

Tamil Nadu is one of the 28 States of India. Its capital is Chennai (formerly known as Madras) the largest city. Tamil Nadu lies in the southern most part of the Indian Peninsula and is bordered by the union territory of Puducherry and the states of Kerala, Karnataka and Andhra Pradesh. Coimbatore is the city in Tamil Nadu, South India. The city is located on the banks of the Noyyal River surrounded by the Western Ghats and is administered by the Coimbatore Municipal. Nirmala college academic campus is located in the southern parts of the Western Ghats. The total area of the college campus is 20 acres. The temperature during both summer and winter varies between 28° c to 34° c. Soil in this area is red loamy soil which is more fertile than sandy soil. Its porosity allows high moisture retention and air circulation.

I. Collection of selected tree sample

For the present study *Mimusops elengi*, was selected in the Nirmala college campus to find out the Morphology and propagation of the selected tree, Physico-chemical parameters of the tree canopy soil, mineral profile of the litter formed by the selected tree canopy, microbial flora of the selected tree canopy soils were analyzed. The data were then processed and represented both in Tables and charts.

Taxonomic Position

Division : Phanerogams
 Class : Dicotyledons
 Sub class : Gamopetalae
 Series : Heteromerae
 Order : Ebinales
 Family : Sapotaceae
 Genus : *Mimusops*
 Species : ***M. elengi*, Linn.;**



Mimusops elengi, Linn.; is a native to India. It is commonly known as Spanish cherry and bullet wood. It is a large medium-sized evergreen tree found in tropical forests in South Asia, northern Australia. This tree is very famous for its shade, fragrant flowers and above all its elegant looks.

This tree is chiefly cultivated for its ornamental appearance and its fragrant flowers. The tree produces small fragrant flowers in abundance during the hot season. Traditionally all different part of this plant, namely leaf, root, fruit, seed, bark and flower are used to cure various kinds of disorders. . The wood is a luxurious wood that is extremely hard, strong and tough, and rich deep red in colour. The bark, flowers, fruits, and seeds of *Bakula* are used in Ayurvedic medicine in which it is purported to be astringent, cooling, anthelmintic, tonic, and febrifuge. It is mainly used for dental ailments such as bleeding gums, dental caries, and loose teeth. It is known for its strengthening property for teeth. It is well documented for several medicinal properties like diuretic effects, gastro protective, antibacterial, antifungal, antihyperglycemic etc.

II. A. Morphological characteristics of the selected tree and propagation

Morphological characters of the selected tree species were recorded. The selected trees total height and width. Leaf, leaflet, flower, fruits - size and colours were measured.

B. Biodiversity of the selected tree

Biodiversity of species such as Ants, Crow, Sparrow, Pigeon, Dragon fly, Mynah, Butterflies, Lac insect, Lizards, Calottes, Chameleon, Spider, Worms, Honey comb, Honey bee, Wasp, Parrots, Grasshopper, Sparrow were observed and recorded during the study period.

C. Average annual litter of dried leaves and logs of the selected tree canopy

The litter of dried leaves and logs of the selected tree canopy were collected throughout the year and the average annual fallings were calculated.

III. Microbial analysis

Collection of the selected tree canopy soil sample

The tree canopy soil samples were collected during the year, 2014-2015. Soil with litter formation and ground vegetation from the selected tree canopy of *Albizia lebeck*, (L.) Benth.; were collected

separately in sterile bags, air dried and sieved for further analysis. Barren land soil, taken from the same campus was kept as control. Soil was taken from the depth of (0-15 cm depth). Soil samples were packed in sterile bags and used for further analysis.

Isolation and culture of microorganisms

Preparation of nutrient medium: Potato-Dextrose Agar (PDA)

120 gms of freshly peeled potato is taken in to a flask and 150 ml of water is added to it. It is boiled for 10 minutes. Then the potato extract is taken and its volume is made up to 150 ml by adding distilled water. To this extract, 7.5 gms of Dextrose is added and thoroughly mixed. Then the solutions were poured in a 500 ml flask and stirred thoroughly. This content is heated in a water bath to dissolve the agar. This medium is dispensed in culture petridishes and kept in laminar air flow for solidification.

Serial dilution method

For the enumeration of microbial population a set of ten selected soil samples (0-15 cm depth) were collected. Soil microbial communities have relied on culturing techniques using PDA (Potato Dextrose Agar) medium. Serially diluted samples were inoculated on petridishes containing PDA medium and incubated in the laboratory for 5 days at 30°C (Kanika Sharma, 2007). The bacterial and fungal colonies were counted using colony counter for three days and the culture was kept in the refrigerator at 4°C. 1 gm of 1% Crystal violet is dissolved in 10 ml of 95% ethyl alcohol and final volume is made up to 100 ml with distilled water. Bacterial colony appears blue and for identification.

Identification of Bacteria (Direct microscopic examination)

An average volume of bacterial cell is 1 cubic micron. They are smallest forms among bacteria. After division the cells may either separate from each other or may remain joined together to form groups of two cells in *Diplococcus*, a tetrad of four cells in *Micrococcus tetragenus* and a chain of cells in *Streptococcus* (Bergey, 1957).

Identification of Fungus

The smear was simple stained to study the morphology of the cells. Basic stain for simple staining Safranin is used for identifying microbes and the data's were recorded. For each experiment replicas were repeated (Mani *et al.*, 2004).

IV. Physicochemical parameters

Physicochemical parameters of the select tree canopy, litter and barren soils were analyzed.

1. pH of the soil

Part of the moist soil samples were air dried and sieved to obtain fine soil samples (2 mm). pH = Hydrogen-ion-concentration, The H^+ concentration i.e., $pH = \log (1/H^+)$

The pH of the medium, if found to be acidic, is brought to the required pH by adding 0.1 (N) NaOH drop wise and testing with pH paper after thoroughly mixing with a glass rod. Conversely, 0.1 (N) HCl is used to get an acidic pH of the medium.

2. Moisture content of the soil

Moisture content is the ratio of the mass of water in the sample to the mass of solids in the sample. Moisture content of the selected tree canopy litter samples were calculated and expressed in percentage (Conventional oven method ASTM, 2001).

3. Water holding capacity and temperature of the soil

Water holding capacity and temperature of the soil were analyzed as per the standard method.

4. Mineral profile of the selected tree canopy soil samples

Mineral like Potassium, Phosphorus, Calcium, Magnesium, Iron and Sodium were analyzed in the standard laboratory by employing Atomic Absorption Spectrophotometer by following the method of Issac and Johnson (1975) and the results were recorded.

Estimation of calcium and magnesium (Jackson, 1967)

5ml of triple acid digested extract was taken in a China dish. To this 10 ml of 10% NaOH and 0.1g of Murexide indicator powder (40 g of potassium sulphate or potassium chloride was ground with 10 g

ammonium purpurate) were added and titrated against 0.02 N versenate (19 g of EDTA was dissolved in 5liters of distilled water) and standardized against 0.2 N Na₂ CO₃ solution and adjusted until the colour changes from red to violet.

Calcium and Magnesium

5ml of triple acid digested extract was taken in a China dish, to this 10 ml of ammonium chloride - ammonium hydroxide buffer pH 10 and few drops of Eriochrome Black T indicator (0.1 g of Eriochrome Black T was dissolved in 25ml of methanol containing 1g of hydroxylamine hydrochloride) were added and titrated against 0.02N versenate solution until the colour changes from red to blue.

Calculation

Percentage of calcium = $\text{Titre value of calcium} \times 100 / 5 \times 100 / 0.5 \times 0.0004$

Percentage of magnesium = $\text{Titre value of calcium + magnesium} - \text{titre value of calcium} \times 0.96$

Calcium and magnesium contents were expressed as mg/100 g of sample

Estimation of Sodium and Potassium

Sodium and potassium were estimated by using Flame Photometer, Model-EFL. The sodium and potassium contents were calculated by referring to the calibration curves of sodium and potassium, respectively, and expressed as mg/100 g on dry weight basis.

Phosphorus estimation (Dickman and Bray, 1940)

One ml of triple acid digested extract was pipetted into 100 ml volumetric flasks. To this 50 ml glass distilled water was added, followed by 5 ml of ammonium molybdate sulphuric acid reagent (Solution A: 25 mg of ammonium molybdate was dissolved in 100 ml of distilled water. Solution B: 280 ml of conc. H₂ SO₄ was diluted to 800 ml). Solution A was added slowly with constant stirring to solution B and the volume was made up to 100 ml with glass distilled water). Blue colour was developed by adding six drops of 2.5% stannous chloride solution. The total volume was made up to 100 ml. The intensity of the blue colour was measured at 650 nm in a spectrophotometer. The phosphorus content present in the sample was calculated by referring to a standard curve of phosphorus and expressed as mg/100 g on dry weight basis.

Estimation of iron by atomic absorption spectrophotometer (Issac and Johnson, 1975)

Estimation

By feeding the sample to an Atomic Absorption Spectrophotometer the iron content was estimated at 246.8 nm wavelength and the readings were expressed in mg/100g of sample on dry weight basis.

V. Analysis of the selected tree canopy litter formed by the selected samples

Collection of tree canopy litter samples

From a composite of litter fall, the fallen fresh/dried leaves, wood logs, flowers, fruits and seeds were collected under the canopy of the ten trees separately and shade dried, packed in sterile bags then powdered and lumped in a composite of sample for chemical analysis. The maximum litter fall of various seasons during the year 2014 (January-March, April-June, July-September, October-December) were analyzed.

1. pH and moisture content

pH and moisture content of the litter were analyzed as per the standard methods.

2. Mineral analysis of the selected tree canopy litter samples

Mineral analysis of Potassium, Phosphorus, Calcium, Magnesium, Iron and Sodium minerals were analyzed in the recognized laboratory by employing Atomic Absorption Spectrophotometer. Mineral profiles of the litter formed by the selected tree canopy, the fallen fresh/dried leaves, wood logs, flowers, fruits and seeds were powdered and kept in airtight container then the mineral profiles were analyzed and the mineral profile of the selected tree canopy soil and litter samples were experimented and recorded by following standard methods of (Association of Official Agricultural Chemists) AOAC, (1990).

RESULTS AND DISCUSSION

Comparative morphology of the selected trees, leaves, inflorescence, flower, fruit, pod (dehiscent/indehiscent) and its propagation, Micro and Macrobial biodiversity were observed and represented in the following Tables.

Sample	Tree	Height in (m)	Breadth in (m)	Leaf		Inflorescence	Flower colour	Fruit		Seed shape and colour	Propagation	Biodiversity
				Type	Shape							
<i>Mimusops elengi</i>	Medium sized evergreen	06.01	01.08	Simple	Oblong elliptic Glossy, oval	Solitary or fascicled	Cream, hairy scented	Berry, yellow, ovoid	Creamy yellow flesh	Cream, hairy scented	Cream, hairy scented	Berry, yellow, ov

Table - 1 Comparative morphological characters, Propagation and the biodiversity of the selected tree

Table - 2 Morphology of the Leaf/ Leaflet length


Sample	Simple/ compound	Leaf length in (cm)	Leaflet length in (cm)	Leaf/ Leaflet of the selected trees
<i>Mimusops elengi</i>	Simple, Oblong elliptic,	07.07	00.00	

Table - 3 Morphology of the inflorescence and flower of the selected trees


Sample	Inflorescence	Flower		Inflorescence and flower of the selected trees
		Colour	Length in (cm)	
<i>Mimusops elengi</i>	Solitary or fascicled	Cream, hairy scented	01.08	

Table - 4 Morphology of the fruits


Sample	Fruit				Fruit of the selected trees
	Type	Colour	Shape	Length in (cm)	
<i>Mimusops elengi</i>	Creamy yellow flesh, Berry	Orange	Berry, Creamy yellow flesh	2.5	

Table - 5 Dehiscent and indehiscent seeds of the selected trees

Sample	Pod
	Dehiscent / Indehiscent
<i>Mimusops elengi</i>	Dehiscent

Table - 6 Biodiversity of the selected trees

Sample	Biodiversity of the selected trees
<i>Mimusops elengi</i>	Ants, Butterflies, Squirrels, Mynah, Wasp.

Table - 7 Average annual litter of dried leaves and logs of the selected tree canopy

Sample	January-March (gm)	April - June (gm)	July-September (gm)	October-December (gm)	Average annual litter of the selected tree canopy in (%)
<i>Mimusops elengi</i>	435.00	751.40	362.40	153.40	4.25

Table - 8 Enumeration of the Bacterial colony of the selected tree canopy soil

Sample	Number of Bacterial Colony								
	Day 1			Day 2			Day 3		
	10 ⁻³	10 ⁻⁶	10 ⁻⁹	10 ⁻³	10 ⁻⁶	10 ⁻⁹	10 ⁻³	10 ⁻⁶	10 ⁻⁹
Control	3	3	2	5	4	3	5	7	6
<i>Mimusops elengi</i>	5	6	8	5	6	8	6	7	8

Table - 9 Bacteria present in the selected tree canopy soil

Sample	Bacteria		
	10 ⁻³	10 ⁻⁶	10 ⁻⁹
Control	<i>Streptococcus sps</i>	<i>Staphylococcus sps</i>	<i>Streptococcus sps</i>
<i>Mimusops elengi</i>	<i>Pseudomonas sps</i>	<i>Diplococcus sps</i>	<i>Corynebacterium sps</i>

Table - 10 Enumeration of Fungal colony of the selected tree canopy soil

Sample	Number of Fungal Colony								
	Day 1			Day 2			Day 3		
	10 ⁻³	10 ⁻⁶	10 ⁻⁹	10 ⁻³	10 ⁻⁶	10 ⁻⁹	10 ⁻³	10 ⁻⁶	10 ⁻⁹
Control	-	-	-	3	3	2	3	3	2
<i>Mimusops elengi</i>	-	-	-	0	1	1	1	1	1

Table - 11 Fungus present in the selected tree canopy soil

Sample	Fungi		
	10 ⁻³	10 ⁻⁶	10 ⁻⁹
Control	<i>Aspergillus niger</i>	<i>Aspergillus glaucus</i>	<i>Aspergillus niger</i>
<i>Mimusops elengi</i>	<i>Aspergillus niger</i>	<i>Rhizopus</i>	<i>Aspergillus niger</i>

Distribution of Microbes present in the selected individual tree canopy soil (Plate-4)

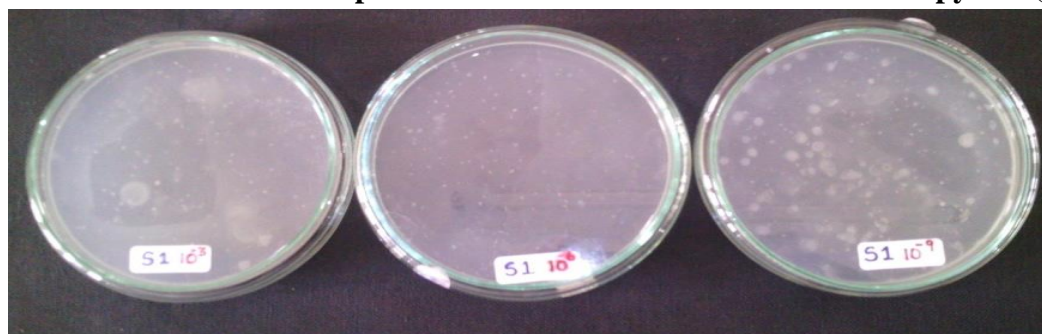


Table - 12 Moisture content and pH of the selected tree canopy soil

Sample	Fresh weight (gm)	Dry weight (gm)	Moisture content (%)	pH
Control	20	18.86	5.7	5.7
<i>Mimusops elengi</i>	20	17.57	12.2	6.9

Table - 13 Mineral profile of the selected tree canopy soil

Sample	Potassium (%)	Phosphorus (%)	Calcium (%)	Magnesium (%)	Iron (%)	Sodium (%)
Control	0.39	0.10	0.31	0.081	0.048	0.18
<i>Mimusops elengi</i>	0.19	0.06	0.48	0.12	0.076	0.30

Table - 14 Moisture content and pH of the selected tree canopy litter

Sample	Fresh weight (gm)	Dry weight (gm)	Moisture content (%)	pH
<i>Mimusops elengi</i>	362.04	153.04	57.72	6.3

Table - 15 Mineral profiles of the selected tree canopy litter

Sample	Potassium (%)	Phosphorus (%)	Calcium (%)	Magnesium (%)	Iron (%)	Sodium (%)
<i>Mimusops elengi</i>	838	216	411	230	18	22

CONCLUSION

India is urbanizing at a very fast pace. The enhancement of urban green spaces or urban green forests is one of the ways, which has the potential to mitigate the adverse effects of urbanization economic or environmental costs. Urban forestry is the art, science and technology of managing trees and forest resources in and around urban community ecosystems for physiological, sociological ecological, educational environmental and aesthetic benefits for society. In urban environment human alter soil forming factors by impacts associated with urban infrastructure for instance, building specifications often results in the scraping, compacting and covering of urban soil which can impact soil organic matter, texture, structure, infiltrations, aeration, root penetration and biological activity. The research on urban greening is very meagre particularly in India. Hence, the study on selected individual tree canopy of the soil and litter in urban greening to enrich the urban soil and to promote plant growth to the urban environment.

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