



# MEDICINAL AND BIOLOGICAL ASPECTS OF *EUPHORBIA HIRTA* PLANT: A REVIEW

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**Abstract:** The common medicinal plant of *Euphorbia hirta* (Family: Euphorbiaceae) has potent molluscicidal and piscicidal activity against freshwater snails and fishes. The toxicological actions of the diterpenoids, rutin,  $\beta$ -sitosterol, botulin, and ellagic acid present in the *Euphorbia hirta* plant. *Euphorbia hirta* is an important medicinal herb, it is a widely used local and traditional medicine in clinical practice. The whole plant is commonly applied to cure various diseases especially gastrointestinal disorders including intestinal parasites, diarrhea, peptic ulcers, heartburn, vomiting, amoebic dysentery, afflictions of the skin, asthma, bronchitis, fever, coughs, and colds. The freshwater harmful snails *Lymnaea acuminata* and *Indoplanorbis exustus* are very common in the Northern part of India. Both snails are vectors of *Fasciola hepatica*, which causes endemic fascioliasis in cattle and livestock in the Northern part of India. Fishing with the aid of plant toxins was formerly very common. Today this easy method of fishing is still practiced in remote areas. The method is simple the poisonous ingredients are pounded and thrown into a pool or dammed-up sections of a small river. After a short time, the fish begin to raise the surface and can then readily be taken by hand. The present work is an extensive review of published literature concerning medicinal and biological aspects of the *Euphorbia hirta* plant to give comprehensive information in an attempt to provide direction for further research. It is commonly known as Dudhi.

**Index Terms:** Molluscicides, Piscicides, Snails, Fishes, Plant

## I. INTRODUCTION

The plant *Euphorbia hirta* is an important medicinal herb, it is widely used in local and traditional medicine in clinical practice. The whole plant is commonly applied to cure various diseases especially gastrointestinal disorders including intestinal parasites, diarrhea, peptic ulcers, heartburn, vomiting, amoebic dysentery, afflictions of the skin, asthma, bronchitis, fever, coughs, and colds (Chopra *et al.*, 1969; Nadkarni, 1976; Zhong, 1986; Zhong, 1999; Bhatnagar *et al.*, 2000; Patil *et al.*, 2009; Huang *et al.*, 2012; Tripathi *et al.*, 2021). In East, Central, and West Africa, a decoction of the herb is used to treat asthma, oral thrust, boils, sores, skin, and wound infections, in addition to its being used as an antispasmodic, antipruritic, carminative, depurative,

diuretic, febrifuge, purgative and vermifuge (Alabashi *et al.*, 1999; Palombo and Semple, 2001; Darwish *et al.*, 2002; Ogbolie *et al.*, 2007).

India has one of the richest herbal medical traditions in the world. Ayurveda is the system of traditional medicine prevalent in India since 2000 B.C. Ayurveda meaning the “science of life” is the oldest existing medical system recognized by WHO which is widely practiced. India is a land of immense biodiversity in which two (the Eastern Himalayas and the Western Ghats of India) out of twenty-five hot spots of the world are located. This country is perhaps the largest producer of medicinal herbs and is rightly called the botanical garden of the world. It is generally estimated that in India over 6000 plants are used in traditional, folk, and herbal medicine, representing about 75% of the medicinal needs of the Third World Countries (Rajshekharan, 2002 and Bigoniya and Rana, 2008). Medicinal herbs as a potential source of therapeutic aids have attained a significant role in the health system all over the world for both human’s animals not only in diseased conditions but also as potential material for maintaining proper health.

Interest in traditional drugs is not new but has been spurred in recent years due to an upsurge of interest in renewable resources in traditional medicine by methodological advances in pharmacology and phytochemistry. Medicinal plants are important for pharmacological research and drug development, not only when plant constituents are used directly as therapeutic agents, but also as starting materials for the synthesis of drugs or as models for pharmacologically active compounds (Mukherjee *et al.*, 2003). A significant number of modern pharmaceutical drugs are thus based on or derived from medicinal plants. Traditional medicine is a powerful source of biologically active compounds. Ethno-pharmacology has become a scientific backbone in the development of active therapeutics based on the traditional medicine of various ethnic groups.

The above-mentioned properties of plant products have opened a new vista. About India, our country possesses a rich biodiversity of medicinal plants that are used for many purposes. The plant products (extracts also) have been used from immemorial. The Vrikshayurveda is the branch of Ayurveda that deals with plant health and recommends drugs possessing specific qualities of treatment of insect attack. During the co-evolution of plants and insects, the plant has bio-synthesized several secondary metabolites to serve as defense chemicals against pest attacks. Although only 10,000 secondary metabolites have been chemically identified so, the total number may exceed 400,000 (Swain, 1977, Cooper and Johnson, 1984). A list of various plant products has been tested during the past decade and has been shown to possess molluscicidal and piscicidal activity. The present review deals with botanical products, which have demonstrated their efficacy in the management of disease vectors as alterations of synthetic pesticides.

The freshwater harmful snails *Lymnaea acuminata* and *Indoplanorbis exustus* are very common in the Northern part of India. Both the snails are vectors of *Fasciola hepatica*, which cause endemic fascioliasis in cattle and live-stock in the Northern part of India (Singh and Agarwal, 1981; Singh and Agarwal, 1990; Singh, 2000; Singh *et al.*, 2000; Yadav and Singh, 2001; 2002; Yadav *et al.*, 2002; Singh and Singh, 2003; 2003; 2003; Singh *et al.*, 2004; 2004; Singh and Singh, 2004; Singh and Singh, 2005; Singh *et al.*, 2005; Yadav *et al.*, 2005; 2006; Yadav and Singh, 2006; 2007; Singh *et al.*, 2009; Yadav *et al.*, 2009; Singh *et al.*, 2010). Fascioliasis, a devastating disease, is associated with significant economic loss in the livestock industry and has public health importance. This disease is caused by *Fasciola hepatica* in Europe, America, and Oceania (Torgerson and Claxton, 1999; Terasaki *et al.*, 2001; 2003). Snail control is a somewhat neglected aspect of the control of fascioliasis. However, it is an essential element of an integrated approach to preventing the infection caused by this parasite, which remains one of the most important diseases in this region. The use of synthetic or petroleum-based molluscicides for controlling vector snails causes serious environmental pollution (Redinger, 1976; Mian and Mulla, 1992; Dua *et al.*, 1998; Susan *et al.*, 1999; Waliszewski *et al.*, 1999). To overcome the problem and to search for eco-friendly molluscicides, several extracts and essential oils and their isolates have been evaluated for use as molluscicides due to their high toxicity, easy availability, low mammalian toxicity, low cost, and easy biodegradability (Kinghorn and Evans, 1975; Marston and Hostettmann, 1985; Singh *et al.*, 1996). These materials have shown encouraging results for vector-controlling properties with various snail species.

Fishing with the aid of plant toxins was formerly very common. Today this easy method of fishing is still practiced in remote areas. The method is simple the poisonous ingredients are pounded and thrown into a pool or dammed-up sections of a small river. After a short time, the fish begin to raise the surface and can then readily be taken by hand. The fish can be eaten without health problems (Singh, 2001). Several biocidal plants have been in use for fish-catching practices by the tribal communities in large numbers including Tharu, Bhotia, Kol,

Gond, Kharwar, and Korwas that inhabit remote villages and forest areas of the States Uttar Pradesh and Bihar (Prakash and Singh, 2000). Different species of plants employed as piscicides have different effects, depending on the species of fish targeted (Van Andel, 2000). The active principles in the plant part used (leaves, seeds, kernals, and bark) have varying potencies and modes of action depending on whether it is applied directly and the forms of extracts (aqueous and solvent extracts) used (Sambasivan *et al.*, 2003; Neuwinger, 2004; Singh and Singh, 2010; Singh *et al.*, 2010; Singh *et al.*, 2013).

Recently *Euphorbia hirta* is one of the medicinal plants currently being investigated in the Philippines for its potential against coronavirus (CoV) disease 2019 (COVID-19). The goal is to develop a formulation utilizing the plant as an adjuvant treatment for mild to moderate COVID-19. A recently published review article identified *E. hirta* as one of the Philippine medicinal plants with immunomodulatory effects and potential against severe acute respiratory syndrome-CoV-2 (SARS-CoV-2) (Dayrit *et al.*, 2021), the virus responsible for COVID-19. In this connection, a parallel and complementary *in silico* study was conducted to investigate the potential of its phytochemicals against a specific COVID-19 drug target, SARS-CoV-2 main protease (Mpro). Mpro is seen as an important COVID-19 drug target because of the role it plays in the regulation of viral replication (Di Micco *et al.*, 2021; Cayona and Creencia, 2022).

## II. MEDICINAL PROPERTIES

*Euphorbia hirta* is a small annual, branched herb prostrate to ascending with branches reaching 70 cm in height, reddish or purplish in color, with abundant latex, and is covered with short hairs (Figure 01). Its leaves are opposite, distichously and simple; its obvious stipules are linear. The leaf blades of *E. hirta* are lanceolate-oblong, long elliptic, or ovate-lanceolate; its base is very unequal; one side is cuneate, the other side is round; the apex is almost acute and its margins are finely toothed, often with a purple blotch near the midvein. The toxicological actions of the diterpenoids, rutin,  $\beta$ -sitosterol, botulin, and ellagic acid present in the *Euphorbia hirta* plant. *Euphorbia hirta* is an important medicinal herb, it is a widely used local and traditional medicine in clinical practice. The whole plant is commonly applied to cure various diseases especially gastrointestinal disorders including intestinal parasites, diarrhea, peptic ulcers, heartburn, vomiting, amoebic dysentery, afflictions of the skin, asthma, bronchitis, fever, coughs, and colds (Zhong, 1999; Bhatnagar *et al.*, 2000; Patil *et al.*, 2009; Huang *et al.*, 2012; Tripathi *et al.*, 2021 and Di Micco *et al.*, 2021; Cayona and Creencia, 2022).

*Euphorbia hirta* mainly contains flavonoids, terpenoids, rutin,  $\beta$ -sitosterol, botulin, ellagic acid phenols, essential oil, and other compounds. The major chemical structures of these compounds are shown in Figure 1. One type of the important constituents of *Euphorbia hirta* is flavonoids including quercetin, quercitrin, quercetin, and derivatives containing rhamnose, quercetin-rhamnose, a chlorophenol acid, rutin, leucocyanidin, leucocyanidol, myricitrin, cyanidin 3,5-diglucoside, pelargonium 3,5-glucoside, and, camphol. The flavonol glycoside xanthorhamnin was also isolated from *Euphorbia hirta*. The stems contain hydrocarbon hentriacontane and myricyl alcohol. The latex contains inositol, taraxerol, friedelin,  $\beta$ -sitosterol, ellagic acid, kaempferol, quercitol and quercitrin. Another type of constituent of the aerial parts of *E. hirta* is terpenoids, including triterpenes:  $\alpha$ -amyrin,  $\beta$ -amyrin, friedelin, taraxerol, and its ester: Taraxerone, 11 $\alpha$ , 12 $\alpha$ -oxidotaraxerol, cycloartenol, 24-methylene cycloartenol, and euphorbol hexacosate. The aerial parts and roots of *Euphorbia hirta* also contain diterpene esters of the phorbol type and ingenious type, including 12-deoxyphorbol-13-dodecanoate-20-acetate, 12-deoxy horbol-13-phenylacetate-20-acetate, ingenol triacetate, as well as the highly toxic tiny toxin, a resiniferous derivative. Some new ent-kaurane diterpenoids were isolated from the ethanol extract of *Euphorbia hirta* and identified as 2-beta, 16-alpha, 19-trihydroxy-ent-kaurane, 2-beta, 16-alpha-dihydroxy-ent-kaurane, and 16-alpha, 19-dihydroxy ent-karate (Yan *et al.*, 2011). The other terpenoids isolated are sterols, including  $\beta$ -sitosterol, campesterol, cholesterol, and stigmasterol (Hazimi *et al.*, 2008; Baslas and Agarwal, 1980).



**Figure 01:** *Euphorbia hirta* plant

Tannins isolated from *Euphorbia hirta* include the dimeric hydrolyzable de-hydro ellagitannins euphorbias A, B, C, E, and terchebin, the monomeric hydrolyzable tannins geraniin, 2,4,6-tri-O-galloyl- $\beta$ -D-glucose and 1,2,3,4,6-penta-O-galloyl- $\beta$ -D-glucose and the esters 5-Ocaffeoylquinic acid (neochlorogenic acid), and 3,4-di-Ogalloylquinic acid, and benzyl gallate. Acids isolated from *Euphorbia hirta* include ellagic, gallic, tannic, malic, and tartaric acids.

The major components of the essential oil include 3,7,11,15-tetramethyl-2hexadecen-1-ol, 6,10,14-trimethyl-2-pentadecanone, hexadecanal, phytol, and n-hexadecanoic acid adding up to 61.01%. The minor constituents of *Euphorbia hirta* include 2-butoxyethanol, tetradecane, phthalic acid, butyl tetradecyl ester, oleic acid, 13-heptadecyn-1-ol, 2-methyl-1-hexadecanol, and 1,2-benzene dicarboxylic acid diisooctylester. The component in essential oil may be responsible for the treatment of asthma and function as a repellent against Anopheles species, and thus, is useful for the treatment of malaria (Ogunlesi et al., 2009). The other compounds found in *Euphorbia hirta* are alkaloids, saponins, amino acids, and minerals. The mineral content of a sample of the dried leaves was: Ca 1.1%, P 0.3%, Fe 0.03%, Mg 0.5%, Mn 0.01%, Zn 0.01%, and Cu 0.002%. Fresh leaves from *Euphorbia hirta* plants of Nigerian origin were found to contain high levels of Mn (189 ppm), Cu (30.5 ppm), Zn (152 ppm), and  $\text{NO}_3$  (4600 ppm). Varying proportions of Fe, Mg, K, Ca, and Na were also found. More recently, two novel butanol rhamnopyranosides (1 and 2), have been isolated from various non-polar and polar extracts of an Indian traditional herb, *Euphorbia hirta*. The structures of the new compounds were elucidated as n-butyl-1-O-L-rhamnopyranoside (1) and n-butyl-1-O-L-rhamnopyranoside (Mallavadhani and Narasimhan, 2009).

### III. BIOLOGICAL PROPERTIES

#### *Molluscicidal properties*

Earlier observations indicate that the aqueous extracts of the leaf and bark of *Euphorbia hirta* plant (Family: Euphorbiaceae) have potent molluscicidal activity against freshwater vector snails *Lymnaea acuminata* and *Indoplanorbis exustus* (Singh et al., 2003; Singh et al., 2004; Singh et al., 2005; Singh et al., 2010; Singh and Singh, 2010). Both the snails are the intermediate host of liver-flukes *Fasciola hepatica* and *Fasciola gigantica*, which cause endemic fascioliasis in the cattle and livestock of Northern part of India (Hyman, 1970; Singh and Agarwal, 1981). The use of plant products as molluscicide is a simple, easily biodegradable, inexpensive, and appropriate technology for focal snail vectors (Kinghorn and Evans, 1975; Kloos and McCullough, 1987; Marston and Hostettmann, 1985; Singh et al., 1996; Yadav and Singh, 2001; Singh et al., 2004; Singh et al., 2005; Azare et al., 2007; Plan et al., 2008; El-Sherbini et al., 2009; El-Kamali et al., 2010). Furthermore, investigation of plants used in traditional medicine or recorded in ethnopharmacological literature provides a ready means of increasing the diversity of available molluscicides and simplifying the choice of selective, ecologically safe snail-killing compounds (Fransworth et al., 1987). The plant-origin pesticides to control the vector snails for the best methods in comparison to synthetic pesticides (Table 1).

**Table 1.** Comparison of LC<sub>50</sub> to plant origin pesticides and synthetic pesticides against snails.

Plants	Plant parts	Active moiety	LC <sub>50</sub> mg/L	Snail species	References
<b>Plant origin pesticides</b>					
<i>Euphorbia hirta</i>	Latex	Unknown	1.29 (24h)	<i>L. acuminata</i>	Singh <i>et al.</i> , 2004a
<i>Euphorbia hirta</i>	Latex (Diethyl ether)	Unknown	0.69 (24h)	<i>L. acuminata</i>	Singh <i>et al.</i> , 2004c
<b>Synthetic pesticides</b>					
Mexacarbate	-	-	3.5 (24h)	<i>L. acuminata</i>	Singh and Agarwal, 1981
Aldicarb	-	-	30 (24h)	<i>L. acuminata</i>	Singh and Agarwal, 1981
Carbaryl	-	-	14.4 (96h)	<i>L. acuminata</i>	Singh and Agarwal, 1981
Formothion	-	-	27 (24h)	<i>L. acuminata</i>	Singh and Agarwal, 1981
Phorate	-	-	15 (96h)	<i>L. acuminata</i>	Singh and Agarwal, 1981
Trichlorofon	-	-	14.4 (96h)	<i>L. acuminata</i>	Singh and Agarwal, 1981
Niclosamide	-	-	11.8 (24h)	<i>L. acuminata</i>	Singh and Agarwal, 1986
Cypermethrin	-	-	0.80 (24h)	<i>L. acuminata</i>	Singh and Agarwal, 1986
Permethrin	-	-	0.73 (24h)	<i>L. acuminata</i>	Singh and Agarwal, 1986
Fenvalerate	-	-	2.5 (24h)	<i>L. acuminata</i>	Sahay <i>et al.</i> , 1991

### ***Piscicidal properties***

Several biocidal plants have been in use for fish-catching practices by the tribal communities in large numbers including Tharu, Bhotia, Kol, Gond, Kharwar, and Korwas that inhabit remote villages and forest areas of the States Uttar Pradesh and Bihar (Prakash and Singh, 2000). Different species of plants employed as piscicides have different effects, depending on the species of fish targeted (Van Andel, 2000). The active principles in the plant part used (leaves, seeds, kernels, and bark) have varying potencies and modes of action depending on whether it is applied directly and the forms of extracts (aqueous and solvent extracts) used (Sambasivan *et al.*, 2003; Neuwinger, 2004; Singh and Singh, 2010; Singh *et al.*, 2010; Singh *et al.*, 2013).

Plant extracts are referred to as botanicals and when poisonous to fish are called piscicides (Burkill, 1985). Such piscicidal plants contain different active ingredients known as resin, tannins, saponins, nicotine, and diosgenin. However, these alkaloids are toxic to fish at high concentrations and wear off within a short time. Several plant materials are toxic to zooplankton (Kreutzweiser *et al.*, 2004), shrimps commercial fish species both in the laboratory and field studies (Singh and Singh, 2002). The Indian major carp *Labeo rohita* (Hamilton) was used as the test animal because it is present in almost all freshwater reservoirs in India and is suitable for toxicity monitoring (Table 2). Recently the freshwater fish *Channa punctatus* was used as the test animal in almost all freshwater reservoirs in India and is suitable for toxicity monitoring (Singh *et al.*, 2013).

**Table 2.** Comparison of LC<sub>50</sub> to plant-origin pesticides and synthetic pesticides against fishes.

Plants	Plant parts	Active moiety	LC <sub>50</sub> mg/L	Fish species	References
<b>Plant origin pesticides</b>					
<i>Euphorbia hirta</i>	Latex	Aqueous extract	4.23 (24h)	<i>Channa punctatus</i>	Yadav and Singh, 2013
<i>Euphorbia hirta</i>	Stem Bark	Aqueous extract	3.82 (24h)	<i>Channa punctatus</i>	Yadav and Singh, 2013
<i>Euphorbia hirta</i>	Latex	Aqueous extract	6.37 (24h)	<i>Labeo rohita</i>	Singh <i>et al.</i> , 2014
<b>Synthetic pesticides</b>					
Methyl parathion		-	8.48 ppm (24h)	<i>Poecilia reticulata</i>	Sharbidre <i>et al.</i> , 2011
Dimecron 100 SCW		-	22.95 ppm (96h)	<i>Channa panctatus</i>	Hossain <i>et al.</i> , 2002
Dimecron 100 SCW		-	375.26 ppm (96h)	<i>Barbodes gonionotus</i>	Hossain <i>et al.</i> , 2002

#### IV. CONCLUSION

In the present review, we have congregated information about the medicinal and biological aspects of molluscicidal and piscicidal claims and scientific studies. The above plant is immense potential and has a broad spectrum of activity for several ailments. *Euphorbia hirta* is an important medicinal herb, it is a widely used local and traditional medicine in clinical practice. The whole plant is commonly applied to cure various diseases especially gastrointestinal disorders including intestinal parasites, diarrhea, peptic ulcers, heartburn, vomiting, amoebic dysentery, afflictions of the skin, asthma, bronchitis, fever, coughs, and colds (Zhong, 1999; Bhatnagar *et al.*, 2000; Patil *et al.*, 2009; Huang *et al.*, 2012; Tripathi *et al.*, 2021). There are a very large number of plants, which contain compounds lethal to target as well as non-target organisms at doses, which are much below those for synthetic pyrethroids. The use of such products has the additional advantage that these contain biodegradable compounds, which are less likely to cause environmental contamination. After all such compounds are not only confined to the plants in which they are found but also possibly get distributed in the environment. We strongly feel that if these herbaceous products are used as molluscicides they would not only control the vector snail, predatory fish, and mosquitoes but would also have the advantage of easy availability, low-cost biodegradability, and greater acceptance amongst the users. Further, we feel that with further progress in biotechnology, such products could be raised from, sources other than those plants in which they are currently found. Production of plant pesticides could, in the long run also become an important industry using biotechnological methods.

#### REFERENCES

1. Alabashi, R.H., Safiyova, S. & Crakor, I.E. 1999. Some antimicrobial activity of some Yemeni medicinal plants. *Journal of Herbs Spices and Medicinal Plants*, 6: 75-83.
2. Baslas, R.K. & Agarwal, R. 1980. Isolation and characterization of different constituents of *Euphorbia hirta* Linn. *Current Science*, 49: 311-312.
3. Bhatnagar, V.P., Kumar, A. & Srivastava, J.N. 2000. Wild medicinal herbs of Agra. *Journal of Medicinal and Aromatic Plant Science*, 22-23: 464-467.
4. Bigoniya, P. & Rana, A.C. 2008. A comprehensive phyto-pharmacological review of *Euphorbia neriifolia* Linn. *Pharmacognosy Review of Supplementary*, 2(4): 57-66.
5. Burkill, H.N. 1985. *The useful plants of West Africa (tropical)* Ed. 2 Vol. I. Families A.D. Royal Botanical Garden, Kewi.
6. Chopra, R.N, Chopra, I.C. & Verma, V.S. 1969. *Supplement to glossary of Indian medicinal plants*. New Delhi, Council of Scientific and Industrial Research.
7. Cayona, R. & Creencia, E. 2022. Phytochemicals of *Euphorbia hirta* L. and Their Inhibitory Potential Against SARS-CoV-2 Main Protease. *Frontiers in Molecular Biosciences*, 8: 801-811.

8. Cooper, M.R. & Johnson, A.W. 1984. In reference book 161. Ministry of Agriculture, Fisheries and Food, London.
9. Darwish, R.M. Aurjai, I. Al-Khalil, N. & Mahafzah, A. 2002. Screening of antibiotic resistant inhibitor from local plant material against two strains of *Staphylococcus aureus*. *Journal of Ethnopharmacology*, 76: 359-364.
10. Dayrit, F.M. Jr A.M.G. & Gloriani, N.G. 2021. Philippine Medicinal Plants with Potential Immunomodulatory and Anti-SARS-CoV-2 Activities. *Philippine Journal of Science*, 15(5): 17.
11. Di Micco, S. Musella, S. Scala, M.C. Sala, M. Campiglia, P. & Bifulco, G. 2021. In Silico Analysis Revealed Potential Anti-SARS-CoV-2 Main Protease Activity by the Zonulin Inhibitor Larazotide Acetate. *Frontier Chemistry*, 8: 609-628.
12. Dua, V.K. Pant C.S. Sharma V.P. & Pathak G.K. Determination of HCH and DDT in finger-prick whole blood dried on filter papers and its field application for monitoring concentrations in blood. *Bulletin of Environmental Contamination and Toxicology*, 1998; 56: 50-57.
13. Hazimi, A. Mohammad, H. & Sarra, A. 2008. Jolkinolide diterpenoids and other constituents from *Euphorbia hirta*. *Journal of Saudi Chemical Society*, 12: 87-93.
14. Hossain, Z. Rahman, M.Z. & Mollah, M.F.A. (2002). Effect of Dimecron 100 SCW on *Anabas testudineus*, *Channa punctatus* and *Barbodes gonionotus*. *Indian Journal of Fisheries*, 49(4): 405-417.
15. Huang, L. Chen, S. & Yang, M. 2012. *Euphorbia hirta* (Feiyangcao): A review on its ethnopharmacology, phytochemistry and pharmacology. *Journal of Medicinal Plants Research*, 6(39): 5176-5185.
16. Kinghorn, A.D. & Evans F.J. 1975. A Biological screen of selected species of the genus *Euphorbia* for skin irritant effects. *Plant Medicine*, 28: 325-335.
17. Kreuzweiser, D.P. Back R.C. Sutton T.M. Pangle K.L. & Thompson D.G. 2004. Aquatic mesocosm assessments of a neem (azadiractin) insecticides at environmentally realistic concentrations-2: zooplankton community responses and recovery. *Ecology of Environmental Safety*, 59: 1 94-204.
18. Mallavadhani, U.V. & Narasimhan, K. 2009. Two novel butanol rhamnosides from an Indian traditional herb *Euphorbia hirta*. *Natural Products Research*, 23(7): 644-651.
19. Marston, A. Hostettmann K. 1985. Plant molluscicides. *Phytochemistry*, 24: 639-652.
20. Mian, L.S. Mulla M.S. 1992. Effects of pyrethroid insecticides on non-target invertebrates in aquatic ecosystem. *Journal of Agricultural Entomology*, 9(2): 73-98.
21. Mohaptra, B.C. Sovan Sahu. 2000. Toxicity of Karanj, *Pongamia pinnata* seed on different fishes. The 5<sup>th</sup> Indian Fisheries Forum. 17-20 January 2000. CIFA Bhubneshwar.
22. Mukherjee, P.K. 2003. GMP for Indian System of Medicine. In: P.K. Mukherjee, R. Verpoorte, eds. GMP for Botanicals: regulatory and Quality Issues on Phytomedicines. New Delhi, PA: Business Horizons, 99-112.
23. Nadkarni, KM. 1976. The Indian Materia medica. 3<sup>rd</sup> ed. Bombay: Popular Prakashan.
24. Neuwinger, H.D. 2004. Review of plants used for poison fishing in tropical African Toxicon, 44(4): 417-430.
25. Ogbolie, J.N. Ogeke, C.C. Okoli, I.C. & Anyanwu, B.N. 2007. Antibacterial activities and toxicology potentials of crude ethanolic extracts of *Euphorbia hirta*. *African Journal of Biotechnology*, 6(13): 1544-1548.
26. Ogunlesi, M. Okiei, W. & Ofor, E. 2009. Analysis of the essential oil from the dried leaves of *Euphorbia hirta* Linn (Euphorbiceae), a potential medication for asthma. *African Journal of Biotechnology*, 8(24): 7042-7050.
27. Palombo, A. & Semple, S.J. 2001. Antibacterial activity of traditional medicinal plants. *Journal of Ethnopharmacology*, 77: 151-157.
28. Patil, S.B. Naikwade, N.S. & Magdum, C.S. 2009. Review on phytochemistry and pharmacological aspects of *Euphorbia hirta* Linn. *Journal of Pharmacological Research and Health Care*, 1(1): 113-133.
29. Prakash, A. & Singh K.K. 2000. Observation on some high valued ethno medicinal plants among the tribal of Uttar Pradesh. *Journal of Medical and Agricultural Plant Science*, (in press).
30. Rajshekharan, P.E. 2002. Herbal medicine. *World of Science, Employment News*, 21-27 November, pp. 3.
31. Redinger, R.F. 1976. Organochlorine residues in adults of six South-Western bat species. *Journal of Wild Life Management*, 40: 677-680.
32. Sambasivan, S. Karpagam, G. Chandran & Khan S.A. 2003. Toxicity of leaf extracts of yellow oleander, *Thevetia nerifolia*, on *Tilapia*. *Environmental Science Pollution Management*, 24: 201-204.

33. Sharbidre, A.A. Metkari, V. & Patode P. 2011. Effect of methyl parathion and chlorpyrifos on certain biomarkers in various tissues of guppy fish, *Poecilia reticulata*. Pesticide Biochemistry and Physiology, 101: 132–141.
34. Singh, O. & Agarwal R.A. 1981. Toxicity of certain pesticides to two economic species of snails in Northern India. Journal of Ecology and Entomology, 74: 568-571.
35. Singh, A. & Agarwal R.A. 1990. Molluscicidal and anti-cholinesterase activity of euphorbiales. Biology Agriculture and Horticulture, 7: 81-91.
36. Singh, A. Singh, D.K. Mishra, T.N. & Agarwal, R.A. 1996. Molluscicides of plant origin. Biology Agriculture and Horticulture, 13: 205-252.
37. Singh, A & Singh, S.K. 2005. Molluscicidal evaluation of three common plant species of India. Fitoterapia, 76: 747-751.
38. Singh, D. 2001. Studies on toxicological and biochemical effects of phytopesticides on non-target freshwater fish *Channa punctatus*. Ph.D. thesis D.D.U. Gorakhpur University, Gorakhpur (U.P.), India.
39. Singh, D. & Singh, A. 2002. Piscicidal effect of some common plants of India commonly used in freshwater bodies against target animals. Chemosphere, 49: 45-49.
40. Singh, S.K. 2000. Studies on molluscicidal properties of some local plants of eastern Uttar Pradesh against harmful snails. Ph.D. thesis, D.D.U. Gorakhpur University, Gorakhpur (U.P.), India.
41. Singh, S.K. Yadav, R.P. & Singh, A. 2000. Molluscicidal activity of *Thevetia peruviana* a common medicinal plant of India. Journal of Medicinal and Aromatic Plant Science, 22/4A & 23/1A: 113-116.
42. Singh, S.K. & Singh, A. 2003. Toxic effect of *Thevetia peruviana* and *Alstonia scholaris* lattices on the freshwater snail *Lymnaea acuminata*. Iberus, 21(2): 19-27.
43. Singh, S.K. & Singh, A. 2003. Molluscicidal and anti-cholinesterase activity of different solvent extracts of stem bark and leaf of *Euphorbia pulcherima* against freshwater harmful snail. First National Interaction Meet on Medicinal and Aromatic Plant, 337-344.
44. Singh, S.K. & Singh, A. 2003. Effect of the plants *Thevetia peruviana* and *Alstonia scholaris* (Family: Apocynaceae) on Acetylcholinesterase activity of *Lymnaea acuminata* snails. Egyptian Journal of Schistosomiasis and Infectious and Endemic Diseases, 25: 31-40.
45. Singh, S.K. Yadav, R.P. Singh, D. & Singh, A. 2004a. Toxic effect of two common Euphorbiales lattices on the freshwater snail *Lymnaea acuminata*. Environmental Toxicology and Pharmacology, 15: 87-93.
46. Singh, S.K. Yadav, R.P. Singh, D. & Singh, A. 2004b. Toxicological effects of stem bark of *Thevetia peruviana* and *Alstonia scholaris* on the freshwater snail *Lymnaea acuminata*. Malaysian Applied Biology, 33(1): 61-68.
47. Singh, S.K. Yadav, R.P. & Singh, A. 2004c. Molluscicidal Activity of Different Organic Solvent Latex Extracts of Some Common euphorbiales against Freshwater Harmful Snails. Journal of Sciences, Islamic Republic of Iran, 15(1): 59-63.
48. Singh, S.K. & Yadav, R.P. Tiwari S and Singh A. 2005. Toxic effect of stem bark and leaf of *Euphorbia hirta* plant against freshwater vector snail *Lymnaea acuminata*. Chemosphere, 59(2): 263-270.
49. Singh, S.K. Yadav, R.P. & Singh, A. 2009. The toxicity of *Thevetia peruviana* and *Alstonia scholaris* plants to target and non-target aquatic organism. Proceedings of the symposium on Functional Biodiversity and Ecophysiology of Animals, 301-307.
50. Singh, S.K. & Singh, A. 2009. Toxic effect of *Euphorbia pulcherima* plant to fingerlings of *Labeo rohita* (Hamilton) in different culturing conditions. World Journal of Fish and Marine Science, 1(4): 324-329.
51. Singh, S.K. & Singh, A. 2010. Molluscicidal activity of different solvent leaf and bark extracts of *Euphorbia hirta* plant against the freshwater vector snails. International Journal of Malacology Argonauta, 21(1-6): 22-33.
52. Singh, S.K. & Singh A. 2010. The toxicity of leaf and stem bark of *Thevetia peruviana* plant to fingerlings of *Labeo rohita* (Ham.) in different conditions. Malaysian Applied Biology, 39(1): 25-31.
53. Singh, S.K. Yadav, R.P. & Singh, A. 2010. Piscicidal activity of leaf and bark extract of *Thevetia peruviana* plant and their biochemical stress response on fish metabolism. European Review for Medical and Pharmacological Science, 14(11): 915-923.
54. Singh, S.K. Yadav, R.P. & Singh, A. 2010. Molluscicides from some common medicinal plants of eastern Uttar Pradesh, India Journal of Applied Toxicology, 30: 1-7.

55. Singh, S.K. Singh, S.K. & Singh, A. 2013. Toxicological and biochemical alterations of apigenin extracted from seed of *Thevetia peruviana*, a medicinal plant. *Journal of Biology and Earth Science*, 3(1): B110-B119.
56. Singh, S.K. Johnson, A. & Singh, A. 2014. Toxic effect of *Euphorbia hirta* plant to fingerlings of *Labeo rohita* (hamilton) in different culturing conditions. *Scientific Journal of Veterinary Advances*, 3(7): 83-90.
57. Susan, T. Anita Veeraiah K. Tilak, K.S. 1999. A study on the bioaccumulation of fenvalerate a synthetic pyrethroid, in the whole-body tissue of *Labeo rohita*, *Catla catla*, *Cirrhinus mrigala* (Ham.) by gas-liquid chromatography. *Pollution Research*, 18(1): 57-59.
58. Swain, T. 1977. Secondary compounds as protective against: *A Annual Review of Plant Physiology*, 28: 479-501.
59. Terasaki, K. Itagaki, T. Shibahara, T. Noda, Y. & Moriyama Gonda, N. 2001. Comparative study of the reproductive organs of *Fasciola* groups by optical microscope. *Journal of Veterinary Medical Science*, 63: 735-742.
60. Terasaki, K. Shibahara, T. Itagaki, T. Fukuda, K. Noda, Y. Mine, M. Motomura, A. & Yamamoto, M. 2003. Gametogenesis and fertilization of *Fasciola hepatica* from Uruguay. *Journal of Veterinary Medicine*, 56: 97-103.
61. Torgerson, P. & Claxton, J. 1999. Epidemiology and control. In: Dalton JP (ed) *Fascioliasis*. CABI Publishing, New York, pp. 113-149.
62. Tripathi, A.N. Sati, S.C. & Kumar, P. 2021. *Euphorbia hirta* Linn - An invasive plant: A Review of its traditional uses, phytochemistry and pharmacological properties. *International Journal of Pharmaceutical Sciences and Research*, 12(12): 6189-6201.
63. Van Andel, T. 2000. The diverse uses of fish-poison plants in Northwest Guyana. *Ecological Botany*, 54: 500-512.
64. Waliszewski, S.M. Aguirre, A.A. Benitez, A. Infanzon, R.M. Infazon, R. & Rivera, J. 1999. Organopesticides residues in Human blood serum of inhabitants of Veracruz, Mexico. *Bulletin of Environmental Contamination and Toxicology*, 62: 397-402.
65. Yadav, R.P. & Singh, A. 2001. Environmentally safe molluscicides from two common euphorbiales. *Iberus*. 19(2): 65-73.
66. Yadav, R.P. & Singh, A. 2002. Toxic effect of latex of *Croton tiglium* on *Lymnaea acuminata* and *Channa punctatus*. *Iberus*, 20(2): 33-45.
67. Yadav, R.P. Singh, S.K. & Singh, A. 2002. Molluscicidal activity of *Codiaeum variegatum*, effect on snail metabolism. *Journal of Ecophysiology and Occupational Health*, 2: 74-84.
68. Yadav, R.P. Tiwari, S. & Singh, A. 2005. Toxic effects of taraxerol extracted from *Codiaeum variegatum* stem bark on target vector snail *Lymnaea acuminata* and non-target fish. *Iberus*, 23(1): 1-13.
69. Yadav, R.P. Singh, D. Singh, S.K. & Singh, A. 2006. Toxic effect of stem bark of *Croton tiglium* on metabolism of freshwater snail *Lymnaea acuminata*. *American Malacology Bulletin*, 21: 87-92.
70. Yadav, R.P. & Singh, A. 2006. Toxic effects of *Jatropha gossypifolia* and its binary and tertiary combinations with other plant molluscicides in natural ponds. *Iberus*. 24(2): 47-54.
71. Yadav, R.P. & Singh, A. 2007. Toxic effects of Euphorbials on freshwater snail *Lymnaea acuminata* in ponds. *Journal of Herbs Spices and Medicinal Plant*, 13(2): 87-94.
72. Yadav, R.P. Singh, S.K. & Singh, A. 2009. Toxic effects of euphorbious plants against freshwater vector snails in ponds. *Proceedings of the symposium on Functional Biodiversity and Ecophysiology of Animals*, 293-300.
73. Yadav, R.P. & Singh, A. 2013. Toxic Effects of Two Common Euphorbiales against Freshwater Fish *Channa punctatus*. *International Journal of Traditional and Natural Medicines*, 3(2): 49-56
74. Yan, S.J. Ye, D.W. & Wang, Y. 2011. Ent-Kaurane Diterpenoids from *Euphorbia hirta*. *Recent Natural Products*, 5(4): 247-251.
75. Zhong, Y.D.D. 1986. Shanghai Science Technology Press. P. 139.
76. Zhong, H.B.C. 1999. Shanghai Science Technology Press. 4: 788.