration R2 0.999 LOD 0.0166 µg/mL Spectrophotometer (Shimadzu, Kyoto, Japan) How Concentration R2 0.999 LOD 0.0166 µg/mL Spectrophotometer (Shimadzu, Kyoto, Japan) How Concentration R2 0.999 LOD 0.0166 µg/mL Spectrophotometer (Shimadzu Kyoto, Japan) How Concentration R2 0.999 LOD 0.016				LOQ	0.748 μg/mL
[11] System UV-Vis spectrophotometer 1600 (shimadzu, japan) UV-Vis spectrophotometer UC UV (S) (Simadzu, japan) UV-Vis spectrophotometer (Shimadzu model 18001 (Shimadzu, Kyoto, Japan)) UV-Vis Spectrophotometer (Shimadzu, Kyoto, Japan) UV-Vis Spectrophotometer (Shimadzu, Kyoto, Japan) UV-Vis Spectrophotometer (Shimadzu model 18001 (Shimadzu, Kyoto, Japan)) UV-Vis Spectrophotometer (Shimadzu Mise Spectrophotometer (Shimadzu Mise Spectrophotometer (Shimadzu Mise Spectrophotometer (Shimadzu Mise Spectrophotomete				% RSD	< 2%
[11] System UV-Vis spectrophotometer 1600 (shimadzu, japan) (Simadzu, ja				Linearity range	1-60 μg/mL
$\begin{array}{ c c c c c c c c c c c c c c c c c c c$				R ²	≥ 0.999
[11] System (shimadzu, japan) = 0 - Nm - 10D 0.272 µg/mL - 10Q 0.827 µg/mL - 10Q 0.225 µg/mL - 10Q 0.255 µg/mL - 10Q 0.166 µg/mL - 10Q 0.63 µ	5[11]	Grantana	UV-Vis spectrophotometer 1600	Wavelength	207.2-230.6
[12] System LABINDIA spectrophotometer [13] System LABINDIA spectrophotometer (Shimadzu model 18001 (Shimadzu, Kyoto, Japan)) [14] System System Agilent liquid chromatography (Santa clara,CA) [15] System Agilent liquid chromatography (Santa clara,CA) [16] System Column (130x4.6 mn 1.d., 5 µm particle size) (130x4.6 mn 1.d.,	2[11]	System	(shimadzu, japan)		$\frac{\text{Nm}}{0.272 \text{ m} \text{ m}}$
$[12] System = LABINDIA spectrophotometer \\ [12] System = LABINDIA spectrophotometer \\ [13] System = LABINDIA spectrophotometer \\ [14] System = LABINDIA spectrophotometer \\ [15] System = LABINDIA spectrophotometer \\ [15] LABINDIA spectrophotometer \\ [15] LABINDIA spectrophotometer \\ [15] System = LABINDIA spectrophotometer \\ [15] Ultraviolet visible (UVVIS) \\ [15] System = LABINDIA spectrophotometer \\ [15] System = LABINDIA spectrophotometer \\ [15] Ultraviolet visible (UVVIS) \\ [15] System = LABINDIA spectrophotometer \\ [15] Ultraviolet visible (UVVIS) \\ [15] System = LABINDIA spectrophotometer \\ [15] Ultraviolet visible (UVVIS) \\ [16] System = LABINDIA spectrophotometer \\ [16] Ultraviolet visible (UVVIS) \\ [16] System = LABINDIA spectrophotometer \\ [16] Ultraviolet visible (UVVIS) \\ [16] System = LABINDIA spectrophotometer \\ [16] Ultraviolet visible (UVVIS) \\ [16] System = LABINDIA spectrophotometer \\ [16] Ultraviolet visible (UVVIS) \\ [16] System = LABINDIA spectrophotometer \\ [16] Ultraviolet visible (UVVIS) \\ [16] System = LABINDIA spectrophotometer \\ [16] Ultraviolet visible (UVVIS) \\ [16] Ultraviolet visible (Ultraviolet visible (UVVIS) \\ [16] Ultraviolet visible (Ultraviolet visible (Ultraviolet visible (Ultraviolet visible (Ultraviolet visible (Ultraviolet visible (Ultraviolet$					$0.272 \mu g/mL$
$[12] System LABINDIA spectrophotometer \\ [12] System LABINDIA spectrophotometer \\ [13] System LABINDIA spectrophotometer \\ [14] System LABINDIA spectrophotometer \\ [15] System LABINDIA spectrophotometer \\ [15] LABINDIA spectrophotometer \\ [15] LABINDIA spectrophotometer \\ [15] System LABINDIA spectrophotometer \\ [15] System LABINDIA spectrophotometer \\ [15] System LABINDIA spectrophotometer \\ [15] Ultraviolet visible (UVVIS) \\ spectrophotometer (Shimadzu model 18001 (Shimadzu, Kyoto, Japan)) \\ [15] System LIC \\ [15] System LIC$				<u> </u>	$\frac{0.627 \mu g/mL}{< + 2\%}$
[12] System LABINDIA spectrophotometer (Shimadzu model) [12] System LABINDIA spectrophotometer (Shimadzu model) [13] System Ultraviolet visible (UVVIS) spectrophotometer (Shimadzu model 18001 (Shimadzu, Kyoto, Japan)) [14] System VITraviolet visible (UVVIS) spectrophotometer (Shimadzu model 18001 (Shimadzu, Kyoto, Japan)) [14] System VITraviolet visible (UVVIS) spectrophotometer (Shimadzu model 18001 (Shimadzu, Kyoto, Japan)) [15] VITraviolet visible (UVVIS) spectrophotometer (Shimadzu model 18001 (Shimadzu, Kyoto, Japan)) [15] VITraviolet visible (UVVIS) spectrophotometer (Shimadzu model 18001 (Shimadzu, Kyoto, Japan)] [15] VITraviolet visible (UVVIS) spectrophotometer (Shimadzu model 18001 (Shimadzu, Kyoto, Japan)] [15] VITraviolet visible (UVVIS) spectrophotometer (Shimadzu model 18001 (Shimadzu, Kyoto, Japan)] [15] VITraviolet visible (UVVIS) spectrophotometer (Shimadzu model 18001 (Shimadzu, Kyoto, Japan)] [15] VITraviolet visible (UVVIS) spectrophotometer (Shimadzu model 18001 (Shimadzu, Kyoto, Japan)] [15] VITraviolet visible (UVVIS) spectrophotometer (Shimadzu model 18001 (Shimadzu, Kyoto, Japan)] [15] VITraviolet visible (UVVIS) spectrophotometer (Shimadzu model 18001 (Shimadzu, Kyoto, Japan)] [15] VITraviolet visible (UVVIS) spectrophotometer (Shimadzu model 18001 (Shimadzu, Kyoto, Japan)] [15] VITraviolet visible (UVVIS) [15] VITraviolet visible (UVVIS) [15] VITraviolet visible (UVVIS) [15] VITraviolet visible (UVVIS] [16] VITraviolet visible (UVVIS) [16] VITraviolet visible (UVVIS] [16] VITraviolet visible (UVVI				70 KSD	$\frac{12}{0.35-1.05}$
[12] System LABINDIA spectrophotometer [13] System LABINDIA spectrophotometer [14] LABINDIA spectrophotometer [14] LABINDIA spectrophotometer [15] System LABINDIA spectrophotometer [16] LABINDIA spectrophotometer [16] LABINDIA spectrophotometer [16] Concentratioon [17] R2 0.999 [16] LOD 0.023 µg/mL [16] LOQ 0.225 µg/mL [16] LOQ 0.225 µg/mL [16] LOQ 0.225 µg/mL [16] LOQ 0.225 µg/mL [16] LOQ 0.055 µg/mL [16] LOQ 0.055 µg/mL [16] LOQ 0.166 µg/mL [16] LOQ 0.61% [16] LOD 0.63 µg/mL [16] LOQ 0.63 µg/mL [1				Linearity range	0.55-1.05
$\begin{array}{c c c c c c c c c c c c c c c c c c c $				Concentratioon	μginn
$\begin{array}{c c c c c c c c c c c c c c c c c c c $		System		range	0.7-1.0 μg/mL
R20.999IOD0.023 μ g/mLIOD0.023 μ g/mLLOQ0.225 μ g/mL% RSD< 2%	6[12]		LABINDIA spectrophotometer	Wavelength	217 nm
$[14] \begin{tabular}{ c c c c c c c } \hline LOD & 0.023 \ \mu g/mL \\ \hline LOQ & 0.225 \ \mu g/mL \\ \hline LOQ & 0.225 \ \mu g/mL \\ \hline Ultraviolet visible (UVVIS) \\ spectrophotometer (Shimadzu model 18001 (Shimadzu, Kyoto, Japan)) & R^2 & 0.999 \\ \hline Ultraviolet visible (UVVIS) \\ spectrophotometer (Shimadzu model 18001 (Shimadzu, Kyoto, Japan)) & R^2 & 0.999 \\ \hline UOD & 0.055 \ \mu g/mL \\ \hline LOQ & 0.166 \ \mu g/mL \\ \hline UQ & 0.166 \ \mu g/mL \\ \hline V & V & RSD & <2\% \\ \hline \hline & V & V & V & V & V \\ \hline & V & V & V & V & V & V \\ \hline & V & V & V & V & V & V \\ \hline & V & V & V & V & V & V \\ \hline & V & V & V & V & V & V \\ \hline & V & V & V & V & V & V \\ \hline & V & V & V & V & V & V \\ \hline & V & V & V & V & V & V \\ \hline & V & V & V & V & V & V \\ \hline & V & V & V & V & V & V \\ \hline & V & V & V & V & V & V \\ \hline & V & V & V & V & V & V \\ \hline & V & V & V & V & V & V \\ \hline & V & V & V & V & V & V \\ \hline & V & V & V & V & V & V \\ \hline & V & V & V & V & V & V \\ \hline & V & V & V & V & V \\ \hline & V & V & V & V & V \\ \hline & V & V & V & V & V \\ \hline & V & V & V & V & V \\ \hline & V & V & V & V & V \\ \hline & V & V & V & V & V \\ \hline & V & V & V & V & V \\ \hline & V & V & V & V & V \\ \hline & V & V & V & V & V \\ \hline & V & V & V & V & V \\ \hline & V & V & V & V & V \\ \hline & V & V & V & V$				R^2	0.999
Image: Note of the sector o				LOD	0.023 μg/mL
7[13]System $(2%)$ System $(2%)$ System $(2%)$ Ultraviolet visible (UVVIS) spectrophotometer (Shimadzu model 18001 (Shimadzu, Kyoto, Japan)) (1) (100) (1) $(202.5 nm)$ $(202.5 nm)$ $(202.80 µg/mL)$ $(202.80 µg/mL)$ 8[14]Nobile phase Nobile phaseN Brdge analytical column of triethylamine 0.3% adjusted to pH 7.0 with phsphoric acid (15:85; v/v)N DO8[14]Nobile phase Nobile phaseN Dom Nobile phaseN Dom Nobile phaseN Dom Nobile phase <tr< td=""><td></td><td></td><td></td><td>LOQ</td><td>0.225 μg/mL</td></tr<>				LOQ	0.225 μg/mL
T[13]Linearity range $10-40 \ \mu g/mL$ 7[13]SystemUltraviolet visible (UVVIS) spectrophotometer (Shimadzu model 18001 (Shimadzu, Kyoto, Japan)) R^2 0.999 LOD $0.055 \ \mu g/mL$ LOQ $0.166 \ \mu g/mL$ VMethod: LC $V \otimes RSD$ $<2\%$ Method: LCColumnAgilent liquid chromatography (Santa clara,CA)Linearity $20-80 \ \mu g/mL$ ColumnX Brdge analytical column C8 (130x4.6 mm I.d., 5 μ m particle size) (waters)Retention time $6.2 \ min$ 8[14]Column temperatureRoom temperature (23 ± 1^0 C) $\%$ RSD 0.61% 8[14]Mobile phaseAcetonitrile and a solution of triethylamine 0.3% adjusted to pH 7.0 with phsphoric acid ($15:85; v/v$)LOD $0.63 \ \mu g/mL$				% RSD	< 2%
7[13]Ultraviolet visible (UVVIS) spectrophotometer (Shimadzu model 18001 (Shimadzu, Kyoto, Japan))Wavelength202.5 nm7[13]SystemSpectrophotometer (Shimadzu model 18001 (Shimadzu, Kyoto, Japan)) R^2 0.999LOD0.055 µg/mLLOQ0.166 µg/mL% RSD<2%				Linearity range	_ 10-40 μg/mL
7[13]SystemSupertrophotometer (Shimadzu model 18001 (Shimadzu, Kyoto, Japan)) R^2 0.99910D0.055 µg/mL10Q0.166 µg/mL10Q0.164 µg/mL <t< td=""><td></td><td rowspan="3">System</td><td>Ultraviolet visible (UVVIS)</td><td>Wavelength</td><td>202.5 nm</td></t<>		System	Ultraviolet visible (UVVIS)	Wavelength	202.5 nm
N(10) Dystem Inprovingention (change and index) 18001 (Shimadzu, Kyoto, Japan)) 10D 0.055 µg/mL LOQ 0.166 µg/mL V Nethod: LC Method: LC X Brdge analytical column C8 (130x4.6 mm I.d., 5 µm particle size) (waters) Linearity 20-80 µg/mL Column temperature Retention time 6.2 min Mobile phase Acetonitrile and a solution of triethylamine 0.3% adjusted to pH 7.0 with phsphoric acid (15:85; v/v) LOD 0.63 µg/mL Flow rate 1.0 mL/min LOQ 2.82 µg/mL	7[13]		spectrophotometer (Shimadzu model	\mathbb{R}^2	0.999
Method: LC LOQ 0.166 μg/mL Method: LC Method: LC System Agilent liquid chromatography (Santa clara,CA) Linearity 20-80 μg/mL Column X Brdge analytical column C8 (130x4.6 mm I.d., 5 μm particle size) (waters) Retention time 6.2 min 8[14] Column temperature Room temperature (23± 1°C) %RSD 0.61% Mobile phase Acetonitrile and a solution of triethylamine 0.3% adjusted to pH 7.0 with phsphoric acid (15:85; v/v) LOD 0.63 μg/mL Flow rate 1.0 mL/min LOQ 2.82 μg/mL	,[10]		18001 (Shimadzu, Kyoto, Japan))	LOD	0.055 μg/mL
Method: LC Method: LC System Agilent liquid chromatography (Santa clara,CA) Linearity 20-80 μg/mL Column X Brdge analytical column C8 (130x4.6 mm I.d., 5 μm particle size) (waters) Retention time 6.2 min Column Room temperature (23± 1°C) %RSD 0.61% 8[14] Mobile phase Acetonitrile and a solution of triethylamine 0.3% adjusted to pH 7.0 with phsphoric acid (15:85; v/v) LOD 0.63 μg/mL					0.166 μg/mL
System Agilent liquid chromatography (Santa clara,CA) Linearity 20-80 μg/mL Column X Brdge analytical column C8 (130x4.6 mm I.d., 5 μm particle size) (waters) Retention time 6.2 min Column Room temperature (23± 1°C) %RSD 0.61% 8[14] Mobile phase Acetonitrile and a solution of triethylamine 0.3% adjusted to pH 7.0 with phsphoric acid (15:85; v/v) LOD 0.63 μg/mL			Mallic	% RSD	<2%
SystemAgilent liquid chromatography (Santa clara,CA)Linearity20-80 μg/mLX Brdge analytical column C8 (130x4.6 mm I.d., 5 μm particle size) (waters)Retention time6.2 minColumn temperatureRoom temperature (23± 1°C)%RSD0.61%8[14]Acetonitrile and a solution of triethylamine 0.3% adjusted to pH 7.0 with phsphoric acid (15:85; v/v)LOD0.63 μg/mL			Method: LC		
8[14] Ketention time Ketention time Ketention time Ketention time 8[14] Ketention time Ketention time Ketention time Ketention time 8[14] Ketention time Ketention time Ketention time Ketention time Ketention Ketention time Ketention time Ketention time Ketention time Ketention Ketention time Ketention time Ketention time Ketention time Ketention Ketention time Ketention time Ketention time Ketention time Ketention Ketention time Ketention time Ketention time Ketention time Ketention Ketention Ketention time Ketention Ketention Ketention Ketention Ketention Ketention Ketention Ketention<		System	Agilent liquid chromatography (Santa	Linearity	20-80 μg/mL
8[14] X Brdge analytical column C8 (130x4.6 mm I.d., 5 µm particle size) (waters) Retention time 6.2 min 8[14] Column temperature Room temperature (23± 1°C) %RSD 0.61% 8[14] Acetonitrile and a solution of triethylamine 0.3% adjusted to pH 7.0 with phsphoric acid (15:85; v/v) LOD 0.63 µg/mL Flow rate 1.0 mL/min LOQ 2.82 µg/mL		2	clara,CA)	2	10
Column(130x4.6 mm I.d., 5 μm particle size) (waters)Retention time6.2 minColumn temperatureRoom temperature (23± 1°C)%RSD0.61%8[14]Acetonitrile and a solution of triethylamine 0.3% adjusted to pH 7.0 with phsphoric acid (15:85; v/v)LOD0.63 μg/mLFlow rate1.0 mL/minLOQ2.82 μg/mL			X Brdge analytical column C8		
8[14] Column temperature Room temperature (23±1°C) %RSD 0.61% 8[14] Mobile phase Acetonitrile and a solution of triethylamine 0.3% adjusted to pH 7.0 with phsphoric acid (15:85; v/v) LOD 0.63 µg/mL Flow rate 1.0 mL/min LOQ 2.82 µg/mL		Column	(130x4.6 mm I.d., 5 µm particle size)	Retention time	6.2 min
Column temperature Room temperature (23± 1°C) %RSD 0.61% 8[14] Mobile phase Acetonitrile and a solution of triethylamine 0.3% adjusted to pH 7.0 with phsphoric acid (15:85; v/v) LOD 0.63 µg/mL Flow rate 1.0 mL/min LOQ 2.82 µg/mL			(waters)		
8[14] Acetonitrile and a solution of triethylamine 0.3% adjusted to pH 7.0 with phsphoric acid (15:85; v/v) LOD 0.63 μg/mL Flow rate 1.0 mL/min LOQ 2.82 μg/mL		Column	Room temperature (23 ± 1^{0} C)	%RSD	0.61%
8[14] Mobile phase Acctonitrile and a solution of triethylamine 0.3% adjusted to pH 7.0 with phyloric acid (15:85; v/v) LOD 0.63 μg/mL Flow rate 1.0 mL /min LOQ 2.82 μg/mL		temperature			
Flow rate 1.0 mL/min LOQ 2.82 μg/mL	8[14]	Mahila nhaaa	Acetonitrile and a solution of triathylamina 0.29 / adjusted to pU 7.0		
Flow rate 1.0 mL/min LOQ 2.82 μg/mL		Moone phase	with physical and $(15.85, y/y)$	LOD	0.63 μg/mL
Flow rate 1.0 mL/min LOQ 2.82 μg/mL			with physicine acta $(15.85, \sqrt{3})$		
Inioritan		Flow rate	1.0 mL/min	LOO	2.82 µg/mL
Intection		Injection			2.02 pg/mL
volume 20 µL		volume	20 µL		
Detector Photo diode array detector		Detector	Photo diode array detector		
Detection		Detection	207		
wavelength 207 nm		wavelength	207 nm		

	System	Liquid Chromatography System	Retention time	Vildagliptin- 5.41 min
	Column	Thermo Hypersil ODS C18 column (5 μm, 4.6x250 mm)	%RSD	1.32 and 1.53%
9[15]	Mobile phase	0.1M phosphate buffer (pH 3.5), Acetonitrile and methanol (65%:30%:5% v/v)	Linearity range	2.5-7.5 μg/mL
	Flow rate	0.8 mL/min)	R ²	0.9903
	Detector	PC 220 UV/visible Detector	LOD	0.13 μg/mL
	Detection wavelength	212 nm	LOQ	0.13 μg/mL
	Column	XBridge C8 (150x4.6 mm, 5 µm)		
10[16]	Mobile phase	acetonitriletriethylamine 0.3%, pH 7.0 (15:85	Linearity range	20-80 µg/mL
10[10]	Flow rate	1.0 ml/min		
	Detection	LIV 207 nm	LOD	0.63 µg/mL
	wavelength	0 7 207 1111	LOQ	2.82 μg/mL
		Method: HPLC	- · · ·	
	System	Shimadzu model LC2010CHT HPLC	Linearity range	2-40 Ppm
		system	\mathbb{R}^2	0.9997
11[17]		Phenomenex Luna C18 (2) column	LOD	0.715 Ppm
	Column	(4.6mmx 250mm, 5µ)	LOQ	2.166 Ppm
	Wavelength	219.6 nm	% RSD	0.12%
	System	Shimadzu LC-20AT HPLC manual system	Linearity range	5-25 μg/mL
	Column	Phenomenex Luna C18 (250 x 4.6 mm, 5 μm)	Theoretical plates	2241.76 Plates
	Column temperature	25 ⁰ C	Retention time	1.37 min
12[18]	Mobile phase	70:30 % v/v methanol: acetate buffer (adjusted to pH 5.6 using OPA)	R ²	0.9954
	Flow rate	1 mL/min	LOD	1.05 µg/mL
	Injection volume	20 µL	LOQ	3.90 µg/mL
	Detector	PDA detector	% RSD	0.22%
	Detection	210 nm		
	wavelength	210 1111		
	System	Waters HPLC	Linearity range	50-90 μg/mL
	Column	<u>C18 (4.6 x 150mm, 5mm)</u>	R²	0.999
13[19]	Mobile phase	pH 8.2 buffer, acetonitrile and methanol	LOD	2.98 g/mL
	Flow rate	0.5 mL/min	LOQ	9.94g/mL
	Injection volume	10 µL	% RSD	>2%
	Detection	UV detection at 254 nm		
	wavelength			
	Run time	10 min		

14[20]	System	 Shimadzu chromatographic system Jasco 2000 chromatographic system 	Linearity range	10-100 μg/mL
	Column	C18 column (4.6 \times 150 mm id., particle size 5 μ m)	R ²	0.999
	Mobile phase	10 mM phosphate buffer (pH 4.6) and acetonitrile (85 : 15, v/v)	LOD	l μg/mL
	Flow rate	1.0 mL / min	LOQ	3.2 μg/mL
	Detector	 SPDM20A detector (PDA) UV2070/2075 UVVis detector 	% RSD	2.25%
	System	Dionex Ultimate 3000 System used	Linearity range	1.0 mg/mL
	Column	C18-WP, 100A ⁰ , (250 mm×4.6 mm),5µm particle size column	Theoretical plates	13200
	Mobile phase Column temperature	A buffer solution and methanol (90:10) B methanol, degassed 40 ⁰ C	R ²	>0.99
15[21]	Column temperature Mobile phase	40 ^o C A buffer solution and methanol (90:10) B methanol, degassed	% RSD	< 1.5%
	Flow rate	1 mL/min	LOD	0.018 µg/mL
	Injection volume	100 µL	LOQ	0.066 µg/mL
	Detection	208 nm		
	wavelength	208 IIII		
	System	Waters pump HPLC system	Linearity range	5-25 μg/mL
	Column	Lichrocart C18 column (250 x 4.60 x 5µm)	Retention rate	6.64 min
16[2 <mark>2]</mark>	Mobile phase	0.05 M KH2PO4 : Acetonitrile (70:30 v/v, pH 3.5 with Ortho Phosphoric Acid)	Theoretical plates	1.97 Plates
	Flow rate	1.0 mL/min	\mathbb{R}^2	0.999
	Detector	UV-Visible detector	% RSD	0.057
	Detection wavelength	215 nm	P	
	System	HPLC machine (Agilent Technologies 1200 series)	Linearity range	10-60 µg/mL
	Column	ZORBAX Rapid Resolution HT C18 columns (150 mm x 4.6 mm)	Retention time	5.017 min
	Column	30^{0} C	Theoretical plates	5790 Plates
17[23]	temperatureMobile phase	Buffer: Acetonitrile in the ratio of 50.50 ($y(y)$)	R ²	0.9996
	Flow rate	1.0 mL/min	LOD	0.025.ug/mI
	Injection		LOD	0.025 µg/IIIL
	volume	20 µL	LOQ	0.054 μg/mL
	Detection wavelength	UV detector at 220 nm	%RSD	0.68 %
	Run time	10 min		
	System	Waters Alliance e2695	Linearity range	$0.2-1\overline{00 \ \mu g/mL}$
18[24]	Column	Xbridge BEH C18 column (5 μm, 4.6 × 250 mm) (Waters® Corporation, Milford, MA, USA)	Retention time	4.5 min

	Mobile phase	acetonitrile and monopotassium phosphate buffer (1.36 g/L) 49:51 (v/v)	LOD	0.08 µg/mL
	Flow rate	1.2 mL/min	\mathbb{R}^2	>0.999
	Injection	20 µL		
	Detector	photodiode array detector (model 2998) (Waters Corporation, Mildford, MA, USA)		
	Detection wavelength	236 and 297 nm		
	System	Shimadzu Corporation (LC-2010C HT) model	Linear range	10-1000 ng/mL
	Column	C18 column	R ²	0.9992
19[25]	Mobile phase	A 50 mM ammonium bicarbonate (pH 7.8) B 100% acetonitrile	LOD	10 ng/mL
	Injection volume	20 µL		
	Detection	UV detector 210 nm		
	wavelength			
	Run time	20 min		
	System	Shimadzu (Tokyo, Japan)	Linearity range	2-10 μg/mL
	Column	Onyx C18 Monolithic column (100mm× i.d., 5μm)	Retention time	2.821 min
2012(1	Mobile phase	MeOH: ACN: KH ₂ PO ₄ at pH 4.0	\mathbb{R}^2	0.998
20[26]	Flow rate	0.4 mL/min	LOD	0,123 ng/mL
	Detection wavelength	220 nm	LOQ	0.374 ng/mL
	Run time	8 min	% RSD	< 2%
	System	HPLC-grade water system	Linearity range	1-100 ppm
2	Column	Unisphere Aqua C18 (4.6 x 150 mm, 3µ) column	R ²	0.9998
	Column temperature	40°C	LOD	1.173 ppm
21[27]	Mobile phase	Buffer(pH-6.50):Acetonitrile:Methanol -55:44:1	LOQ	3.555 ppm
	Flow rate	1.2 mL/min	% RSD	1.07 %
	Injection	20 μL		
	volume	·		
	Detection	210		
	wavelength	210 nm		
	System	Shimadzu prominence LC20AP (Shimadzu Corporation, Tokyo, Japan)		
22[28]	Column	InertSustainSwift C18 (G L Sciences, Eindhoven, Netherlands) 500 mm × 30 mm, 10 m particle size		
	Mobile phase	Water and acetonitrile		
	Flow rate	10 mL/min		
	Detection	210	1	
	wavelength	210 nm		
23[29]	System	RP-HPLC Shimadzu (Tokyo,Japan)	Retention Time	5 min
	Column	C18 segment (100 x 4.6 mm id, 5 µm molecule size)	R ²	0.998

IJCRT2308472 International Journal of Creative Research Thoughts (IJCRT) www.ijcrt.org e387

	Mobile phase	MeOH, ACN, 0.01mM KH ₂ PO ₄ (pH 3.5 0.5)adjusted with freshly prepared 10% orthophosphoric acid	LOD	0.013 μg/mL
	Flow rate	0.3-0.5 mL/min	LOQ	0.039 μg/mL
	Injection volume	20 μL		
	Detector	SPD-M20A prominence diode array detector		
	Detection wavelength	210 nm		
	System	Waters 2695 HPLC framework system	R ²	0.999
	Column	C18 section (150x4.6mm, 5 µm)	%RSD	0.649 %
	Column temperature	25°C	LOD	0.25 μg/mL
24[30]	Mobile phase	Phosphate cushion, Acetonitrile and Menthol in the proportion of 30:05:65)	LOQ	0.08 µg/mL
	Flow rate	1 mL/min		
	Detector	UV locator and Engage 2 programming Detector are used		
	Detection	249 nm		
	wavelength			
5	System	Technologies, USA)	Linearity range	20-320 μg/mL
	Column	Inertsil ODS-3,4.6x250 mm,5 μm porosity (GL Science, Japan)	Recovery range	40-120 μg/mL
25[21]	Column temperature	34 ⁰ C	%RSD	1.0-1.4%
23[31]	Mobile phase	MeOH ad TFA (0.1%v/v) in the proportion 52:48(v/v)	LOD	0.06 µg/mL
	Injection volume	20 μL	LOQ	0.20 μg/mL
	Detector	Diode array detector		
	Detection wavelength	321 nm		
	System	RP-HPLC System	Retention time	4.296 min
	Column	Kromsil C18 (4.6x250 mm, 5 mm) column	LOD	0.1 µg/mL
26[32]	Mobile phase	Phosphate buffer pH-5.8 and Acetonitrile in ratio of 80:20	LOQ	0.4 µg/mL
	Flow rate	1 mL/min	Percentage assay	99.2%
	Detector	Electronspray ionization- tandem mass spectrometry	Percentage recovery	100.1%

	Detection wavelength	215 nm		
	System	HPLC water 2469 system	Run time	10 min
	Column	C8 column 150x4.6 mm, 5 µm column	Retention time	6.5 min
	Column	25 ⁰ C		
	temperature	25 C		
27[33]	Mobile phase	Buffer pH-3.0 and (ACN:Methanol) in the ratio of (80:19:1)		
	Flow rate	1.2 mL/min		
	Injection volume	10 µL		
	Detector	UV/PDA detector		
	System	Waters Alliance 2690 or 2795 HPLC system (water, Milford, MA)		
	Column	YMC ODS-AR C18 column (3 μm, 4.6x150 mm)		
	Column	35 ° C		
	temperature	5 mM annuar inna an tair inna		
28[34]	Mobile phase	5 mM ammonium acetate containing 0.1% trifluoroacetic acid (pH-2.3)		
	Flow rate	1.0 mL/min		
	1 low fute			
	Injection volume	10 μL		
		Radioactivity detector with 250- µL		
	Detector	liquid cell (INUS B- RAM, Tampa,		
		FL)		
	LIDLC	Method: HPLC-MS/MS		<u> </u>
	MS/MS Plates	Oasis HLB 96-Well extraction plates using an automated system	LLOQ 2.0 ng/mL	
	Column	X Tera MS C18 5 μm column (150x2.1 mm; waters corp; Milford, MA, USA)	¥	
29[35]	Mobile phase	(A): 40% (10 mmol/L ammonium acetate (pH 8) methanol (95:5, v/v)		
	Flow rate	0.2 mL/min		
	Detector	API 3000 electrospray ionization mass spectrophotometer (Applied Biosystems, Foster city, CA, USA)		
30[36]	System	6460 triple quadrupole mass spectrometer.	Retention time	5.3 min
	Column	C18 column (250 × 4.6 mm, 5- Hypersil Gold)	\mathbb{R}^2	0.9999
	Mobile phase	acetonitrile and water $(\overline{40:60)}$	Linearity range	2-12 μg/mL
	Flow rate	1 mL/min	LOD	3.61 ug/mL
	Detection wavelength	220 nm	LOQ	10.96 ug/mL
	1	Method: UHPLC		
31[37]	System	Agilent 1290 series Ultra High- Performance Liquid Chromatography (US-CA)	Linearity range	20-100 μg/mL

	Column	Agilent Zorbax Eclipse Plus C18 (150×4.6mm, 5µm) column	R ²	0.999
	Column temperature	30 ⁰ C	LOD	2.20 μg/mL
	Mobile phase Acetonitrile and Potassium dihydrogen phosphate buffer (80:20, v/v)		LOQ	7.33 μg/mL
	Flow rate	0.6 mL/min	% RSD	< 2%
	Injection volume	5 μL		
	Detector	DAD detection		
	Detection wavelength	270 nm		
	0	Method: UHPLC-MS	•	
	System	Dionex Ultimate 3000RS device (Dionex, Sunnyvale, CA, USA)		
	~ 1	\sim Kinetex XB-C18 column (150 \times 2.1		
	Column	mm. 1.7 um)		
	Column	25 ⁰ C		
	temperature	25 0		
32[38]	temperature	A: 0.1% formin and in doionized		
	Mahila nhaqa	A. 0.176 formic acid in deformized		
	Mobile phase			
		B: 0.1% formic acid in acetonitrile		
	Flow rate	0.3 mL/min		
	Detection	100 450 nm		
	wavelength	190-450 mii		
		Method: UPLC		
	System	Highly sensitive UPLC System	Linearity range	2.5-15 μg/mL
		Acquity UPLC BEH C18 (2.1×50)	Theoretical	
	Column	mm 1.7 µm) column	plates	9417 Plates
	Colum		plates	
	tomporatura	25°C		
	temperature	0.05 M		
001001		0.05 M ammonium acetate buffer at		
33[39]	Mobile phase	pH 5.1 and methanol in the ratio of	Retention time	3.84 min
		45:65 (v/v)	-	
	Flow rate	0.3 mL/min	\mathbb{R}^2	0.9995
	Detector	PDA detector	LOD	0.03 µg/mL
	Detection			
	wavelength	215 nm	LOO	0.01 µg/mL
	U			10
		Method: UPLC-MS/MS		
		Tandem Triple quadrupole mass		
	System	spectrometer (AOUITV TOD)	Lincority	2.16 ng/mI
		spectrometer (AQUITITQD)	Lincarity	2-10 lig/lilL
	Calver	$C_{18}(50x^2 + xx^2 + 17xx^2) = 10xx^2$	LOD	1 I /
	Column	$C18 (50x2.1 \text{ mm}, 1.7 \mu\text{m}) \text{ column}$	LOD	1 mL/min
34[40]	Column temperature	15-20 ⁰ C	LLOQ	accuracy or 20 ng/mL
	Mobile phase	0.5% Acetic acid in Methanol and 0.02M aqueous Ammonium Acetate (10:90, v/v)	Accuracy	99.78 ±0.78

	Flow rate	0.5 mL/min		
	Injection	10 uI		
	volume	10 μL		
	Detector	Tandem mass detector		
	System	ACQUITY UPLC sysem from waters		T 1 5 0 0/
	5	corp.(Milford)	%RSD	Less than 5.0 %
		ACOUITY LIPLC BEH C8 column		
	Column	$(2.1 \times 50 \text{ mm}, 1.7 \text{ µm}, \text{waters})$	\mathbb{R}^2	0.998
	Column		LOD	0.015 / 1
	temperature	35°C	LOD	0.015 μg/mL
	Mobile phase	(A): water with 0.1% formic acid		
		(B): methanol containing 0.1% formic	LOQ	0.03 µg/mL
35[41]		acid		
	Flow rate	0.3 mL/min	Accuracy	14.1 μg/mL
	Injection	1 μL		
	volume			
		Xevo G2 O-TOR mass spectrometer		
	Detector	(waters) was equipped with an		
		electrospray ionization source (ESI)		
	Detection	220 nm		
	wavelength	220 IIII		
	Ì	HPG-3400 pump (Dionex UltiMate		
	System	3000 RSLC, Thermo Fisher Scientific,	Linearity range	0.025-1 μg/L
		Germany)		
		1.Acciaim RSLC C18 column (2.1 ×		
_		Scientific (Drejejch, Germany)		
	Column	2. ACOUITY UPLC BEH C18 1.7 µm.		
		VanGuard Pre-Column, Waters		
26[42]		(Ireland)		2
30[42]	Column	30°C	R ²	0.98
	temperature	500		0.90
		A H2O:MeOH (90:10) with 5 mM		
	Mobile phase	ammonium formate and 0.01 % formic		
		B MeOH with 5 mM ammonium		
		formate and 0.01 % formic acid		
	Injection			200/
	volume	5 µL	% RSD	< 20%
		Method: LC-MS		
	System	Model 515 pump system used	Linearity range	40-190 μg/mL
	Column	Purospher RP-18 end capped column	\mathbb{R}^2	0.9997
		$(125 \times 4.0 \text{ mm}, 5 \mu\text{m})$		
37[/3]	Mobile phase	2 mW ammonium acetate-acetonitrite (80.20 y/y)	LOD	2.99 μg/mL
37[43]	Flow rate	$(80.20, \sqrt{v})$	LOO	9.09.ug/mI
	Detector	model 2487 UV DAD detector	% RSD	0.26-0.55%
	Detection		/0100	0120 010070
	wavelength	210 nm		
38[44]	System	Shimadzu, Nexera-X2 (Shimadzu		
	System	Corporation, Japan)		
	Column	Waters X Bridge C-18, 250 mm × 4.6		
		mm, 5.0 m (Milford, MA, USA)		
		column		

		Column	0	
		temperature	45°C	
		1	Solvent A 0.05% of ammonia solution,	
		Mobile phase	pH - 9.2 with 5 M acetic acid (v/v) ,	
			Solvent-B: mixture of methanol-	
		Flow rate	$\frac{1}{0.6 \text{ mL/min}}$	
		Detection		
		wavelength	210 nm	
		Sustam	API3000 (Applied biosystems foster	
		System	city, calif) LOQ	2 ng/mL in 0.2 mL
		Column	V T MC C10.5	
			X Terra MS C18 5 μ m(150x2.1 mm)	
			corumi	
		Column	20.00	
30[/	151	temperature	30 °C	
57[7	[]		A(40%) : 10 mM Ammonium acetate-	
		Mobile phase	Methanol (95:5, v/v), pH8	
			B(00%): Acctomittle- Methanol (95:5 y/y)	
	/	Flow rate	0.2 mL/min	
		ĺ		
		Detector	Quantum Discovery (Thermo finnigan,	
		Detector	San Jose, calif) mass spectrometer	
		System	Micromass Quattro LC water	
		LC/MS/MS		1.
	4	Plates	96-well polypropylene plate	
		Column	Polaris 5- µm C18-A 50x2.0 mm	
		Column	column	
40[4	1 6]	temperature	45° C	
			(A): Methanol/10mM Ammonium	
		Mobile phase	acetate, pH 8.0 (5:95, v/v)	
			(B): acetonitrile/Methanol (10:90, v/v)	
		F1		
		Flow rate	0.2mL/min	
		volume	10 µL	
		System	LC-MS system	
			- -	
		Column	X Terra Ms C18 High- Performance	
			liquid chromatography (HPLC)	
41[47]			Column (150x2.1 mm, waters Corp; Milford MA_USA)	
			(A): 40% (10 m mol/L ammonium	
	1 7]	Mobile phase	acetate (pH-8): Methanol (95:5, v/v)	
			(B): 60% (Acetonitrile: Methanol	
			(10:90, v/v))	
		Flow rate	02 mL/min	
	Injection	10 µL		
		voluiile		

		API 4000 electrospray ionization mass						
	Detector	spectrometer (Applied Biosystems,						
		Foster city, CA, USA)						
Method: GC-MS								
	System	6890 N Agilent GC	Linearity range	3.5-300 ng/mL				
	Column	5973 N mass selective detector	LOD	1.5 ng/mL				
	Injection							
	volume	1 µL	LOQ	3.5 ng/mL				
42[48]								
	Detector	5% phenyl methylpolysiloxane						
		capillary column (30 m \times 0.25 mm i.d.						
		with 0.25 µm film thickness, Agilent						
		Technologies, USA)						
	Run time	6 min						

CONCLUSION:

In conclusion, this comprehensive review has delved into the multifaceted aspects of vildagliptin, encompassing its pharmacological, pharmaceutical, and analytical attributes. Through a meticulous analysis of existing literature, we have gained a profound understanding of vildagliptin's mechanisms of action, therapeutic indications, and clinical efficacy in managing type II diabetes mellitus. The review has also highlighted the critical role of formulation strategies in ensuring the drug's stability, bioavailability, and patient compliance. As we conclude this review, it is evident that vildagliptin holds a prominent place in the therapeutic landscape for type II diabetes management. Its unique mechanism of action and favourable safety profile make it an attractive option for clinicians and patients alike. Furthermore, advancements in pharmaceutical formulations and analytical techniques continue to enhance our understanding of vildagliptin's characteristics and optimize its administration. The analytical methods discussed herein underscore the significance of accurate and sensitive techniques in assessing vildagliptin's presence and concentration. These methodologies not only aid in quality control during pharmaceutical production but also contribute to bioequivalence studies and pharmacokinetic investigations, thereby promoting the drug's safe and effective use.

This comprehensive review serves as a valuable resource for researchers, clinicians, and pharmaceutical professionals, offering a consolidated understanding of vildagliptin's multifaceted nature and inspiring further scientific inquiry in the realm of diabetes therapeutics.

CONFLICT OF INTEREST:

All the authors have no conflict of interest.

References:

- 1. Croxtall JD, Keam SJ,2008: Vildagliptin: a review of its use in the management of type 2 diabetes mellitus. Drugs.;68(16):2387-409.
- Vaishnavi Aher, S. D. Mankar, G. S. Shinde,2021: Review on Analytical Methods for Estimation of Vildagliptin in Bulk and Pharmaceutical Dosage form, Research Journal of Science and Technology. 13(2):157-162.
- 3. Mathieu C, Degrande E, 2008: Vildagliptin: a new oral treatment for type 2 diabetes mellitus. Vasc Health Risk Manag;4(6):1349-60. doi: 10.2147/vhrm.s3005.
- 4. <u>https://www.ema.europa.eu/en/documents/product-information/galvus-epar-product-information_en.pdf</u>
- 5. https://www.caymanchem.com/msdss/14705m.pdf
- 6. Colleen D. Lauster; Teresa P. McKaveney; Sarah V. Muench,2007: Am J Health Syst Pharm;64(12):1265-1273.
- 7. Mayer T. Narkhede, Sachin S. Rane, Rajesh Y. Chaudhari and Vijay R. Patil, 2021: Development and Validation of UV Spectrophotometric Method for The Simultaneous Estimation of Vildagliptin and Metformin in Bulk Drugs and Pharmaceutical Dosage Form, World Journal of Pharmaceutical Research, Volume 10, Issue 13.

- 8. Prachi Joshi And Rajendra Kotadiya ,2023: Simultaneous Estimation of Remogliflozin Etabonate and Vildagliptin in A Tablet Formulation: UV Spectrophotometric And HPLC-PDA Method, Journal of Chemical Metrology.
- 9. Karajgi Santosh Raveendra, Shahid Momin Gaffar and Navanath V. Kalyane ,2016: First Derivative Spectrophotometric Simultaneous Determination of Vildagliptin and Metformin in Tablet Formulations, Pharmacophore, Vol. 7 (2), 109-117.
- Sujan Banik, Palash Karmakar and Md. Anowar Hossain Miah ,2015: Development and Validation of A UV-Spectrophotometric Method for Determination of Vildagliptin and Linagliptin in Bulk and Pharmaceutical Dosage Forms, Bangladesh Pharmaceutical Journal, 18(2):163-168.
- 11. Mahesh Attimarad, Katharigatta N. Venugopala, Bandar E. Al-Dhubiab, Rafea Elamin Elgack Elgorashe and Sheeba Shafi ,2021: Development of Ecofriendly Derivative Spectrophotometric Methods for The Simultaneous Quantitative Analysis of Remogliflozin and Vildagliptin from Formulation, Molecules, 26, 2160, 1-15.
- 12. Usharani Gundala , Chandra Shekar BhUVanagiri , Devanna Nayakanti ,2013: Simultaneous Estimation Of Vildagliptin And Metformin In Bulk And Pharmaceutical Formulations By UV Spectrophotometry , American Journal Of Pharmtech Research;3(1).
- Samer Housheh, Hanan Mohammad, Youssef Alahmad, 2013: Spectrophotometric Method For The Determination Of Vildagliptin In Bulk And Pharmaceutical Dosage Forms, International Journal Of Pharmaceutical Sciences Review And Research, 58(2), Pg.No.117-120.
- 14. Amanda Thomas Barden, Barbara Salamon,2012: Elfrides Eva Sherman Schapoval and Martin Steppe. Stability Indicating RP-LC Method for the Determination of Vidagliptin and Mass Spectrometry Detection for a Main Degradation Product. Journal of Chromatographic Science; 50:426-432.
- 15. Abdul SHAKOOR, Rashida BASHIR, Shaziz KHURSHID, Tabassam BASHIP, Saba IBRAHIM, Sajad HUSSAIN and Ahmad ADNAN, 2020: Liquid Chromatography Technique for Stimulaneous Estimation of Metformin and Vildagliptin. Application to Pharmacokinetic in Healthy Rabbits. Latin American Jounal of Pharmacy. 39 (3): 490-7.
- 16. Anna Gumieniczek, Anna Berecka ,2016: Analytical tools for determination of new oral antidiabetic drugs, glitazones, gliptins, gliflozins and glinides, in bulk materials, pharmaceuticals and biological samples, DE GRUYTER OPEN ,14: 215-242.
- 17. J. Panchal, B. Dhaduk And J. Dhalani ,2023: Stability Indicating Isocratic RP-HPLC And Second Derivative UV Spectroscopic Methods For Simultaneous Determination Of Remogliflozin Etabonate And Vildagliptin Hydrochloride, Rasayan J.Chem, Vol. 16, No. 2, 579-587.
- 18. Prachi Joshi And Rajendra Kotadiya ,2023: Simultaneous Estimation of Remogliflozin Etabonate and Vildagliptin in A Tablet Formulation: UV Spectrophotometric and HPLC-PDA Method, Journal of Chemical Metrology.
- 19. Pragati Ranjan Satpathy, V. Mohan Goud, Bhoga Bhagya, Jvc. Sharma & N.Shyamala ,2014: Development And Validation Of A Rp-HPLC Method For The Assay Of Vildagliptin, World Journal Of Pharmacy And Pharmaceutical Sciences, Vol 3, Issue 2.
- 20. A. M. Kashida , D. A. Ghorpadea , P. P. Toranmala , And S. C. Dhawaleb , 2015:Development And Validation Of Reversed Phase HPLC Method For The Determination Of Vildagliptin Using An Experimental Design , Journal Of Analytical Chemistry, Vol. 70, No. 4, Pp. 510–515.
- 21. Enas Al-Qudah, Sharif Arar, Kamal Sweidan, 2020: Forced Degradation Studies Of Vildagliptin Raw Material Alone And In The Presence Of Excipients Using HPLC-UV Analysis, J. Excipients And Food Chem. 11 (2).
- 22. Shrikrishna B. Baokar, Sugandha V. Mulgund , Nisharani S. Ranpise , 2013:Development And Validation Of Rp-HPLC Method For Simultaneous Estimation Of Vildagliptin And Metformin , Research Journal Of Pharmaceutical Dosage Forms And Technology , 5(2) , 95-98.
- 23. Razia Sultana, Sitesh C. Bachar and Fatema Rahman ,2013; Development and Validation of Stability Indicating Assay Method of Vildagliptin in Bulk and Tablet Dosage Form by Rp-HPLC, International Journal of Pharmacy & Life Sciences, 4(4): April.
- 24. Cristina Lopez, Raquel Diez, Jose M. Rodriguez, Matilde Sierra, Juan J. Garcia, Nelida Fernandez, M. Jose Diez and Ana M. Sahagun, 2013: Determination of Menbutone: Development and Validation

of a Sensitive HPLC Assay According to The European Medicines Agency Guideline, Seperations ,9,28.

- 25. Mahaveer Sharma And S. S. Agrawal ,2021: Bioequivalence Study of Different Brands of Vildagliptin in Healthy Human Subjects, Future Journal of Pharmaceutical Sciences ,7:155.
- 26. Krishnan Balamurugan, Kirtimaya Mishra ,2020: Optimization and Validation of The Simultaneous Determination of Vildagliptin and Metformin, In Bulk and Formulation by A Reverse Phase HPLC Method Using D-Optimal Experimental Design, Journal of Global Pharma Technology, Vol.12, Issue 08, 01-12.
- 27. J. Panchal, B. Dhaduk And J. Dhalani ,2023: Stability Indicating Isocratic Rp-HPLC And Second Derivative UV Spectroscopic Methods For Simultaneous Determination Of Remogliflozin Etabonate And Vildagliptin Hydrochloride, Rasayan J.Chem , Vol. 16, No. 2, 579-587.
- 28. Neeraj Kumar, Subba Rao Devineni, Gurmeet Singhb, A. Kadirappa, Shailendra Kumar Dubeya, Pramod Kumar ,2016: Identification, Isolation and Characterization of Potential Process-Related Impurity and Its Degradation Product in Vildagliptin, Journal of Pharmaceutical and Biomedical Analysis, 119, 114-121.
- 29. Balamurugan Krishnan, Kirtimaya Mishra, 2010: Quality by Design Based Development and Validation of Rp-HPLC Method For Simultaneous Estimation of Sitagliptin and Metformin in Bulk and Pharmacuetical Dosage Forms, Int.J. Pharm.Investigation;10(4):512-518.
- 30. S.K.Godasu, Pothula.Raju, Gopinadh.Vuyyala, Varun.Dasari, Perli.Krantikumar Sri Indu Institute Of Pharmacy, Sheriguda (V), Ibrahimpatnam (M), Hyderabad, 2023: Rp-HPLC Method For Stimulaneous Estimation Of Anti Diabetic Drugs In Api Dosage Form. Journal Of Engineering Sciences Vol 14 Issue 03, 96-107.
- Gedeon Richter Romania 540306, Tirgu Mures, Romania.2020: Reversed Phase HPLC For Strontium Ranelate:Method Development And Validation Applying Experimental Design. Acta Pharm.68 171-183.
- 32. Anil Kumar Tallam , Alapati Sahithi , Mohana Vamsi Nuli. International Joural Of Health Care And Biological Sciences. Int Jou Hea Bio Sci, 4(1), 2023, 18-24.
- 33. Ahire Sujeetkumar, Ahire Sandhya, Dr. Patil P.R., Jain Akshata, Chavhan Archana, 2023: Development Of Rp-HPLC Method For Vidagliptin In Pharmacuetical Dosage Form. International Jounal Of Pharmacuetical Sciences Review And Research, January-February; Article N.06, Page;36-40.
- 34. Handan He, Phi Tirn, Hequm Yin, Harold Smith, Dennis Flood, Roger Kramp, Ron Filipeck, Volker Fischer And Dan Howard, 2023: Disposition Of Vildagliptin, A Novel Dipeptidyl, Peptidase 4 Inhibitor, In Rats And Dogs. Dmd Fast Forward. Published On December 15.
- 35. Yan-Ling He, Ron Sabo, Franck Picard, Yibin Wang, Jerry Herron, Monica Ligueros-Saylan and William P, 2019: Dole. Study Of the Pharmacokinetic Interaction of Vidagliptin and Metformin in Patients with Type 2 Diabetes. Current Medical Research and Opinions. Vol.25, No. 5,1265-1272.
- 36. Chaitali Dhale1 and Janhavi R Rao ,2019: Stability Indicating HPLC MS Method for Determination of Degradation Products in Vildagliptin, Journal of Analytical & Bioanalytical Techniques ,10:2.
- 37. Viralkumar Patel, Chintan Pandya, Aditee Pandya, Dharmesh Patel, Zalak Patel ,2021: Novel UHPLC DAD Method for Simultaneous determination of Vildagliptin and Metformin in Bulk and its Tablet formulation, Research J. Pharm. and Tech. 14(8).
- 38. Anna Gumieniczek, Anna Berecka-Rycerz , Emilia Fornal , Barbara Zy zy nska-Granica and Sebastian Granica , 2021:Comprehensive Insight into Chemical Stability of Important Antidiabetic Drug Vildagliptin Using Chromatography (LC-UV and UHPLC-DAD-MS) and Spectroscopy (Mid-IR and NIR with PCA), Molecules , 26, 5632, 1-28.
- 39. Syed Mastan Ali, Ponnuri Bharath, Syed Khasim Sharif, D. Ramachandran, 2021: Simple and Fast Stability Indicating UPLC Method for the Simultaneous Quantification of Vildagliptin and Remogliflozin Etabonate in Bulk Drug and Formulations, Current Trends in Biotechnology and Pharmacy Vol. 15 (4) 401 407.
- 40. Ramzia I. EIBagary , Hassan M.E. Azzazy, Ehab F. ElKady and Faten Farouk. 2013: Simultaneous determination of metformin, vildagliptin, anad 3 amino-1-adamantanol in human plasma: Application to pharmacokinetic studies. Journal of liquid Chromatography and Related Technologies, At:11:21.

- 41. Camila Ferrazza Alves Giordani, Sarah Campanharo , Nathalie Riberio Wingert , Livia Maronesi Bueno , Joanna Wttckind Manoel , Cassia Virginia Garcia , Nadia Maria Volpato , Gabrielle Dineck lop , Paoka de Azevedo Mello , Erico Marlon de Moraes Flores , Elfrides Eva Scherman Schapoval and Martin Steppe,2020: UPLC-ESI/Q-TOF MS/MS Method for Determination of Vildagliptin and its Organic Impurities. Journal of Chromatographic Science,Vol.58,No.8, 718-725.
- 42. Pablo Gago-Ferrero , Anna A. Bletsou , Dimitrios E. Damalasa , Reza Aalizadeha , Nikiforos A. Alygizakisa , Heinz P. Singerc , Juliane Hollenderc, , Nikolaos S. Thomaidis ,2020: Wide-scope target screening of > 2000 emerging contaminants in wastewater samples with UPLC-Q-ToF-HRMS/MS and smart evaluation of its performance through the validation of 195 selected representative analytes , Journal of Hazardous Materials , 387, 121712.
- 43. Anna Gumieniczek , Anna Berecka-Rycerz , Emilia Fornal , Barbara Zy zy nska-Granica and Sebastian Granica ,2021: Comprehensive Insight into Chemical Stability of Important Antidiabetic Drug Vildagliptin Using Chromatography (LC-UV and UHPLC-DAD-MS) and Spectroscopy (Mid-IR and NIR with PCA), Molecules , 26 , 5632 , 1-28.
- 44. Neeraj Kumar, Subba Rao Devineni, Gurmeet Singhb, A. Kadirappa, Shailendra Kumar Dubeya, Pramod Kumar, 2016:Identification, isolation and characterization of potential process-related impurity and its degradation product in vildagliptin, Journal of Pharmaceutical and Biomedical Analysis, 119, 114-121.
- 45. Y.-L. He. R. Sabo, J. Campestrinil.Y.Wang. M. Ligueros-Saylan. K. C. Lasseter.S.C. Dilzer. D. Howard. W.p. Dole.2017: The Infulence of hepatic impairement on the pharmacokinetics of the dipeptidyl peptidase IV (DPP-4) inhibitor vildagliptin. Eur J Clin Pharmacol 63(7):677-686.
- 46. Handan He, Phi Tran, Heqn Yin, Harold Smith, Yannok Batard, Lai Wang, Heidi Einolf, Helen Gu, James B.Mangold, Voler Fisher, and Dan Howard, 2009: Absorption, Metabolism, and Excretion of [14 C] Vidagliptin, a Novel Dipeptidyl Peptidase 4 Inhibitor, in Humans. Drug Metabolism And Disposition, DMD 37:536-544.
- 47. Sachiko Mita, Shripad D. Chitnis, Kenneth Kulmatycki, Atish Salunke, Yan-Ling He, Wei Zhou, and Hikoe Suzuki, 2016: Bioequivalence and food effect assessment for vidagliptin/metfromin fixed-dose combinations of vildagliptin and metformin in Japnese healthy subjects. International Journal of Clincal Pharmacology and Therapeutics, Vol. 54- No. 4/2016.
- 48. Ebru Ucakturk ,2015: Development of Sensitive and Specific Analysis of Vildagliptin in Pharmaceutical Formulation by Gas Chromatography-Mass Spectrometry , Journal of Analytical Methods in Chemistry Volume 2015, 7 pages.