



# ANALYSIS OF LEVEL OF TOTAL CHOLESTEROL IN HUMAN FEMALE SALIVA UNDER DIFFERENT REPRODUCTIVE CONDITIONS AND METABOLIC DISORDERS.

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**Abstract:** Saliva is a complex diagnostic fluid, which contains a variety of enzymes, hormones, antibodies, antimicrobial constituents and growth factors. In our body fluids like blood, saliva, tears, sweat and urine are crucial resources of pathological diagnosis. Changing in level of total cholesterol level in human female saliva was investigated in 60 women volunteers during various stages of reproduction (pre-pubertal, parous, non-parous, menopausal and diabetogenic). According to findings, level of total cholesterol level is significantly increased during ovulatory phase, which was due to hormonal metabolic changes in the period of menstrual cycle.

**Keywords:** Saliva, Total cholesterol, prepubertal, parous, non-parous, menopausal and Diabetogenic.

## Introduction:

Cholesterol is a waxy fat like substance found in cells of human body. It is an organic compound having 27 carbons with hydrocarbon tail, central sterol nucleus made up of four hydrocarbon rings and a hydroxyl group. It belongs to the steroid family and its molecular structure is  $C_{27}H_{46}O$ . It is an essential structural component of animal cells membrane and its major synthesis site is liver. It plays a role in forming and maintaining cell membranes and structures (Krause, 2014). It provides stability and fluidity. Cholesterol also plays a crucial role in regulating cell function (Rahmati et al., 2019, Ding et al., 2019). Total cholesterol includes low density lipoprotein (LDL) cholesterol and high-density lipoprotein (HDL) cholesterol. In human saliva variety of

electrolytes including calcium, magnesium, potassium, cholesterol, fatty acids, triglycerides, glycolipid and nitrogenous products are found (**Marini and Cabassi 2002, Actis et al., 2005, Agha-hosseini et al., 2006 and Caufal et al., 2003**). The concentrations of total lipids in parotid, submandibular and whole stimulated saliva were 0.2, 0.9 and 1.3 mg/dl respectively. Cholesteryl esters, cholesterol, triglycerides, diglycerides, monoglycerides and free fatty acids accounted for 96-99 percent of the total salivary lipids. Total Cholesterol is a direct precursor of steroid hormones, including corticosteroids, androgens, estrogens, progesterone and vitamin D, some of which are produced in the placenta (**Byanes, 2009**). Cholesterol is essential for making a number of critical hormones, including the stress hormone cortisol. According to **Harvard publishing** Cholesterol is also used to make the sex hormones testosterone, progesterone, and estrogen. In women total cholesterol levels rise as estrogen levels increase during the monthly menstrual cycle and drop shortly before ovulation, then decrease more rapidly after ovulation occurs. High Total cholesterol level is one of the primary risk factors for heart disease, the leading cause of death among women (**Lloyd, 2009**). The researchers found that as the level of estrogen rises, high-density lipoprotein (HDL) cholesterol also rises, peaking at the time of ovulation. HDL cholesterol is believed to be protective against heart disease. And when total cholesterol and low-density lipoprotein (LDL) cholesterol levels as well as another form of blood fat known as triglycerides declined as estrogen levels decline. Total cholesterol, LDL cholesterol and triglyceride levels reached their lowest just before menstruation began. **Alagendran et al., 2009** in their study assessed the usefulness of saliva as a biomarker of ovulation detection which showed that saliva can be used to test cholesterol and phospholipids instead of blood. **Al Rawi, 2010 & 2011** did two different studies and compared plasma and salivary lipid profile in individuals with ischemic heart stroke and the diabetes mellitus and suggested that lipid fractions particularly TGL can be assessed in saliva and may be used alone or in combination with other lipid parameters for monitoring disease activity and severity in such studies. The results of our study suggest that saliva can be used to assess, not only TGL but also TC, HDLC, VLDLC and to some extent LDLC. **Alagendran et al., 2009** conducted a study with 50 women between the ages 19 and 40 years and concluded that the lipid and its metabolites undergo consistent variations during the menstrual cycle. With significant review of literature 14 elevations of total cholesterol, low density lipoprotein (LDL), high density lipoprotein (HDL), phospholipids and triglycerides corresponding with peak estradiol levels at ovulation. High total and LDL cholesterol concentration was seen during the ovulatory phase and preovulatory phase of the menstrual cycle. The increased salivary HDL cholesterol levels in preovulatory phase may be attributed to decreased salivary triglycerides and reduced hepatic lipase activity. In a healthy person level of total cholesterol is less than 170 mg/dL. As women and men get older, their cholesterol levels rise. Before the age of menopause, women have lower total cholesterol levels. With high cholesterol, you can develop fatty deposits in your blood vessels. Sometimes, those deposits can break suddenly and form a clot that causes a heart attack or stroke. As earlier report of **Schwartz, 1981 and Mishell and Davajan 1979** indicated that during menopause ovulation fails to occur & female sex hormones diminish rapidly to almost none at all. In this condition non-utilization of cholesterol in steroidogenesis may be possible which increase the total cholesterol level in menopausal saliva in

women. As earlier report of **Meuram et al., 1998** and **Dodds and Dodds 1997** suggested that diabetes caused neuropathic changes in the salivary parenchyma with lymphocytic gland infiltrate. **Ship et al., 2002** also reported reduced salivary flow rate in uncontrolled diabetic patients. It was shown in a recent survey that only one in four women associates menopause with high cholesterol leading to a lack of awareness of the need to consider having cholesterol level checked around the time of the menopause (**Jackson, 2008**). A person's cholesterol levels can increase during or after menopause due to reduced levels of the hormone estrogen in the body. Salivary cholesterol and triglycerides of diabetic patients were estimated lower than that of non-diabetic patients. In contrast, **Priya et al. 2019** found a higher level of salivary lipids (cholesterol and triglyceride) in patients with type-1 diabetic. With the exception of Mycoplasma, which needs cholesterol for growth, it is not present in prokaryotes (bacteria and archaea).

## Materials and Methods

The studies were performed in 60 different human female volunteers of age group (19 to 40 yrs.) categorized as prepubertal, parous, non-parous, menopausal and diabetogenic. Prepare each patient prior to collecting salivary sample. Patients must avoid chewing gum for at least 30 minutes prior to sample collection. In some cases, patients may need to fast overnight for 12-14 hours prior to sample collection.

1. Each patient brushed their teeth thoroughly without toothpaste.
2. Next, each patient had floss.
3. Rinsed patient's mouth with distilled water and sample collected.
4. Stored sample on ice.

**Sample Preparation:** Samples collected from each patient were centrifuged at high speeds and collected the supernatant. For immediate testing, stored supernatant on ice, otherwise stored supernatant at  $-20^{\circ}\text{C}$ . Diluted the supernatant 25 to 100-fold with assay buffer (component B, Cat No. 40006). Added  $50\ \mu\text{L}$ /well of sample to test.

## Results and Discussion:

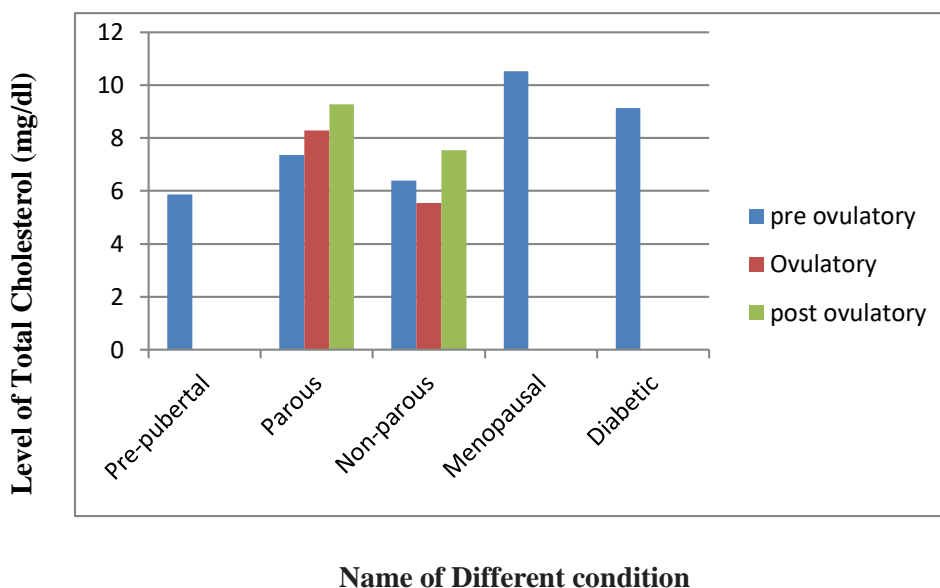
As the findings a highly significant ( $p < 0.001$ ) increased total cholesterol in parous ovulatory, post ovulatory, menopausal and diabetogenic condition in women's saliva was observed whereas highly significant ( $p < 0.01$ ) increased in parous preovulatory and significant ( $p < 0.01$ ) increased in parous preovulatory and significant ( $p < 0.02$ ) increased level in non-parous postovulatory were observed in comparison to prepubertal. It might be due to the active steroid synthesis by ovarian tissue than prepubertal stage. A highly significant ( $p < 0.001$ ) decreased total cholesterol level in non-parous preovulatory & ovulatory than parous preovulatory, ovulatory and post ovulatory was found whereas, highly significant ( $p < 0.01$ ) decreased level were observed in non-parous post ovulatory than parous post ovulatory. In menopausal women's saliva showed a highly significant ( $p < 0.001$ )

increased level than parous preovulatory, ovulatory, postovulatory and non-parous preovulatory, ovulatory & post ovulatory phase.

**Table 1:** Level of Total Cholesterol in saliva of different conditions in human female subjects.

SL No.	Name of different conditions with symbols	Level of total cholesterol (mg/dl) mean $\pm$ SE of 5 samples	P-Value
1	Pre pubertal –(a)	5.86 $\pm$ 0.265	
2	Parous I. Pre ovulatory –(b) II. Ovulatory – (c) III. Post ovulatory–(d)	7.35 $\pm$ 0.105 8.28 $\pm$ 0.076 9.27 $\pm$ 0.258	a to b – (p<0.01) HS a to c – (p<0.001) HS a to d – (p<0.001) HS
3	Non- parous I. Pre ovulatory-(e) II. Ovulatory-(f) III. Post ovulatory-(g)	6.39 $\pm$ 0.167 5.55 $\pm$ 0.153 7.54 $\pm$ 0.425	a to g – (p<0.02) S
4	Menopausal –(h)	10.53 $\pm$ 0.154	a to h – (p<0.001) HS b to h –(p<0.001) HS c to h –(p<0.001) HS d to h – (p<0.01) HS e to h – (p<0.001) HS f to h – (p<0.001) HS g to h – (P<0.001) HS
5	Diabetogenic –(i)	9.14 $\pm$ 0.316	b to i –(p<0.001) HS c to i –(p<0.05) HS e to i – (p<0.001) HS f to i – (p<0.001) HS g to i – (p<0.02) S h to i – (p<0.01) S

Salivary total cholesterol might be possible to reproductive physiological disturbances and steroid hormones synthesis impairment whereas highly significant (p<0.01) decreased level is found than diabetogenic. Diabetogenic women's saliva showed a highly significant (p<0.001) increased total cholesterol than parous preovulatory, non-parous preovulatory and ovulatory phase and a highly significant (p<0.01) decreased level of salivary total cholesterol were observed than menopausal women saliva. A highly significant increased total cholesterol level in parous, non-parous, menopausal and diabetogenic women in comparison to prepubertal saliva might be an indication of its dependence on ovarian steroid hormone synthesis & secretion during ovulation.



### Conclusion:

Lipids are one of the major constituents of the cell. Our body needs some cholesterol to work properly. But if we have too much cholesterol in our blood, we have a higher risk of coronary artery disease. An excessive cholesterol level can lead to cardiovascular diseases such as stroke, hypertension, elevated levels can cause serious problems. With high cholesterol, we can develop fatty deposits in our blood vessels. Eventually, these deposits grow, making it difficult for enough blood to flow through the arteries. Sometimes, those deposits can break suddenly and form a clot that causes a heart attack or stroke. Thus, saliva offers an alternative to serum as a biologic fluid for diagnostic purposes.

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### References:

1. Rahmati-Ahmadabad S, Broom DR, Ghanbari-Niaki A, Shirvani H. 2019. Effects of exercise on reverse cholesterol transport: A systemized narrative review of animal studies. *Life Sci.* May 01, 224, 139-148.
2. Ding X, Zhang W, Li S, Yang H. 2019. The role of cholesterol metabolism in cancer. *Am J Cancer Res.* 9 (2), 219-227.
3. Luo J, Yang H, Song BL. 2020. Mechanisms and regulation of cholesterol homeostasis. *Nat Rev Mol Cell Biol.* Apr, 21(4), 225-245.
4. Krause MR, Regen SL. 2014. The structural role of cholesterol in cell membranes: from condensed bilayers to lipid rafts. *Acc Chem Res.* 47(12), 3512-21.

5. Harvard Health Publishing. How it's made: cholesterol production in your body.
6. Byanes J, Dominiczak M. 2009. *Medical Biochemistry*. 3rd edn Mosby: Elsevier.
7. Lloyd-Jones D, Adams R, Carnethon M et al. 2009. Heart disease and stroke statistics – 2009 update: a report from the American Heart Association Statistics Committee and Stroke Statistics Subcommittee. *Circulation*. 119 (3), 480–486.
8. Alagendran S, Archunan G, Neelamathi E, Anusha R, Miller Samson, and Puspha, N. 2009. Lipid fluctuations in women saliva during Menstrual cycle, *Journal of Cell and Tissue Research*, 9 (2), 1915-1919.
9. Kaufman E, Lamster IB. 2002. The diagnostic applications of saliva: A review. *Crit Rev Oral Biol Med*. 13, 197–212
10. Alagendran S, Archunan G, Prabhu SV, Orozco BE, Guzman RG. 2010. Biochemical evaluation in human saliva with special reference to ovulation detection. *Indian J Dent Res*. 21, 165–8.
11. Al-Rawi NH. 2010. Salivary lipid peroxidation and lipid profile levels in patients with recent ischemic stroke. *J Int Dent Med Res*. 3, 57–64.
12. Rawi NH. 2011. Oxidative stress, antioxidant status and lipid profile in the saliva of type 2 diabetics. *Diab Vasc Dis Res*. 8, 22–28.
13. Jackson G. 2008. Gender differences in cardiovascular disease prevention, *Menopause Int*, 14, 13–17.
14. Saeedi P, Petersohn I, Salpea P, Malanda B, Karuranga S, Unwin N, et al. 2019. Global and regional diabetes prevalence estimates for 2019 and projections for 2030 and 2045: Results from the International Diabetes Federation Diabetes Atlas. *Diabetes Research and Clinical Practice*. 157, 107843.
15. Marini A., Cabassi E. 2002. La saliva: approccio complementare nella diagnostica clinica e nella ricerca biologica. *Ann Fac Med Vet Pharma*, 22, 295-311.
16. L Adriana B. Actis Nilda R. Perovic Daniela Defago Cecilia Beccacece Aldo R. Eynard. 2005. Fatty acid profile of human saliva: a possible indicator of dietary fat intake. *Arch Oral Biol*. 50 (1), 1-6.
17. F Agha- Hosseini, I M Dizgah, S Amirkhani. 2006. The composition of unstimulated whole saliva of healthy dental students *J Contemp Dent Pract*, 7 (2), 104-11.
18. P Coufal, J Zuska, T van de Goor, V Smith. 2003. Separation of twenty underivatized essential amino acids by capillary zone electrophoresis with contactless conductivity detection; electrophoresis. 24, 671-7.
19. Schwartz R, Brunzell JD. 1981. Increase of adipose tissue lipoprotein lipase activity with weight loss. *The Journal of clinical investigation*. May 1, 67 (5), 1425-30.
20. Mishell DR and Davajan V. 1979. eds *Reproductive Endocrinology, Infertility and Contraception*. Philadelphia F-A. Davis Co. 173-179.
21. Meurman, J.H., Collin, H.L., Niskanen, L., Töyry, J., Alakuijala, P., Keinänen, S. and Uusitupa, M. 1998. Saliva in non-insulin-dependent diabetic patients and control subjects: The role of the autonomic nervous system. *Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology, and Endodontology*, 86 (1), 69-76.

22. Dodds, M.W. and Dodds, A.P. 1997. Effects of glyceemic control on saliva flow rates and protein composition in non-insulin-dependent diabetes mellitus. *Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology, and Endodontology*, 83(4), 465-470.
23. Ship, J.A., Pillemer, S.R. and Baum, B.J. 2002. Xerostomia and the geriatric patient. *Journal of the American Geriatrics Society*, 50 (3), 535-543.

