

EFFECT OF DUAL INOCULATION OF AM FUNGI AND *Rhizobium* ON THE DROUGHT TOLERANCE IN BLACK GRAM (*Vigna mungo* L.) VAR VBN4

¹D. Ananda kumar, ²K.Sivakumar and ³S.Chitharanjan

Department of Agricultural Microbiology, Faculty of Agriculture, Annamalai University, Annamalai Nagar – 608 002, Tamilnadu, India

Abstract

Biofertilizers are the microbial inoculum which enhanced the soil fertility and crop yield. This study was initiated with the interaction of AM Fungi, Rhizobium and drought stress on the growth of black gram (*Vigna mungo* L.) var VBN4. The dual inoculation of AM fungi and Rhizobium showed the synergistic effect on morphological features and physiological tolerance of such legume species under green house condition with few days of drought condition. The efficiency of such biofertilizers in dual and individual applications on the quantitative yield of chickpea plant was comparatively higher than the non-inoculated control plants. This study has suggested the dual application of such biofertilizers not only enhanced the legume plant growth but also increased the soil fertility, drought tolerance and reduced the risk of application of chemical fertilizers in the agricultural field. Thus, they are generally termed as eco-friendly fertilizer and do not cause the pollution of any sort.

Key words: *Rhizobium*, *AM fungi*, *Drought Stress*.

I. INTRODUCTION:

AM fungi is the most abundant kind of mycorrhizae found in association with every taxonomic group of plants and the list of species not infected is probably far shorter than the infected ones. These fungal associations are beneficial to crop plants in many ways, including enhancing the nutrient availability especially phosphorus, enhancing water uptake, inducing resistant against diseases and increasing the yield (Lekberg and Koids, 2005). Aher *et al.* (2007) compared the efficiency of four VA-mycorrhizal fungi on *Pennisetum typhoides*. Among the mycorrhizal fungi, *G. fasciculatum* and *G. mosseae* were the most effective in increasing the shoot and root growth, dry weight, per cent infection over control. Natural soil offers consortium of indigenous mycorrhizal fungi and often used as source of inoculum. This can be produced on a large scale by pot culture technique. Since isolation and selection of AM species (effective for growth promotion) and rising of pure culture of these species is difficult, a suitable host is required to maintain the AM culture. The beneficial use of AM inoculum in agriculture and raising nurseries has been reported (Muthukumar *et al.*, 2001).

The legume-*Rhizobium* symbiosis is a classic example of mutualism where rhizobia supply ammonia or amino acids (as dicarboxylic acids – malate and succinate) as a carbon and energy source. Different *Rhizobium* sources are *Rhizobium meliloti*, *Rhizobium trifolii*, *Rhizobium leguminosarum* and *Rhizobium phaseoli*. Artificial inoculation helps to increase the native population of a particular species in the soil when that crop is grown. India is the largest producer and importer of the leguminous crop (Shakya *et al.*, 2008).

It is interesting to note that interaction of AM fungi with other beneficial micro organisms increases the growth of plants by enhancing the uptake of minerals especially phosphate. This AM fungi confer other benefits to their host plant such as drought tolerance, protecting the host against diseases, salinity and temperature extremes, producing plant growth hormones and movement of carbohydrates from one cell to another. The wide spread presence AM symbiosis in nodulated legumes and the role of AM fungi in improving nodulation and rhizobial activity within the nodules, are both universally recognized processes (Barea *et al.*, 2005b). The dual application of *Rhizobium* and VAM showed synergistic effect on all mung bean cultivars. Among form cultivars tested variety “Vaibhav” was found host responsive to root nodulation, growth parameters and grain yield (Manke *et al.*, 2008).

Among these, drought stress is particular importance, the former showed water deficit which refers to situation in which plant water potential and turgor are reduced to interfere the normal functioning of plants and the later showed homeostasis disruption in water potential and ion distribution. This has lead to research into drought

tolerance with the aim of improving crop plants inoculated with biofertilizers. This study was initiated to know the interaction of biofertilizers (*Rhizobium* and AM Fungi) and drought stress on the growth of chickpea (*Vigna mungo* L.). Such work was an ecofriendly beneficial to all in some way.

Material and methods:

Legume plant grown under green house condition:

Seeds of black gram (*Vigna mungo* L.) var VBN4 was surface sterilized with 0.1% mercuric chloride for 5min and washed with sterile water repeatedly. Sterile field soil was field to fill the cement pots (30cm height; 45 cm diameter). About 10 kgs of soil were. Ten seeds were sown in each pots. After germination the seedlings were thinned out to 6 in each pot. All experimental plants were maintained in the green house under condition of broad day light. Sterile tap water was used to water the plants.

Treatments :

T1-Control

T2- Single inoculation of *Rhizobium*

T3-Single inoculation of AM Fungi

T4-Dual inoculation of AM Fungi + *Rhizobium*

Inoculation with AM Fungi:

The selective AM Fungal inoculum was mass cultured in the host maize plants in plants in sterile soils under potted conditions. Five gram of soil inoculum with AM Fungal spores and sporocarps and infected roots bits were spread over the lower layer of soils (2 kgs) Then 8 kgs of soils was layered over the inoculum before sowing.

Inoculation of *Rhizobium* :

The selective *Rhizobium* inoculum was mass cultured under laboratory conditions. The carrier based inoculum was mixed thoroughly to form inoculum slurry and was mixed with seed properly .then such seeds were shade dried and the *Rhizobium* (10^9).

Induction of tolerance:

Drought tolerance was given by withholding water supply from 31th day to 35th day for 5 days

Determination of growth :

The legume plants vegetative growth was measured for the following parameters at regular interval of 15 days.

Determination of fresh and dry mater:

The plant materials were cut into bits and weighed. Then they were and dried in a dried in an oven at 90°C until the weight became constant.

Shoot and root length determination:

The shoots and root lengths of the plants were measured using a meter scale.

Determination of root nodule number:

The number of root nodules per plants were collected and counted in all bio-inoculated legume plants.

Assessment of AM Fungal infection:

The root materials of AM Fungi treated legume plants were cleared and stained using the improved procedure of Phillips and Hayman (1970).

Chlorophyll estimation :

The chlorophyll content of leaf tissue was estimated by the methods of Arnon (1949).

Carotenoid estimation:

The carotenoids content of the leaf tissue was estimated by Ridley's (1977) method.

Proline estimation:

The free proline content in the leaves of black gram was determined by the methods of Bates *et al.*, (1973).

Total nitrogen estimation :

The total nitrogen content of the dried biomass of black gram was estimated (Umbreit *et al.*, 1972) by micro kjeldahl method.

Total Phosphorous estimation:

The total potassium content of the dried biomass of blackgram was estimated (Bartlett,1959) by micro kjeldahl method.

Total potassium estimation:

The total potassium content of the dried biomass of black gram was estimated by flame photometer methods.

Estimation of proteins:

The protein content of fresh leaf tissue and dried seed was estimated by Lowry's method (Lowry *et al.*, 1951).

Statistical analysis:

The data collected in this study was subjected to analysis of variance (ANOVA) and means comparison has done using Duncan's multiple range test (DMRT) (Little and hills,1978).

Results and Discussion:

Legumes are consumed as a source of human food and animal feed. Their importance as food lies primarily in their high protein content. Its grain protein is the natural supplement to cereal grain protein. They also provide fat and carbohydrates. Moreover, legumes are high in bone building minerals and vitamins essential for good health (Porres *et al.*, 2003). Biofertilizers are inputs containing microorganisms which are capable of mobilizing nutritive elements from non-usable form to usable form through biological processes; they include mainly the nitrogen fixing, phosphate solubilizing and plant growth promoting microorganisms (Goel *et al.*, 1999). Sharma *et al.* (2005) addressing the effect of legume rhizo deposition on bacterial communities, showed a distinct plant-dependent rhizosphere effect on the distribution of different bacterial groups present in legume rhizosphere. In the present study, the dual mixture of two biofertilizers (*Rhizobium* and AM fungi) was significantly more effective than single species inoculum. The fresh weight of the black gram was gradually increased from 1.10g to 5.30g and dry weight 0.35g to 2.60g in *Rhizobium* with AM fungi inoculated plants when compared with control and individual inoculated plants (Table 1).

Lin *et al.* (1993) have investigated that the mycorrhizal inoculation with *Rhizobium trifolii* on *Trifolium repens* significantly increases the dry weight of shoots and roots, nodulation, nitrogen fixation, total nutrient uptake, final dry matter and phosphorus absorption. Drought stress is one of the major abiotic stresses limiting the productivity of crops in agriculture worldwide (Bohnert *et al.*, 1995). It is also a significant yield-limiting factor in

black gram production as the major black gram growing areas are in arid and semi-arid zones and about 90% world's black gram is grown under rain fed conditions (Kumar and Abbo, 2001). Blackgram showed the mechanisms for overcoming this condition. The present study showed that the mild drought stress in black gram plants had little effect on fresh and dry matter yield of both individual and combined inoculation of *Rhizobium* and AM fungi treated legume plants and control plants. However, the dry matter yield during the recovery period (35 to 45 days) was remarkable in the mycorrhizal plants. These results were positively related to the inoculation effect of single and dual inoculation with *Gigaspora rosea*, *Glomus intraradices* with *Gigaspora rosea* and *Glomus etunicatum* with *Glomus intraradices* on the growth and nutrients uptake (NPK) on *Medicago sativa*. It showed the significant increase in dry weight of shoot and root (Khan *et al.*, 2008).

The present investigation reported that the shoot and root length of the control and all biofertilizers treated black gram plants increased progressively with age. Significant increase in shoot and root length was found in *Rhizobium* with AM fungi inoculated as compared to the other treated black gram plants (Table-1). The response of arbuscular mycorrhizal fungi and *Rhizobium* inoculation on the growth and chlorophyll content of *Vigna unguiculata* (L) Walp Va. Pusa 151 was investigated (Arumugam *et al.*, 2010) and recorded as a significant increase over control in root length (45.6 cm), shoot length (12.2 cm), dry weight of root (0.4 g) and shoot (1.8 g), total number of nodules (39.6 nos.), dry weight of nodules (0.5 g), percentage of mycorrhizal infection (96.6%), chlorophyll *a* (0.83 mg/g fr.wt), chlorophyll *b* (1.19 mg/g fr.wt) and total chlorophyll (2.24 mg/g fr.wt) in dual inoculated (AM Fungi and *Rhizobium*) plants than plants with individual inoculation. The number of root nodules was higher in *Rhizobium* with AM fungi treated black gram (15±2.0 in numbers.) plants than the AM fungi and *Rhizobium* alone treated and control plants. The biofertilizers (*Rhizobium* and AM fungi) inoculated legumes showed normal growth period under drought stress. The wide spread presence of the AM symbiosis in nodulated legumes and the role of AM fungi in improving nodulation and rhizobial activity within the nodules, are both universally recognized processes (Barea *et al.*, 2005b). Colonization of a legume by AMF can increase the number of nodules (Garg and Manchanda, 2008). This may indicate a positive influence of AMF on legume-nitrogen fixing bacterial symbiosis. The present study revealed that the AM fungal colonization in roots of black gram was maximum as 98 in *Rhizobium* with AM fungi inoculated plants and was minimum (89) in AM fungi treated black gram plants. Drought stress does not affect the percent of AM infection in both single and dual inoculation with *Rhizobium* treated plants. The presence of oval, round or irregularly lobed vesicles occurring between or inside cortical cells, attached to hyphae and containing oil globule was a sign of AM fungal infection in black gram plant roots.

These vesicles were act as storage structures. The presence of arbuscules in the infected roots are intended to serve as two way channels for transport of nutrients, more particularly carbohydrates structures known as appressoria connect AM fungal ramifications inside roots with the mycelium of the fungus outside the root and serve as absorbing elements from soil to roots. In the present study, the AM fungi either alone or in combination with *Rhizobium*, caused about significant difference in chlorophyll *a*, *b* and total chlorophyll content (73.60±0.30µ g; 34.40±0.37µ g and 104.00±0.50µ g) was noticed at 50th day period of black gram. But these contents were reduced during drought and salt stress condition in all plants. However on re watering, the biofertilizers inoculated legumes recovered fast in comparison with the control plants. Such results were previously reported by Manivannan *et al.*, 2007. The decrease in chlorophyll under drought stress is mainly the result of damage to chloroplasts caused by active oxygen species (Simirnof, 1995). The carotenoid contents of black gram showed minimum increase in biofertilizers treated plants compared to control and was observed maximum during drought stress period.

During recovery period, the carotenoid content declined progressively in both control and biofertilizers inoculated drought stressed plants. Plants can partly protect themselves against mild drought stress by accumulating osmolytes. Proline is one of the most common compatible osmolytes in drought stressed plants. For, example, the proline content increased under drought stress in Pea (Alexieva *et al.*, 2001). Proline accumulation might also be a part of the stress signal influencing adaptive responses (Maggio *et al.*, 2002). Its metabolism in plants, however, has mainly been studied in response to osmotic stress (Verbruggen and Hermans, 2008). It induced accumulation of

soluble sugars and proline in two maize varieties was studied (Mohammadkhani and Heidari, 2008). Proline accumulation may also be part of the stress signal influencing adaptive responses (Maggiio *et al.*, 2002). This accumulation has been advocated as a parameter of selection for stress tolerance (Jaleel *et al.*, 2007). The effect of drought stress on yield, proline and chlorophyll contents in three black gram cultivars was studied (Mafakheri *et al.*, 2010).

Table 1: Effect of Dual inoculation of AM Fungi and *Rhizobium* on the drought tolerance in black gram(*Vigna mungo L.*) var VBN4

| Treatment | Growth parameter | 15 DAS | 30 DAS | 35DAS | 50 DAS |
|--|------------------------|--------|--------|-------|--------|
| T1-Control | Shoot length(cm/plant) | 7.5 | 16.80 | 17.50 | 23.90 |
| | Root length (cm/plant) | 3.20 | 8.20 | 8.35 | 11.50 |
| | Fresh weight(g/plant) | 0.60 | 2.10 | 2.12 | 3.55 |
| | Dry weight(g/plant) | 0.12 | 0.25 | 0.26 | 1.05 |
| T2-Single inoculation Of <i>Rhizobium</i> | Shoot length(cm/plant) | 7.90 | 17.80 | 18.50 | 26.00 |
| | Root length (cm/plant) | 3.70 | 9.36 | 9.80 | 12.50 |
| | Fresh weight(g/plant) | 0.65 | 2.14 | 2.16 | 3.85 |
| | Dry weight(g/plant) | 0.15 | 0.30 | 0.30 | 1.40 |
| T3-Single inoculation Of AM Fungi | Shoot length(cm/plant) | 9.90 | 25.00 | 26.50 | 33.50 |
| | Root length (cm/plant) | 4.35 | 11.30 | 11.65 | 16.10 |
| | Fresh weight(g/plant) | 0.65 | 2.40 | 2.44 | 4.90 |
| | Dry weight(g/plant) | 0.14 | 1.20 | 1.24 | 2.10 |
| T4-Dual inoculation Of <i>Rhizobium</i> and AM Fungi | Shoot length(cm/plant) | 10.12 | 26.90 | 28.82 | 36.20 |
| | Root length (cm/plant) | 4.70 | 11.60 | 13.00 | 19.20 |
| | Fresh weight(g/plant) | 1.10 | 2.60 | 2.69 | 5.30 |
| | Dry weight(g/plant) | 0.35 | 1.27 | 1.30 | 2.60 |

In the present investigation the proline content in the control and biofertilizers inoculated black gram plant was minimum at all stages of the plant. After drought stress period, its level declined sharply in both control and biofertilizers inoculated plants. This proline accumulation in both legume plants was stimulated by the biofertilizers under mild drought conditions. Further, the diffusion of proline after rehydration of legumes might be taken to indicate that proline served as a storage compound during stress. The leaf protein content was increased gradually in black gram of both control and biofertilizers treated plants as on plant age. This content was declined during the drought stress. After its recovery, it was observed maximum (8.82 ± 0.07 ; 8.96 ± 0.04 mg) as in dual inoculation of *Rhizobium* and AM fungi treated plants compared to control (3.68 ± 0.04 ; 4.78 ± 0.05 mg) plants. After harvesting, the seed protein content of black gram was estimated and it was significantly higher (<31%) in *Rhizobium* and AM fungi treated Plant than the control (20%). But in *Rhizobium* and AM fungi the protein content was 23% and 26%. Generally, in legumes the AM fungi increased nodulation and nitrogen fixation as a consequence of improved phosphorus nutrition (Athar, 2005). Many researchers have reported enhancement of phosphate uptake and growth of leguminous plants by vesicular arbuscular mycorrhizal fungi (Atimanav and Adholeya, 2002). The present investigation revealed that the black gram plant's total nitrogen, phosphorus and potassium contents were higher in biofertilizers inoculated plants than the control, at all stages of growth. Drought stress did not affect the nitrogen accumulation in *Rhizobium* and AM fungi at both individual and dual inoculated plants.

There was an increase in total phosphorus and potassium content was found in the mycorrhizal plants followed by *Rhizobium* inoculated plants even during the drought stress period Morte *et al.* (2000) have explained that mycorrhizal plants have accumulated more potassium in shoots and roots than control plants under both normal irrigation and drought – stress conditions. The main advantage of mycorrhiza is its greater soil exploration and increasing uptake of nitrogen, phosphorus, potassium, zinc, copper, sulphur, iron, calcium, magnesium and manganese supply to the host roots (Malik, 2000). The *Glomus etunicatum* inoculated maize plants in sandy loam soil, under water stressed conditions, absorbed more phosphorus than non-mycorrhizal plants (Muller and Hofwer, 1991). To the well-known positive impacts of AM fungi on plant yields such as a better survival rate of colonized plants, the maintenance of plant biodiversity, the improvement of soil microflora (Boer *et al.*, 2005), the resistance to biotic (Dalpe, 2005) and abiotic environmental stresses (Neumann and George, 2009), the improvement of soil structure and reduction of pesticide use (Strack *et al.*, 2003).

Conclusion

Arbuscular mycorrhizal fungi are ubiquitous in soil habitats and form beneficial symbiosis with the roots of angiosperms and other plants. Most terrestrial plants associate with root colonizing mycorrhizal fungi, which improve the fitness of both the fungal and plant associates. *Rhizobium* is a symbiotic nitrogen fixing bacteria applied as biofertilizers in agricultural crop improvement and management. They played a major role in the plant metabolism and crop productivity. The present study experimentally reported that the dual inoculation of AM fungi and *Rhizobium* could help the growth and yield of black gram plants. The results from this study showed that AM fungal symbiosis could protect such legume plants against the mild drought stress. It could be confirmed that the mycorrhizal plants were resistant to mild drought stress than control plants. Crop plants with dual symbiotic association possess both nutritional and ecological advantages to compensate nutrient deficient situation, and the establishment of these association can be improved by inoculation with its mutualistic partners *Rhizobium* and arbuscular mycorrhizal fungi. This study suggested the both biofertilizer (AM fungi and *Rhizobium*) application in agricultural fields of economically important crops can yield higher quantity and quality of seeds and also these plants showed mild drought tolerance when compare to non-biofertilizer inoculated crops. They were considered as environment friendly fertilizers and do not cause any kind of pollution for a generation to come.

REFERENCES

- Aher, K. 2007. Mass multiplication of *Glomus fasciculatum* using different hosts Biofertilizer Newsletter, 3(4): 85.
- Alexieva V, Sergiev, I., Mapelli, S. and Karanov, E. 2001. The effect of drought and ultraviolet radiation on growth and stress markers in pea and wheat. Plant cell Environ, 24: 1337-1344
- Arnon, D.I. 1949. Copper enzymes in isolated chloroplasts. Polyphenol oxidase in *Beta vulgaris*. Plant Physiol. 24:1-11
- Arumugam, R., Rajasekaran, S. and Nagarajan, S.M. 2010. Response of Arbuscular mycorrhizal fungi and *Rhizobium* inoculation on growth and chlorophyll content of *Vigna unguiculata* (L) Walp Var. Pusa 151. J. Appl. Sci. Environ. Manage., 14(4):113-115.
- Athar, M. 2005. Nodulation of native legumes in Pakistani range lands. Agric. Conspect. Sci., 70:49-54.
- Atimanav, G. and Adholeya, A. 2002. AM inoculations of five tropical fodder crops and inoculum production in marginal soil amended with organic matter. Biol. Fertil. Soil., 35:214-218.
- Barea, J.M., Werner, D., Azcón-Aguilar, C. and Azcón, R. 2005b. Interactions of arbuscular mycorrhiza and nitrogen fixing symbiosis in sustainable agriculture. In: Werner D, Newton WE, eds. Agriculture, forestry, ecology and the environment. The Netherlands: Kluwer Academic Publishers.
- Bartlett, G.R. 1954. Phosphorus assay in column chromatography. J. Biol. Chem. 234:466-468.
- Bates LS, Waldran RP and Treare ID 1973. Rapid determination of free proline for water stress studies. Plant soil 39:205-208
- Boer, W., Folman, L.B., Summerbell, R.C. and Boddy, L. 2005. Living in a fungal world: Impact of fungi on soil bacteria niche development. FEMS Micro. Biol. Rev., 29:795-811.
- Bohnert, H.J., Nelson, D.E. and Jensen, R.G. 1995. Adaptation to environmental stress. Plant Cell., 7: 1099-1111.
- Chaurasia, B. 2001. Ecological study of tropical forest trees with special reference to vesicular arbuscular mycorrhizal (VAM) association. Ph.D. Thesis, Dr.H.S. Gour University, Sagar, M.P. India. 172.
- Goel, A.K., Laura, R.D., Pathak, D.V., Anuradha, G. and Goel, A. 1999. Use of biofertilizers: Potential, constraints and future strategies review. International Journal of Tropical Agriculture, 17: 1-18.
- Guriqbal, S., Sekhon, H.S., Poonam, S., Singh, G. and Sharma, P. 2001. Effect of *Rhizobium*, Vesicular arbuscular mycorrhiza and phosphorus on the growth and yield of lentil (*Lens culinaris*) and field pea (*Pisum sativum*). Environ. Ecol. 19(1): 40-42.

- Jaleel, C.A., Gopi, R., Sankar, B., Manivannan, P., Kishore Kumar, A., Sridharan, R. and Panneerselvam, R.2007. Studies on germination, seedling vigour, lipid peroxidation and proline metabolism in *Cathranthus roseus* seedlings under salt stress. *South Afri. J. Bot.*, 73: 190-195.
- Khan, I.A., Ayub, N., Mirza, S.N., Nizami, S.M. and Azam, M.2008. Synergistic effect of dualinoculation (vesicular-arbuscular mycorrhizae) on the growth and nutrients uptake of *Medicago sativa*, *Pak.J.Bot.* 40(2): 939-945.
- Kumar, J. and Abbo.S.2001. Genetics of flowering time in chickpea and its bearing on productivity in semiarid environment. *Adv. Agron.* 72: 107-138.
- Lekberg, Y. and R.T. Koids. 2005. Arbuscular mycorrhizal fungi, rhizobia available P and nodulation of groundnut (*Arachis hypogea* L.) in zimbabwe. *Agric. Ecosys. Environ.*, 110: 143-148.
- Lin, X.G., Hao, W.Y. and Wo, T.H. 1993. The beneficial effects of dual inoculation of vesicular-arbuscular mycorrhizae and *Rhizobium* on growth of white clover. *Tropiculture.* 11: 151-154.
- Little, T.M. and Hills, F.C. 1978. *Agricultural experimentation* John Wiley and Sons Inc, U.S.A.
- Lowrey, O.H., Rosenbrough, N.J., Farr, A.L. and Randall, R.J.1951. Protein measurement with the folin phenol reagent. *J.Biol.chem.*193:265-275.
- Mafakheri, A., Siosemardeh, A., Bahramnejad, B., Struik, P.C. and E.Sohrabi.2010. Effect of drought stress on yield, proline and chlorophyll contents in three chickpea cultivars. *Australian Journal of Crop Science*, 4(8): 580-585.
- Maggio, A., Miyazaki, S., Veronese, P., Fujita, T., Ibeas, J.I., Damsz, B., Narasimhan, M.L., Hasegawa, P.M., Joly, R.J. and Bressan, R.A. 2002. Does proline accumulation play an active role in stress-induced growth reduction. *Plant J.*, 31: 699-712.
- Mallik, M.A.2000. Association of arbuscular mycorrhizae with some varieties of Tobacco(*Nicotiana tobacum* L.) and its effect on their growth, nutrition and certain soil borne diseases, Ph.D. Thesis, Bharathidasan University, Tiruchirapalli, S.india, pp.104.
- Manivannan, P., Abdul Jaleel, C., Sankar, B., Kishorekumar, A., Somasundaram, R., Lakshman, G.M.A. and Panneerselvam, R. 2007. Growth, biochemical modifications and proline metabolism in *Helianthus annus* L. as induced by drought stress. *Colloids and surfaces: B: Biointerfaces*, 59:141-149.
- Manke, N.M., Potdukhe, S.R., Bramhankar, S.B. and Padghan, P.R. 2008. Effect of *Rhizobium* and VAM on growth parameters and yield of mungbean. *Journal of Plant Disease Sciences.* 3(2): Print ISSN: 0973-7456.
- Mohammadkhani, N. and Heidari, R. 2008. Drought – induced Accumulation of soluble sugars and proline in two Maize varieties. *World Applied Sciences Journal*, 3(3):448- 453.
- Morte, A., Lovisola, C. and Schubert, A.2000. Effect of drought stress on the growth and water relations of the mycorrhizal association *Helianthemum almeriense*- *Terfezia clavary*. *Mycorrhiza*.10 (3): 115-119.
- Muller, I. and Hofner, W.1991. Influence of arbuscular mycorrhiza on phosphorus uptake and recovery potential of maize(*Zea mays* L.) under water-stressed conditions, *Mycol.Res.*98:593-603.
- Muthukumar, T., Udaiyan, K. and Rajeshkannan, V.2001. Response of neem (*Azadiracta indica* A.Juss) to indigenous arbuscular mycorrhizal fungi, phosphate- solubilizing and symbiotic nitrogen- fixing bacteria under tropical nursery conditions. *Biology and Fertility of Soils*.34:417-426.
- Neumann, E. and George. E.2009. The effect of arbuscular mycorrhizal root colonization on growth and nutrient uptake of two different cowpea (*Vigna unguiculata*(L.) Walp. Genotypes exposed to drought stress. *Emin. J. Food. Agric.*, 21:1-17. SPACE-wheat project. *Eur.J.Agron.*,10: 197-203.
- Philips, J. M. and Hayman, D. S. 1970. Improved procedures for clearing and staining parasites and vesicular-arbuscular mycorrhizal fungi for the rapid assessment of infection. *Trans.Br.Mycol.Soc.* 55: 158-161.

Porres, J.M., Jurado, M.L., Aranda, P. and Urbano, G. 2003. Effect of heat treatment and mineral and vitamin supplement on the nutritive use of protein and calcium from lentils (*Lens esculinaris*, M.) in growing rats. *Nutrition*, 19(5): 451-456.

Rabie, G.H. and Almadini, A.M. 2005. Role of bioinoculants in development of salt-tolerance of *Vicia faba* plants under salinity stress. *African Journal of Biotechnology*. 4:210-222.

Ridley, S.M. 1977. Interaction of chloroplast with inhibitors. Induction of chlorosis by diuron during prolonged illumination in vitro. *Plant Physiol.* 59:724-732.

Sharma, S., Aneja, M., Mayer, J., Schloter, M. and Munch, J.C. 2005. Characterization of bacterial community structure in rhizosphere soil of grain legumes. *Microbiol. Ecol.*, 49, 407-415.

Simirnof, N. 1995. Antioxidant systems and plant response to the environment. In Simirnof, V. (Edn.). *Environment and plant metabolism: Flexibility and Acclimation*, BIOS Scientific Publishers, Oxford, U.K..

Strack, D., Fester, T., Hause, B., Schliemann, W. and Walter, M.N. 2003. Arbuscular mycorrhiza: Biological, chemical and molecular aspects. *J. Chem. Ecol.*, 29: 1955-1979.

Umbreit, W.W., Burris, R.H. and Stauffer, J.F. 1972. Method for Nitrogen. In: *Manometric and Bio-chemical Techniques* (5th ed). Burgess publishing company, Minnesota, pp.259-260.

Verbruggen, N. and Heramns, C. 2008. Proline accumulation in plants: A review, *Amino Acids*, 35: 753-759.

