BIOLOGICAL INVESTIGATION OF SUBSTITUTED ACETOPHENONE DERIVED SCHIFF BASE NICKEL (II) COMPLEX

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Abstract: A tridentate ligand,2-(((2-aminoethyl)imino)methyl)phenolobtained by the condensation of 2-hydroxyacetophenone and ethylenediamine andits Nickel complexhas been synthesized and characterized by spectroscopic studies viz., FT-IR, 1H NMR and UV–vis. The synthesized ligand and its complex has beentested against *Escherchia coli*, *Proteus mirabilis*, *Klebsiella pneumonia*, *Salmonella typhi*, *Salmonella paratyphi 'A'*, *Pseudomonas aeroginosa* and *Staphylococcus aereus* and the results suggested that Ni (II) complex has significant antimicrobial activity. A comparative study of minimum inhibitory concentration (MIC) values of Schiff base ligand and its Ni (II) complex.

Keywords: Antibacterial, 2-hydroxyacetophenone, Haemolytic assay, Escherchia coli, Proteus mirabilisand Staphyloccoccusaueus

I. INRODUCTION

Schiff bases have been studied as a class of ligands [1] and are known to coordinate with metal ions through the azomethine nitrogen atom. The metal based drugs, where metal plays an important role, occupy a prominent place in pharmaceutical chemistry; this has been highlighted in many reviews [2]. Other useful applicationsare sensor [3], electrodes [4], conducting polymer [5], energystorage [6], enzymatic application [7], solar cell [8], antiviral[9], antifungal [10], anti-inflammatory [11], antitumor [12], antiparasitic[13], antibacterial [14], anti-HIV [15], anticancer [16], etc. Nickel is usually dipositive in its compounds, but itcan also exist in the oxidation states 0, 1+, 3+, and 4+. Inaddition to the simple nickel compounds or salts, nickelforms a variety of coordination compounds. Currently,the bioinorganic chemistry of nickel is a topic ofincreasing interest because the study of the interactions nickel (II) with Schiff bases offers an opportunity to understand various properties of nickel (II) complex of nickel (II) ion with these Schiff base have been prepared and characterized. The prepared Schiff base and nickel (II) complex has been tested for possiblebiological (antibacterial and haemolytic) activities.

Metal complexes of Schiff base ligands have recently been used as precursors in the preparation of nanostructures of the respective metal oxides. They are important compound due to their wide range of biological activities and industrial application. The objectives of the present investigations (i) To synthesis of Schiff base ligand and its Ni (II) complex. (ii) To characterization by FTIR, ¹H NMR and UV-Visible spectroscopy. (iii) To study the antibacterial activity. (iv) Tostudy haemolytic assay.

II. EXPERIMENTAL PROCEDURE

2.1. Materials

All the chemicals used were chemically pure and AR grade. Solvents were purified and dried according to standard procedures. 2-hydroxy acetophenone and nickelchloridehexahydrate were obtained from Merck specialties Ltd., Mumbai, India

2.2. Synthesis of Schiff base ligand

The tritendateSchiff base was prepared by condensation of ethylenediamine with 2-hydroxyacetophenone (Scheme 1). Equimol (0.5 mmol) quantity of 2-hydroxyacetophenone and ethylenediamine was dissolved in 20 mL of ethanol and the solution was refluxed for 4 h under constant stirring. This condensation reaction was carried out by using acid catalyst (few drops of glacial acetic acid). The formed water was removed from the reaction mixture using sodium sulfate (dehydrating agent). After completion of the reaction, the mixture was reduced to half of its original volume using a water bath and kept aside at room temperature. Yellow crystals of ligand were obtained from slow evaporation (Yield: 83%). The Schiff base is characterized by FTIR, UV- Vis and Proton NMR spectroscopy.

2.3. Synthesis of Schiff base Nickel (II) complex

Nickel(II) chloride (0.1 mmol) and the potential tridentate Schiff base ligand (0.2 mmol) were dissolved in acetone (20 ml) and the mixture was heated to reflux for 5 h and the reaction was monitored by TLC. After partial evaporation of the solvent,

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solid metal(II) Schiff base complex (Scheme 2) were separated and dried in vacuum (Yield: 86%) and character by FTIR, UV-Vis and Proton NMR spectroscopy.

2.4. Antibacterial Assay (MIC)

Seven bacterial isolates; Escherchia coli, Proteus mirabilis, Staphylococcus aereus, Klebsiella pneumonia, Salmonella typhi, Salmonella paratyphi 'A' and Pseudomonas aeroginosaprocured fromMicrobiology Laboratory, KAPV Medical College, Tiruchrappalli, Tamilnadu were sub cultured periodically and had been stored in glycerol semi solid media. These pure isolates were inoculated in 1% Peptone water and incubated at 37°C for overnight.

The prepared acetophenone derivatives 2-(((2-aminoethyl)imino)methyl)phenol Bis(2-(1-((2and aminoethyl)imino)ethyl)phenoxy)zinc(II)were tested for Minimum Inhibitory Concentration for seven different bacterial isolates adopting the method of [17]. All the wells designed for the protocol were filled with100µl of 1% peptone water and hundred micro liter of the product (mg/ml) was pipetted into each well of the first column of the plates designed for bacterial strains. Then a serial dilution was made to ensure that the first well of each organism loaded had 50µl of the product and the succeeding wells had half of volume of protein solution from the previous well in serially descending concentrations. Thereafter, 10µl of the bacterial suspension following the same volume of resazurin indicator were added and incubated for 24 hours. The MIC values were determined by visual observation of colour change. The colour change from purple to pink or colourless was recorded as the presence of bacterial growth whereas the retaining of original colour of the dye, purple means inhibition of growth. The experiments were done in triplicate and the data were presented in mean values.

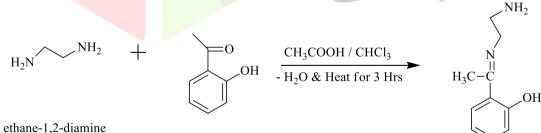
2.5. Haemolytic Assay

The MHC (Minimum Haemolytic Concentration) of the products was carried out in micro well plates according to [18] with little modification. The human blood sample was gifted by a blood bank located at Tiruchirappalli, Tamilnadu, India. The red blood cells were sedimented and repeatedly washed with 0.9% NaCl. The cells were made into 4% suspension with normal saline. All the test wells were filled with 100 µl of normal saline. The first wells of the marked column for the product were filled with 100µl of the respective sample solutions and serially diluted to ensure that each first well had 50µl and serially descending concentration of the product. Then 100µl of 4% hRBCs was added in the wells of the respective rows. The 4% hRBCs alone and 4% hRBCs in 10% SDS were used as 0% (Negative Control) and 100% (Positive Control) haemolytic controls respectively. After 2 hours of incubation the button formation and absence of colour change was recorded as absence of haemolysis and the vice versa was recorded as presence of haemolysis. The MHC of a test sample is defined as the lowest concentration of protein at which 100% haemolysis occurs. The experiments were done in triplicate and the data were presented in mean values.

III. RESULTS AND DISCUSSION

3.1 Reaction Synthesis of Ligand and Complex

Schiff base ligand prepare by the condensation of derivative of acetophenone and ethylenediamine with molar equation, by Scheme 1

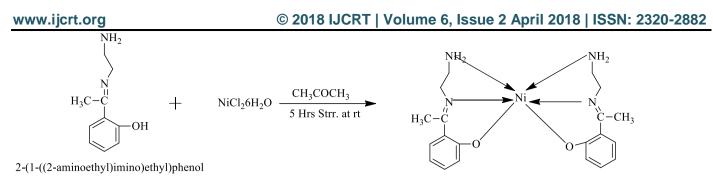


1-(2-hydroxyphenyl)ethanone

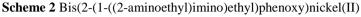
2-(1-((2-aminoethyl)imino)ethyl)phenol

Scheme 1 Preparation of 2-(1-((2-aminoethyl)imino)ethyl)phenol

The Nickel (II) complex prepare by 2:1 ratio of Schiff base ligand and Zn(II) metal salt by Scheme 2.



bis(2-(1-((2aminoethyl)imino)ethyl)phenoxy)nickel(II)



3.2 UV-Visible spectra

The UV-visible spectra of Preparation of 2-(1-((2-aminoethyl)imino)ethyl)phenoland Bis(2-(1-((2-aminoethyl)imino)ethyl)phenoxy)nickel(II)were recorded in aqueous solution (10^{-3} M) in the range of 200-1100 nm at room temperature. The electronic spectrum of the ligand shows a broad band at 378.90 nm and 98.9759 due to Π - Π * transition of the azomethine (-N=C<) chromospheres, the intense absorption band at high energy, 0.1925 (AU) in the region 208.20 nm presumably for the Π - Π * transition of the benzylidene ring of ligand shows in **Fig. 1**, **2**. The electronic spectrum of the Ni(II) complex shows a broad band at 360.00 nm and 95.635 due to d-d transitions of Ni (II) complex, the intense absorption band at high energy, 2.6704 (AU) in the region 207.00 nm shows in **Fig. 3**, **4**.

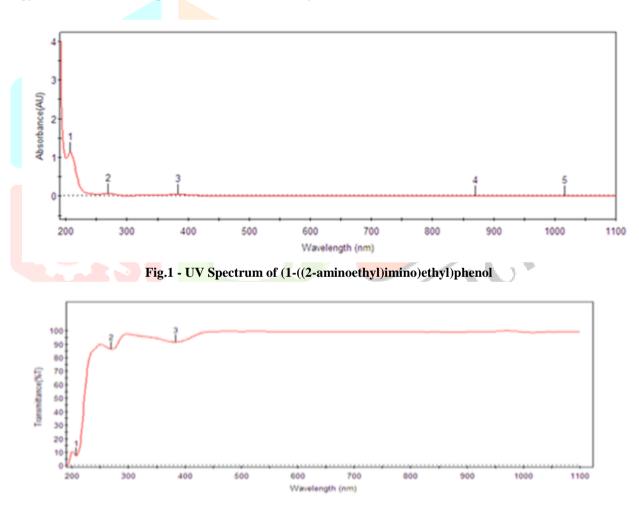


Fig.2 - Visible Spectrum of (1-((2-aminoethyl)imino)ethyl)phenol

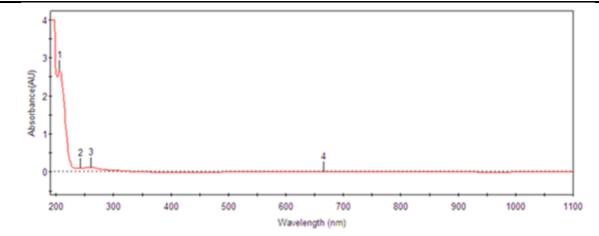


Fig3 - UV Spectrum of Bis(2-(1-((2-aminoethyl)imino)ethyl)phenoxy)nickel(II)

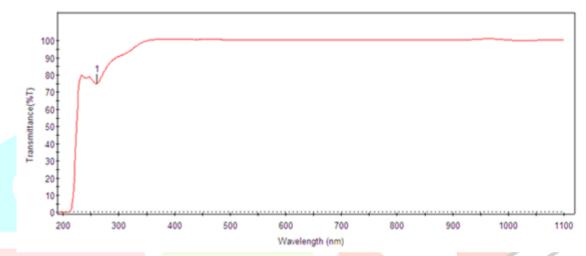
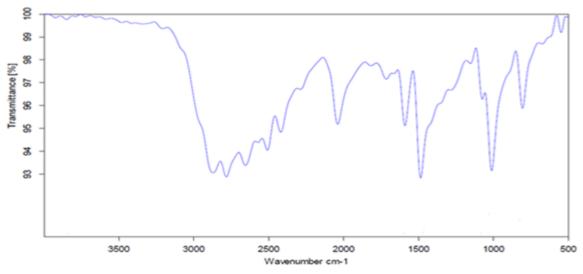
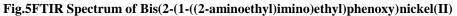


Fig. 4 - Visible Spectrum of Bis(2-(1-((2-aminoethyl)imino)ethyl)phenoxy)nickel(II)

3.3 IR Spectra

The vibrational spectra is valuable information regarding thenature of functional group of the 2-(1-((2-aminoethyl)imino)ethyl)phenotynickel(II). The IR spectra of the Schiff base ligand data showed in **Fig. 5** and **Fig.6**, a broad band in the region $3480-3466 \text{ cm}^{-1}$ indicating the presence of $-\text{NH}_2$ group. On complexation this band isshifted to lower frequency in the range ($339-3440 \text{ cm}^{-1}$) shows the involvement of primary amine nitrogen in coordination tometal ion for all the Schiff base complexes [19]. A sharp peak in the region 1556 cm^{-1} indicating azomethine group and its frequency is increasing 1591 cm^{-1} after complexation The free amine broad band at 3466 cm^{-1} was decreasing to 3446 cm^{-1} [20]. The frequency of 1403 cm^{-1} indicating methyl group of 2-hydroxyacetophenone and which is decreasing after coordination to 1486 cm^{-1} . The aromatic phenolic hydrogen streeting and bending frequency bands shows in the region $3100 \text{ and } 1279 \text{ cm}^{-1}$ [21] were absence and form new sharp band form in the region 548.69 cm^{-1} for confirmatingnickel metal oxygen covelating [22].





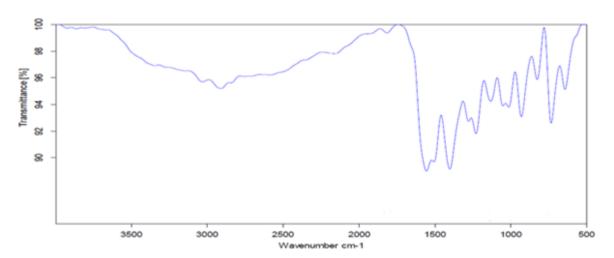


Fig.6 FTIR of 2-(1-((2-aminoethyl)imino)ethyl)phenol

3.4¹HNMR Spectra

The ¹HNMR results show2-(1-((2-aminoethyl)imino)ethyl)phenol and Bis(2-(1-((2aminoethyl)imino)ethyl)phenoxy)zinc(II)in Fig.7 and Fig.8, a sharp absorption singlet peak in the region 16.0 ppm which corresponds to phenolic alcohol of ligand, which peak is absence in Ni (II) complex. This indicates that the deprotated phenolic Oxygen atom is involved in chelation. Four multiplets (2-doublets and 2-triplets) between the region 6.7 and 7.6 ppm, 7.6 (t, 1H) (J=8.1), 7.2 (t, 1H) (J=9.6), 7.2 (d, 1H) (J=3.3), 6.7 (d, 1H) (J=8.1)indicate protons shifts of benzylidene ring of ligand and decrease between 6.6 and 7.2ppm, 7.2 (t, 1H) (J=6.9), 7.0 (t, 1H) (J=14.1), 6.7 (d, 1H) (J=8.7), 6.6 (d, 1H) (J=24.6) for Zn(II) complex. Two triplets between 3.0 and 4.0 ppm, 3.7 (t, 2H) (J=9.0), 2.8 (t, 2H) (J=9.9) indicate aliphatic alkane shifts of ligand and decrease its range after coordination between 2.8 and 3.8 ppm, 3.6 (t, 2H) (J=10.8), 2.5 (t, 2H) (J=10.5). A sharp singlet peak in the range 6.4 which is corresponds to primary amine and decrease 5.11 ppm. This is indicates that coordination bond formed between Nitrogen atom of amine and nickel metal atom. In multiplets, coupling constants are increasing after coordination respectively.

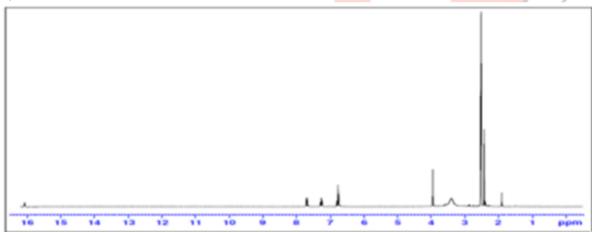
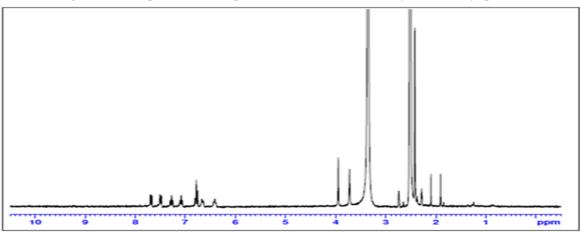


Fig.7 ¹HNMR Spectrum of Preparation of 2-(1-((2-aminoethyl)imino)ethyl)phenol





3.5 ANTIBACTERIAL STUDY

The acetophenone derivative of 2-(1-((2-aminoethyl)imino)ethyl)phenol and Bis(2-(1-((2-aminoethyl)imino)ethyl)phenoxy)zinc(II) complexwere the aim of production and synthesis of antimicrobialcompound is to inhibit the causal microbe without any side effectson the patients. In addition, it is worthy to stress here on the basic idea of applying any chemotherapeutic agent which depends essentially on the specific control of only one biological function and not multiple ones. 2-(1-((2-aminoethyl)imino)ethyl)phenol The acetophenone derivatives of and Bis(2-(1-((2aminoethyl)imino)ethyl)phenoxy)zinc(II) complex involves in bacteriostatic show a remarkable biological activity against differenttypes of Gram-positive (G+ve) and Gram-negative (G-ve) bacteria effect shows in Table 1. The importance of this unique property of the investigated Schiff base complexes lies in the fact that, it can be applied safely in the treatment of infections and some common diseases e.g. septicaemia, gastroenteritis, urinary tract infections and hospital acquired infections. These compounds showed significant antimicrobial activities against Escherchia coli, Proteus mirabilis, Klebsiella pneumonia, Salmonella typhi, Salmonella paratyphi 'A', Pseudomonas aeroginosa and Staphylococcus aereus (G+ve) by well diffusion with serial dilution method up to the lowest concentration $(15.62 \,\mu g)$ from highest concentration $(250 \,\mu g)$. In the higher concentration (250 µg) of the prepared Schiff base and nickel complex against Escherchia coli, Proteus mirabilisand Staphylococcus aereussuppressed by ligand not nickel(II) complex. There is slight moderate growth of inhibition of Klebsiella pneumonia bacteria organism at the concentration of 125 µg of ligand complex and no effects for ligand From the Table.1, both ligand and complex are very good activities against all bacterial strains of gram-positve and gram-negative bacterias in all high to lower concentrations. A strong result of bacterial growth of the Bis(2-(1-((2-aminoethyl)imino)ethyl)phenoxy)nickel(II) have Salmonella typhi, Salmonella paratyphi 'A' microorganisms have at the 125 µg only but inhibited growth of these organisms at the higher concentration (250 µg) and lower concentrations are 62.5, 31.25 and 15.62 µg. Hence, the acetophenon derivativeBis(2-(1-((2aminoethyl)imino)ethyl)phenoxy)nickel(II) in this study could be a potential antibiotic for the enteric fever, typhoid. However, these compounds are 2-(1-((2-aminoethyl)imino)ethyl)phenol and Bis(2-(1-((2-aminoethyl)imino)ethyl)phenoxy)nickel(II) may be used for treating all infectious disease caused by the tested organisms.

Table 1: Minimum Inhibitory Concentrations of 2-(((2-aminoethyl)imino)methyl)phenol and Bis(2-(1-((2-

	Organism/	250) µg	125	μg	62.	5 µg	31.2	5 µg	15.6	2 µg	
	Concentration	L	C	L	С	L	С	L	С	L	С	
	Escherchia coli	-	+	-	-	-	//	1	-	-	-	
	Proteus mirabilis	_	+	-	-	-	1	-	-	-	-	
Ĩ	Klebsiella pneumonia	-	-	-	+/-	-	-	-	-	/		1
	Pseudomonas Aeruginosa	-	-	-	-	-	1	-	-	-		
	Salmonella typhi	-		-	+	-	-	-	1	_	×	
	Salmonella paratyphi 'A'	-	-	1	+	-	-	-	_	3	~	
	Staphylococcus aureus (+	-	+	-	-	-	-/		C	11		
	ve)			0,-		-) 🗂		

aminoethyl)imino)ethyl)phenoxy)zinc(II) against human pathogenic bacteria.

(+) Bacterial growth.

(-) Bacterial Inhibition.

(+/-) Bacterial growth Inhibition

Serial dilution Concentrations from (250 to 15.62)

µg – microgram

L - 2-(((2-aminoethyl)imino)methyl)phenol

C - Bis(2-(1-((2-aminoethyl)imino)ethyl)phenoxy) zinc(II)

3.6 HAEMOLYTIC ASSAY

The acetophenone derivatives are 2-(1-((2-aminoethyl))mino)ethyl)phenol and Bis(2-(1-((2-aminoethyl))mino)ethyl)phenoxy)zinc(II)haemolytic assay result in**Table 2**. There is no toxic effect of both ligand and its complex with human blood sample from highest concentration (250 µg) to lowest concentration (15.62 µg). Here, 10 % sodiumdodcylsulphate as positive control and saline as a negative control. From the table 2 2-(1-((2-aminoethyl))mino)ethyl)phenol and Bis(2-(1-((2-aminoethyl))mino)ethyl)phenol and Bis(2-(1-((2-aminoethyl))mino)ethyl)phenoxy)zinc(II) have there is no toxicity in these tested concentrations and volumes also.

Table 2: Haemolytic Assay of 2-(((2-aminoethyl)imino)methyl)phenol and Bis(2-(1-((2-aminoethyl) imino)ethyl)phenoxy)zinc(II) against human pathogenic bacteria.

Compound / Conc.	250 µg	125 µg	62.5 μg	31.25 µg	15.62 µg	Positive Control	Negative Control
L	-	-	-	-	-	+	-
С		-	-	-	-	+	-

Negative control - Normal saline Positive control - 10% Sodium DodycylSulphate Serial dilution Concentrations from (250 to 15.62) μg – microgram

L - 2-(((2-aminoethyl)imino)methyl)phenol

785

C - Bis(2-(1-((2-aminoethyl)imino)ethyl)phenoxy) zinc(II)

IV. CONCLUSIONS

In the present research studies, our efforts were to synthesis and characterize of aceophenone derivative compounds by condensation method. These synthesized compounds were characterized by various physicochemical and spectral analyses. The compounds were tested against a large number of human pathogenic strains of Gram-negative bacteria including staphylococcus aureus (+ve). Compounds are exhibited good growth inhibition activity against all human pathogenic microorganisms and are promising to act as a potential antimicrobial agent. Based on the results obtained haemolytic assay work proves that cytotoxicity of the prepared 2-(1-((2-aminoethyl))phenol and Bis(2-(1-((2-aminoethyl))phenoxy)zinc(II)no toxic effect lowest concentration (15.62 µg).

ACKNOWLEDGEMENT

The authors thanks to UGC, New Delhi, India for the financial support through the Minor Research Project (**MRP6282/UGC/SERO**) in this batch of 2015-2016.

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