IJCRT.ORG

ISSN: 2320-2882



INTERNATIONAL JOURNAL OF CREATIVE RESEARCH THOUGHTS (IJCRT)

An International Open Access, Peer-reviewed, Refereed Journal

FORMULATION AND EVALUATION OF POLY HERBAL GRANULES AS DIETARY SUPPLEMENT

Prachi N Dighe*1, Ganesh N Kardile1, Vishal L Khilari2, Rutuja A Shinde2

1- Department of Pharmacognosy, PRCOP, Pravaranagar Ahmednagar, MS, India

ABSTRACT

The present research work is based on the formulation of poly herbal granules by incorporating the leaves extract of Curcuma longa, Tinospora cordifolia and Withania somnifera. The turmeric widely uses this plant for treatment of various diseases and disorder. The dried leaves powder of the plant was extracted and subjected to preliminary chemical tests. Then it was formulated and then evaluated for various parameters like angle of repose, bulk density, tab. Density, disintegration time and stability studies. The preliminary chemical studies show that the extract contains carbohydrate, alkaloids, flavonoid, glycoside and protein. The formulated poly herbal granules exhibited excellent flow properties which showed good angle of repose, bulk density and tapped density.

KEYWORDS: Curcuma longa, Tinospora cordifolia, Withania somnifera, poly herbal granules

INTRODUCTION

CURCUMA LONGA

Turmeric, derived from the rhizomes of *Curcuma longa*, is a perennial plant having short stem with large oblong leaves, and bears ovate, pyriform or oblong rhizomes, which are often branched and brownish-yellow in color. Accounting for about 78 percent of world turmeric production, India is the largest producer of turmeric.

Family: Zingiberaceae

Common name: Turmeric, haridra, haldi



Fig. 1: Curcuma longa

TINOSPORA CORDIFOLIA

Tinospora cordifolia is a large glabrous, perennial, deciduous, climbing shrub ofweak and fleshly stem found throughout India.

Family: Menispermaceae

Synonyms:

Sanskrit: Guduchi, Gulvel

English: Heartleaf moonseed



Fig. 2: Tinospora cordifolia

WITHANIA SOMNIFERA

Withania somnifera is a small, woody shrub that grows about two feet inheight. It can be found growing in Africa, the Mediterranean, and India.

Family: Solanaceae

Synonyms:

English: Winter cherry

Hindi: Ashwangadha

Sanskrit: Turangi-gandha



Fig. 3: Withania somnifera

MATERIAL AND METHODS

Plant Material:

Collection and drying:

The botanical staple consisting of three medicinal plants *Curcuma longa* (rhizome), *Withania somnifera* (roots), *Tinospora cordifolia* (leaves), were procured from the authenticated local market of Ahmednagar district.

Dried material was coarsely powdered in grinder and powder was used for extraction.

Authentication:

The identities of the plants were confirmed by the Botany department of sangamner college of arts, science & commerce, Tal: Sangamner dist.: Ahmednagar.

Extraction of plant material:

Method: Soxhelet extraction

Solvents: Using H2O and alcohol.

The collected plant parts were crushed to powder.

The powder was extracted with 7:3 hydro alcoholic mixtures, at 600C temperature, for 6 hrs, in a 500ml round bottom flask. After 6 hrs. of extraction, round bottom flask was cooled to temperature and the extract were filtered and picked up. Decoction was prepared by evaporating the extract to at least one third of its volume.

Decoction was poured onto a glass tray and dried at 100°C.

Dried extract was pulverized and stored in a desicator.



Fig. 4: Extraction process

Preliminary phytochemical screening for various extracts: (Khandelwal, 2005)

Test for carbohydrates:

Molisch test (General test):

Two ml of extract solution was added with few drops of 15 methanolic alpha- naphthol solutions in a test tube and 2ml of concentrated sulphuric acid was added carefully along the side of the test tube. The formation of reddish violet ring at the junction of two layers indicates the presence of carbohydrates. IJCR

Test for reducing sugar:

Benedict's test:

Mix equal volume of Benedict's reagent and extract solution in the test tube. Heat in a boiling water bath for 5 min. solution appears green, yellow or red depending on amount of reducing sugar present.

Fehling's test:

Five ml of extract solution was mixed with 5 ml Fehling's solution (equal mixture of Fehling's solution A and B) and boiled. Development of reddish brown precipitated indicates the presence of reducing sugars.

Test for Alkaloids:

Evaporate all extracts separately. To residue, add dilute HCL. Shake well and filter. Use filtered solution for test.

Dragendroff's test:

2-3 ml test solution and 0.1 ml Dragendroff's reagent was added in test tube.

Formation of orange brown precipitate indicates the presence of alkaloids

Mayer's test:

2-3 ml test solution and 0.1 ml of Mayer's reagent were added. Formation of yellowish buff precipitate indicates the presence of alkaloids.

Hager's test:

2-3 ml test solution and 0.1 ml of Hager's reagent. Formation of yellowish precipitate indicates the presence of alkaloids.

Wagner's test:

2-3ml filtrate with few drops Wagner's reagent gives reddish brown ppt.

Acid test:

Test solution treated with tannic acid solution gives buff colored precipitate.

Test for Flavonoids:

Shinoda test:

To dry powder or extract, add 5 ml 95% ethanol/t-butyl alcohol, few drops conc.HCL and 0.5g magnesium turnings. Orange, pink, red to purple color appears (flavonols, dihydro derivatives and xanthenes).

Sulphuric acid test:

On addition of sulphuric acid (66% or 80%) flavones and flavonols dissolve into it and deep yellow solutions. Chalcones and aurones give red or red-bluishsolutions. Flavones give orange to red colors formed.

Addition of accelerating amount of sodium hydroxide to the residue shows coloration, which decolorizes after addition of acid.

Heat test solution with zinc and HCL, pink to red color is observed.

Test for Tannins:

- To 2-3 ml of aqueous or alcoholic extract, add few drops of following reagents.
- 5% FeCl₃solution: Deep blue-black color.
- Lead acetate solution: White ppt.
- Gelatin solution: White ppt.
- Bromine water: Decolouration of bromine water.
- Acetic acid solution: Red color solution.
- Potassium dichromate: Red ppt.
- Dilute Iodine solution: Transient red color.

Detection of Tannins:

- Solvent system used:
- Toluene: Acetone: Ethyl acetate (3:1:2)
- Ethyl acetate: Formic acid: Acetic acid: water (100:11:11:26)

- Spray reagents:
- 5% FeCl3 in 0.1N HCL. The developed TLC plate was sprayed with reagent, heated at 100°C for 5-10 minutes.
- Color observed: bluish black spots.

Preparation of Poly herbal granules:

Selection of excipients:

Starch was chosen as disintegrant, calcium phosphate dibasic as bulking agent,magnesium stearate as antiadherant, and methyl and propyl parabens as preservatives. To mask the acute bitter taste, sucralose was used as sweetener, citric acid as taste masker.

Formulation of poly herbal granules:

Granules were prepared by using wet granulation technique. Extract (powder) and citric acid were mixed in a mortar to which sucralose were added. This was followed by subsequent addition of starch, calcium phosphate dibasic and thereforethe parabens. Sufficient quantity of distilled water was added to form a lumpy mass which was then passed through sieve no. 22 to form granules. Granules were dried within the oven. Magnesium stearate was added at the end for lubrication.

Table 4. Formulation table

Sr.no	Ingredients	Quantity(mg)	Category
1	Extract	125	
2	Starch	150	Disintegrant
3	Magnesium stearate	2.5	Antiadherant Antiadherant
4	Calcium phosphate dibasic	250	Bulking agent
5	Pearlitol	312.5	Bulking agent
6	Citric acid	125	Taste masker
7	Methyl parabens	2 ml	Preservative
8	Propyl parabens	0.5 ml	Preservative
9	Orange Flavor	Qs	Flavoring agent
10	Sucralose	Qs	Sweetening agent
11	Color	Qs	Coloring agent

Evaluation of poly herbal granules: (Lachman- Liberman 1999, USP 2007)

Angle of Repose:

The angle of repose is that the angle formed by the horizontal base of the bench surface and the edge of a cone-like pile of granules. After the cone from 5 g of sample was built, height of the granules forming the cone (h) and therefore the radius

(r) of the base were measured.

The angle of repose (θ) was calculated as follows: θ = tan-1 (h/r)

Results were only considered valid when a symmetrical cone of powder was formed. The funnel method was used to perform the test.

Loss on drying:

This test was performed by drying a weighed quantity of the product in the oven at 105°C until constant weight was obtained.

Bulk density:

- It is that the ratio of total mass of powder to the bulk volume of powder.
- Db = m / VO
- Where, m: Mass of the blend
- VO: Untapped Volume
- A graduated glass cylinder was used to perform the test

Tapped Density:

Tapped density is the ratio of mass of powder to the tapped volume. Tapped volume is the volume occupied by the same mass of the powder after a standard tapping of a measure.

- Dt= m / Vi
- Where, m: Mass of the blend.
- VI: Tapped Volume
- Graduated glass cylinder was used for the test which was subjected to 50tapping and the volume was noted.

Disintegration time:

The test was performed using a beaker containing simulated saliva fluid maintained at 37 °C for evaluating fast disintegration. The formulation was added to it and the disintegration time was noted.

Stability Studies:

The success of an effective formulation can be evaluated only through stability studies. The purpose of stability is to obtain a stable product which assures its safety and efficacy up to the end of shelf life at defined storage conditions and peak profile. The optimized formulation of the drug was subjected to accelerated stability studies at specified conditions of temperature and relative humidity of 25°C/60% RH, 30°C/60% for 3 months. After the completion of three month the samples were analyzed visually for any color changes due to physical and chemical interaction within excipients and with the drug. (ICH guidelines1993).

Screening of Polyherbal granules for antioxidant activity:

Formulated poly herbal granules were evaluated for its antioxidant action using DPPH radical scavenging method:

DPPH assay (2, 2-Dipheny l-1-Picrylhydrazyl):

The free radical scavenging activity of extract was determined by using DPPHassay. The decrease in the absorption of the DPPH solution after the addition of an antioxidant was measured at 517nm. Ascorbic acid (50mg/ml) in methanol was used as reference standard.

Principle:

2,2 Diphenyl 1,1- Picryl Hydrazyl is a stable (in powder form) free radical withred color which turns yellow when scavenged. The DPPH assay uses this character to show free radical scavenging activity.

Procedure:

Evaluation of antioxidant activity by DPPH radical scavenging method Free radical scavenging activity of extract were measured by 1, 1- diphenyl-2-picryl hydrazyl (DPPH). In brief, 0.1 mM solution of DPPH in ethanol was prepared. This solution (1 ml) was added to 3 ml. of different extracts in ethanol at different concentration (25, 50,100 µg/ml). Here, only those extracts are used which are Solubilise in ethanol and their various concentrations were prepared by dilution method. The mixture was shaken vigorously and allowed to stand at room temp for 30min. then, absorbance was measured at 517 nm. By using spectrophotometer (UV- VIS Shimadzu). Reference standard compound being used was ascorbic acid. The IC 50 value of the sample, which is the concentration of sample required to inhibit 50% of the DPPH free radical, was calculated using formula. Lower absorbance of the reaction mixture indicated higher free radical activity.

The percent DPPH scavenging effect was calculated by using following equation:

Percent inhibition = $A0 - A1 / A0 \times 100$.

Where, A0 was the Absorbance of control reaction

A1 was the Absorbance in presence of test or standard sample.

EXPERIMENTAL RESULTS

Phytochemical screening:

Table 5. Preliminary Phytochemical Screening

Type of phytoconstituents	Inference	
Carbohydrates	+	
Alkaloids	+	
Tannins	+	
Flavonoids	+	

Excipients profile:

Starch takes up water from the body fluids which cause it to swell and thereby leading to disintegration of the granules. Calcium phosphate dibasic used as bulking agent. Pearlitol also aids in faster disintegration and acts as a non-calorific sweetening agent. Magnesium stearate helps to prevent attrition between the granules and formation of fines. Methyl and propyl parabens are nontoxic, non-irritating and are used in combination to prevent decomposition of the formulation. Citric acid helps to stimulate salivary secretions and hence leading to disintegration of the granulesin the oral cavity, thereby obviating the need to consume water along with the formulation.

Sucralose and the flavoring agent help to mask the bitter taste of extract. As they are triturated with the drug at the very beginning of the preparation before the addition of other excipients, they form a coating over the drug particles and hence in spite of disintegration within the oral cavity it makes the formulation highly palatable.

Evaluation of Polyherbal granules:

The values of angle of repose are below 30° thereby indicating excellent flow properties. Lower values of bulk and tapped density indicate higher porosity implying the time required for disintegration would be lower. % LOD test values comply with the official limits and indicate lower moisture content in the formulation. The disintegration test implies that the granules can disintegrate within 15 sec, thereby leading to quicker absorption and onset of action of the drug as compared to that in its other dosage form

Stability study discussion:

After the three months of completion the samples were analyzed visually for any color changes and hence no color changes were observed due to physical and chemical interaction within excipients and with the drug.

Table 6. Evaluation of Polyherbal granules

Parameters	Results
Angle of repose	26.38
Loss on drying	0.355
Bulk density	0.498
Tapped density	1.20
Disintegration time	Within 15 sec
Stability study	Stable

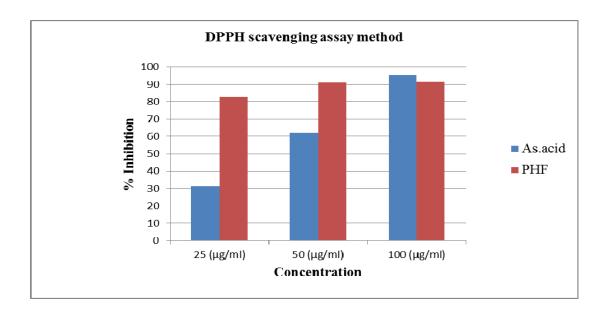
Screening of polyherbalgranules for antioxidant activity:

Total antioxidant percent:

Absorbance of with standard as corbic acid at 517 nm by UV visible spectrophotometer (dpph scavenging assay method)

Table 7. Total antioxidant percent

Sr. no	Co <mark>ncentrat</mark> ion	Ascorbic	acid	Polyherbal formulation
	(µg/ml)	(%inhibition)		(%inhibition)
1	25	31.22 %		82.56 %
2	50	61.95 %		90.97 %
3	100	95.12 %		91.34 %



REFERENCES

- 1) Junaid Niazi, Priyanka Poonia, Vikas Gupta, Narindenpal Kaur. Pharmacotherapeutics of Curcuma longa-A potent patent.International Journal of Pharma Professional's Research, 2010;1(1):24-33
- 2) Nilanjana Deb, Purba majumbar, Ajay Kumar Ghosh.Pharmacognostic and Phytochemical Evaluation of the Rhizomes of Curcuma Longa Linn. Journal of Pharamasci-Tech, 2013; 2(2):81-86.
- 3) C.K.Kokate, A.P.Purohit, S. B. Gokhale. Pharmacognosy.Nirali Prakashan, 2014; 49:14.132-134.
- 4) Seema Kaur P. comparative study of pharmacognostical and preliminary phytochemical investigation of Curcuma longa leaves and rhizomes. Pharma Tutor, 2016; 4(10):31-36.
- 5) K.M.Nadkarni. Indian plants and drugs. Jay book service, 2010; 1:271-273.
- 6) M.A.Pawar, S.S. Patil, D.M. Nagrik. Phytochemical and physicochemical investigation of Curcuma longa Linn rhizome. International Journal of Chemical and Physical Sciences, 2015; 4:458-463.
- 7) R.S. Sawant and A.G. Godghate. Qualitative phytochemical screening of rhizome of Curcuma longa limns. International Journal of Science Environment, 2013;2(4):634-641.
- 8) Shilika Shetty and K.K Vijay, A. laxmi. Phytochemical investigation of extract /solvent fraction of Piper nigrum Linn. Seeds and Piper betle Linn. Leaves. International Journal of Pharma and Bio Science, 2012; 3(2):344-349.
- 9) V. Jyotiprabha and Venkatachalam.Preliminary phytochemical screening of different solvent extract of selected Indian spices. International Journal of Current Microbiology and applied sciences, 2016; 5(2):116-122.
- 10) Debjit Bhowmik, Chiranjib, K.P. Sampat Kumar, Margret chandira, B. Jayakar. Turmeric:A

herbal and traditional medicine. Archives of Applied Science Research, 2009;1(2):86-108.

- M. Rasool, P. Varalakshmi. Immunomodulatory role of Withania somnifera 11) root
- Powder on experimental induced inflammation: An in vivo and in vitro study. Vascul. 12) Pharmacol.
- 13) S.K. Kulkarni, I. Ninan. Inhibition of morphine tolerance and dependence by
- Withania somnifera in mice. J. Ethnopharmacol. 14)
- M. Ganzera, M.I. Choudhary, I.A. Khan. Quantitative HPLC analysis of with anolides 15)
- In Withania somnifera. Fitoterapia M.A. Weiner, J. Weiner. Ashwagandha (India ginseng). 16)

In: Herbs that Heal. Quantum Books, Mill Valley

- Somasundaram, J. Sadique, A. Subramoniam. Influence of extra-intestinal 17)
- 18) Inflammation on the in vitro absorption of 14C-glucose and the effects of antiinflammatory
- Drugs in the jejunum of rats. Rana V, Thakur K, Sood R, Sharma V, Sharma TR. Genetic 19) diversity analysis of Tinospora cordifoliagermplasm collected from northwestern Himalayan region of India. J Genet. 2012; 91:99–103
- Gupta R, Sharma V. Ameliorative effects of Tinospora cordifolia root extract on 20) histopathological and biochemical changes induced by aflatoxin-b (1) in mice kidney. Toxicol Int. 2011; 18:94–8.
- Kapil A, Sharma S. Immunopotentiating compounds from Tinospora cordifolia. J 21) Ethnopharmacol. 1997; 58:89-95.
- Kim SK, Kim HJ, Choi SE, Park KH, Choi HK, Lee MW. Anti-oxidative and inhibitory 22) activities on nitric oxide (NO) and prostaglandin E2 (COX-2) production of flavonoids from seeds of Prunus tomentosa Thunberg. Arch Pharm Res.
- Swaminathan K, Sinha UC, Bhatt RK, Sabata BK, Tavale SS. Structure of tinosporide, a 23) diterpenoid furanolactone from Tinospora cordifolia Miers. Acta Crystallography C. 1989; 45:134–6.
- Afolabi, S.O., Akindele, A.J., Awodele, O., Anunobi, C.C. and Adeyemi, O.O. 2012. A 90 day 24) chronic toxicity study of Nigerian herbal preparation DAS-77 in rats. BMC Complementary and Alternative Med., 12: 79.
- 25) World Health Organization (1998). Quality control methods for medicinalplant material.
- S.S Khadabadi, S.L Deore and B.A Baviskar. Experimental Phytopharmacognosy, A 26) Comprehensive Guide. Nirali Prakashan India. Preliminary Chemical evaluation.2011
- Ayur shop, India store. Adulsa Compund 100ml am rut Pharmaceuticals. Indian pharmacopoeia 27) 2007. Volume 3
- Rakhi B. Shah, Mobin A.; Comparative Evaluation of Flow for Pharmaceutical Powders and 28) Granules; AAPS PharmSciTech,
- FatKatzhendler, ReuvenAzoury, Michael Friedman. Crystalline properties of Carbamazepine 29) in sustained release hydrophilic matrix-tablets based on hydroxyl propyl methyl cellulose. Journal of

controlled release;

- 30) Basak SC, ShrinivasaR, Manavalan. Controlled release HPMC matrix tablet of propranolol HCl. Indian J. Pharm. Sci. 2004
- 31) NIH. U.SNational Library of Medicine. Daily med. Montelukast Sodium-montelukast sodium granule.
- 32) Ajazuddin and S. Saraf. Applications of novel drug delivery system for herbal formulations. Fitoterapia. Volume 81, Issue 7, October.
- 33) Watson RR, Gerald JK, Prredy VR (Eds). *Nutrients, Dietary Supplements, and Neutraceuticals*. *Cost Analysis versus Clinical Benefits*. Springer Science & Business Media, Berlin, 2011.
- 34) Cupp MJ, Tracy TS (Eds). *Dietary Supplements. Toxicology and Clinical Pharmacology*. Humana Press, New York, 2003.
- 35) Begins K, Kreft S (Eds). *Dietary Supplements. Safety, Efficacy and Quality*. Woodhead Publishing (inprint of Elsevier), Amsterdam, 2014.
- 36) Bjelakovic G, Nikolova D, Gluud C. Antioxidant supplements to preventmortality. *JAMA* 310, 1178-1179, 2013.
- Watson RR (Eds). *Nutrients, Dietary Supplements and Neutraceuticals. Cost Analysis Versu Clinical* Benefits, Nutrition and Health. Springer-Science, Heidelberg, Berlin, New York. 2011.
- 38) Bergins K, Kreft S (Eds). *Dietary Supplements. Safety, Efficacy and Quality.* Woodhead Publishing inprint of Elsevier, Amsterdam, New York, 2014.
- Joshi S. Vitamin supplementation in the elderly. *Clin Geriatr Med* 31(3), 355-266, 2015
- 40) Kähkönen, M.P.; Hopia, A.I.; Heikki, J.V.; Rauha, J.P.; Pihlaja, K.; Kujala, T.S.; Heinonen, M. Antioxidant activity of plant extracts containing phenolic compounds. *J. Agric. Food Chem.* **1999**, 47, 3954–3962
- 4l) Blois, M.S. Antioxidant determinations by the use of a stable free radical. *Nature* 1958.
- 42) Proestos, C.; Chorianopoulos, N.; Nychas, G.J.E.; Komaitis, M. RP-HPLC analysis of the phenolic compounds of plant extracts. Investigation of their antioxidant capacity and antimicrobial activity. *J. Agric. Food Chem.* **2005**, *53*, 1190–1195
- 43) Gokhale, S.B., Kokate C.K., Purohit, A.P. Pharmacognosy. 37th ed. Nirali Prakashan, 2008. pp.581.
- 44) Khandelwal K.R., Practical Pharmacognosy. Techniques and Experiments. Nirali Prakashan, pp. 15-29, 149-56. 2007. 13th Edition