



INTERNATIONAL JOURNAL OF CREATIVE RESEARCH THOUGHTS (IJCRT)

An International Open Access, Peer-reviewed, Refereed Journal

Niosomes In Skin Cancer

Yogesh Shendage, shivani rathod

Pharmacy practice, Shivlingeshwar College of pharmacy Almala, dist latur, India

Abstract : The prevalence of skin diseases, particularly skin cancer, presents a global health challenge an increasing impact on the economy and workforce. Melanoma, arising from dysfunctional melanocytes, is the most aggressive form of skin cancer, in contrast non-melanoma skin cancers (NMSC) like squamous cell carcinoma (SCC) and basal cell carcinoma (BCC) account for the majority of cases. The incidence of NMSC has risen significantly, with men being at higher risk due to genetic, phenotypic, and environmental factors. Advances in polymeric micro/nano carrier-based therapies offer promising avenues for skin cancer management, including drug and gene delivery, and combination therapies. These innovative approaches hold the potential for overcoming current treatment limitations and advancing both research and clinical applications in the field. Niosomes can be important nanomolecules For the cancer management and drug delivery. In this review, we are discussing the niosomes as a nanocarrier for drug delivery in skin cancer.

Keywords : niosomes, skin cancer, nanocarrier, penetration enhancers, cancer treatment

Introduction :

The goal of targeted drug delivery is to try to concentrate the drug in the tissues that are of interest while decreasing the drug's concentration in the rest of the tissues. Accordingly, the drug is confined to the designated site. As a result, the drug does not effect on the tissues around it. Additionally, due to drug localization, drug loss is prevented, ensuring maximum medication efficacy. Various transporters have been utilized for focusing of medication, like immunoglobulin, serum proteins, engineered polymers, liposomes, microspheres, erythrocytes and niosomes.[1]

Niosomes :

Niosomes are truly outstanding among these transporters. The self-gathering of non-ionic surfactants into vesicles was first detailed during the 70s by analysts in the restorative business. Niosomes (non-ionic surfactant vesicles) got on hydration are tiny lamellar structures framed after joining non-ionic surfactant of the alkyl or dialkyl polyglycerol ether class with cholesterol.[2] The non-ionic surfactants structure a shut bilayer vesicle in fluid media in light of its amphiphilic nature involving some energy for example heat, actual fomentation to shape this construction. The hydrophilic heads remain in contact with the aqueous solvent while the hydrophobic parts of the bilayer structure are oriented away from it. By altering their composition, size, lamellarity, tapped volume, surface charge, and concentration, the vesicles' properties can be altered. Different powers act inside the vesicle, eg, van der Waals powers among surfactant particles, shocking powers rising up out of the electrostatic associations among charged gatherings of surfactant atoms, entropic awful powers of the head gatherings of surfactants, short-acting horrendous powers, and so forth. These powers are liable for keeping up with the vesicular design of niosomes. In any case, the security of niosomes is impacted by the kind of surfactant, nature of

the typified drug, capacity temperature, cleansers, utilization of film-crossing lipids, the inter-facial polymerization of surfactant monomers in situ, consideration of charged particle. Because of presence of hydrophilic, amphiphilic, and lipophilic moieties in the construction, these can oblige drug particles with an extensive variety of solubility.[3] These may go about as a stop, delivering the medication in a controlled way. The restorative exhibition of the medication particles can likewise be worked on by deferred leeway from the course, shielding the medication from the natural climate and limiting impacts to target cells.[4] Baneful made of alpha, omega-hexadecyl-bis-(1-aza-18-crown-6) (Bola-surfactant)-Length 80-cholesterol (2:3:1 molar proportion) is named as Bola-Surfactant containing niosome.[5] The surfactants utilized in niosome readiness ought to be biodegradable, biocompatible and non-immunogenic. A dry item known as proniosomes might be hydrated preceding use to yield fluid niosome scatterings. Niosomes behave in vivo like liposomes, prolonging the circulation of entrapped drug and altering its organ distribution and metabolic stability.[7] Like liposomes, the properties of niosomes depend on the composition of the bilayer as well as the method of their production. Niosomes also provide additional convenience in transportation, distribution, storage, and dosing.[6] Liposomes and niosomes have distinct characteristics, particularly since niosomes are made from cholesterol and uncharged single-chain surfactant, whereas liposomes are made from double-chain phospholipids (neutral or charged). It has been reported that the intercalation of cholesterol in the bilayers reduces the entrapment volume during formulation and, consequently, entrapment efficiency.[8] The convergence of cholesterol in liposomes is substantially more than that in niosomes. Accordingly, drug entanglement proficiency of liposomes becomes lesser than niosomes. In addition, liposomes are costly, and their fixings, like phospholipids, are synthetically unsound due to their inclination to oxidative debasement; In addition, these necessitate special handling and storage, and the purity of natural phospholipids varies.

Niosomal drug conveyance is possibly pertinent to numerous pharmacological specialists for their activity against different sicknesses. To develop a novel drug delivery system, it can also be used as a vehicle for drugs that are difficult to absorb. It improves the bioavailability by crossing the physical obstruction of the gastrointestinal lot through transcytosis of M cells of Peyer's patches in the digestive lymphatic tissues.[9] The niosomal vesicles are taken up by the reticuloendothelial framework. Such restricted drug gathering is utilized in treatment of illnesses, for example, leishmaniasis, in which parasites attack cells of the liver and spleen.[10,11] Some non-reticulo-endothelial frameworks like immunoglobulins likewise perceive lipid surface of this conveyance system.[2-8,10-12] Exemplification of different enemies of neoplastic specialists in this transporter vesicle has limited drug-prompted harmful aftereffects while keeping up with, or in certain occasions, expanding the counter cancer efficacy.[13] Doxorubicin, the anthracycline anti-toxin with wide range hostile to growth movement, shows a portion subordinate irreversible cardio-poisonous effect.[14,15] Niosomal conveyance of this medication to mice bearing S-180 growth expanded their life expectancy and diminished the pace of expansion of sarcoma. S-180 tumor-bearing mice were given methotrexate entrapped in niosomes via intravenous administration, which resulted in total tumor regression, a higher plasma level, and a slower elimination. It has great command over the delivery pace of medication, especially for treating cerebrum harmful cancer.[16] Niosomes have been utilized to concentrate on the idea of the resistant reaction incited by antigens.[17] Niosomes can be utilized as a transporter for hemoglobin.[18,19] Vesicles are porous to oxygen and hemoglobin separation bend can be changed in much the same way as non-epitomized hemoglobin. The primary drawback of the transdermal route of administration is the drug's slow skin penetration.[20] To increase the therapeutic efficacy of certain anti-inflammatory medications like flurbiprofen and piroxicam and sex hormones like estradiol and levonorgestrel, niosomes are frequently administered transdermally. This vesicular framework likewise gives better medication focus at the site of activity managed by oral, parenteral and skin courses. Supported discharge activity of niosomes can be applied to drugs with low restorative record and low water solvency. Drug conveyance through niosomes is one of the ways to deal with limited drug activity as to their size and low vulnerability through the epithelium and connective tissue, which keeps the medication restricted at the

site of the organization. Confined drug activity improves the adequacy of strength of the medication and, simultaneously, lessens its foundational poisonous impacts, eg, antimonials epitomized inside niosomes are taken up by mononuclear cells, bringing about localisation of medication, expansion in power, and thus decline in portion as well as toxicity.[13] The development of niosomal drug conveyance innovation is currently at the phase of earliest stages, yet this kind of medication conveyance framework has shown guarantee in disease chemotherapy and against leishmanial treatment.

Characterisation of niosomes

Size State of niosomal vesicles is thought to be round, and their mean breadth not entirely set in stone by utilizing laser light dispersing method.[28] Likewise, measurement of these vesicles still up in the air by utilizing electron microscopy, atomic strainer chromatography, ultracentrifugation, photon connection microscopy and optical microscopy[29,30] and freeze break electron microscopy. Freeze defrosting of niosomes builds the vesicle breadth, which may be credited to a combination of vesicles during the cycle.

Bilayer development Gathering of non-ionic surfactants to shape a bilayer vesicle is described by a X-cross development under light polarization microscopy.[31]

Number of lamellae not entirely settled by utilizing atomic attractive reverberation (NMR) spectroscopy, little point X-beam dissipating and electron microscopy.[29]

Film inflexibility Layer unbending nature can be estimated through portability of fluorescence test as an element of temperature.[31]

Entanglement proficiency In the wake of getting ready niosomal scattering, untrapped drug is isolated and the medication remained ensnared in not set in stone by complete vesicle disturbance utilizing half n-propanol or 0.1% Triton X-100 and dissecting the resultant arrangement by fitting examine technique for the drug.[32] It very well may be addressed as:

Ensnarement proficiency (EF) = (Sum captured/aggregate sum) × 100

In vitro Delivery Study

A technique for in vitro discharge rate study was accounted for with the assistance of dialysis tubing.[33] A dialysis sac was washed and absorbed refined water. The suspension of vesicles was pipetted into a tubing-filled bag and sealed. The sack containing the vesicles was then positioned in 200 ml cradle arrangement in a 250 ml measuring utensil with steady shaking at 25°C or 37°C. An appropriate assay method was used to check the drug content of the buffer at various intervals. In another strategy, isoniazid-epitomized niosomes were isolated by gel filtration on Sephadex G-50 powder saved in twofold refined water for 48 h for swelling.[34] from the get go, 1 ml of arranged niosome suspension was put on the highest point of the segment and elution was done utilizing typical saline. Niosomes embodied isoniazid elutes out first as a somewhat thick, white opalescent suspension followed by free medication. Isolated niosomes were filled in a dialysis cylinder to which a sigma dialysis sac was connected to one end. A magnetic stirrer was used to stir the dialysis tube into phosphate buffer with a pH of 7.4, and samples were taken at specific intervals and analyzed using the high-performance liquid chromatography (HPLC) method.

Types of niosomes

Niosomes can be divided into three groups based on the size of their vesicles. These are little unilamellar vesicles (SUV, size=0.025-0.05 μm), multilamellar vesicles (MLV, size=>0.05 μm), and huge unilamellar vesicles (LUV, size=>0.10 μm).

Method of preparation

The sizes and distribution of the vesicles, the number of double layers, the aqueous phase's entrapment efficiency, and the vesicle membrane's permeability all influence how niosomes are made.

Preparation of unilamellar

Sonication The drug's aqueous phase is added to the surfactant and cholesterol mixture in a scintillation vial.[11] A sonic probe is used to homogenize the mixture for three minutes at 60°C. The vesicles are little and uniform in size.

Miniature fluidisation Two fluidised streams push ahead through exactly characterized miniature channel and connect at super high speeds inside the communication chamber.[21] Here, a typical door is organized to such an extent that the energy provided to the framework stays inside the area of niosomes development. The end result is better reproducibility, smaller size, and greater uniformity.

Preparation of multilamellar

Hand shaking strategy (Slight film hydration procedure) In the hand shaking technique, surfactant and cholesterol are broken down in an unpredictable natural dissolvable, for example, diethyl ether, chloroform or methanol in a rotational evaporator, leaving a flimsy layer of strong blend saved on the mass of the flask.[11] The dried layer is hydrated with watery stage containing drug at typical temperature with delicate unsettling.

Trans-membrane pH gradient (inside acidic) drug uptake process (remote loading) Chloroform is used to dissolve surfactant and cholesterol.[22] The solvent is then evaporated at a lower pressure to form a thin film on the wall of the round-bottom flask. Vortex mixing hydrates the film with 300 mM citric acid (pH 4.0). The multilamellar vesicles are frozen and defrosted multiple times and later sonicated. Aqueous solution containing 10 mg/ml of the drug is added to this niosomal suspension and vortexed. With 1M disodium phosphate, the sample's pH is then raised to 7.0-7.2. The desired multilamellar vesicles are made by heating this mixture for ten minutes at 60°C.

Preparation of large unilamellar

Reverse phase evaporation technique (REV) This technique involves dissolving cholesterol and surfactant in a mixture of ether and chloroform.[23] Next, an aqueous phase containing a drug is added, and the two resulting phases are sonicated at 4-5°C. Following the addition of a small amount of phosphate buffered saline, the clear gel is further sonicated. The natural stage is taken out at 40°C under low tension. Niosomes are produced by heating the viscous niosome suspension for ten minutes in a water bath at 60°C with phosphate-buffered saline.

The ether injection method essentially consists of slowly injecting niosomal ingredients in ether through a 14-gauge needle at a rate of approximately 0.25 ml/min into a preheated aqueous phase maintained at 60°C.[11,24] The ether gradient that extends towards the aqueous-nonaqueous interface may be the reason for the formation of larger unilamellar vesicles due to the slow vapourization of The previous

might be liable for the development of the bilayer structure. The fact that a small amount of ether is frequently present in the vesicle suspension and is difficult to remove is one of this method's drawbacks.

Diverse Multiple Membrane Extrusion Technique A thin film is made by evaporating a mixture of surfactant, cholesterol, and diacetyl phosphate in chloroform.[20] The film is then hydrated with an aqueous drug solution, and the suspension is extruded through polycarbonate membranes that are arranged in a series for up to eight passages. Niosome size can be controlled effectively using this strategy.

Niosome arrangement utilizing polyoxyethylene alkyl ether The size and number of bilayer of vesicles comprising polyoxyethylene alkyl ether and cholesterol can be changed utilizing an option method.[25] Temperature increase above 60°C changes little unilamellar vesicles to huge multilamellar vesicles (>1 µm), while energetic shaking at room temperature shows the contrary impact, ie, change of multilamellar vesicles into unilamellar ones. Since it is known that polyethylene glycol (PEG) and water remix at higher temperatures due to the breakdown of hydrogen bonds between water and PEG moieties, it is possible that the characteristic of polyoxyethylene alkyl ether (ester) surfactant is the transformation from unilamellar to multilamellar vesicles at higher temperatures. By and large, free medication is taken out from the typified drug by gel pervasion chromatography dialysis strategy ,or centrifugation technique. Frequently, thickness contrasts among niosomes and the outer stage are more modest than that of liposomes, which makes division by centrifugation undeniably challenging. Expansion of protamine to the vesicle suspension works with detachment during centrifugation.

Emulsion technique The oil in water (o/w) emulsion is ready from a natural arrangement of surfactant, cholesterol, and a fluid arrangement of the drug.[26,27] The natural dissolvable is then vanished, leaving niosomes scattered in the watery stage.

Lipid infusion technique This strategy doesn't need a costly natural stage. The surfactant-lipid mixture is first melted before being injected into a heated, highly agitated aqueous phase containing dissolved drugs. Here, the medication can be broken into liquid lipids and the blend will be infused into a fomented, warmed fluid stage containing surfactant.

Niosome readiness utilizing Micelle Niosomes may likewise be shaped from a blended micellar arrangement by the utilization of catalysts. A blended micellar arrangement of C16 G2, dicalcium hydrogen phosphate, and polyoxyethylene cholesteryl sebacetate diester (PCSD) converts to a niosome scattering when brooded with esterases. PCSD is divided by the esterases to yield polyoxyethylene, sebacic corrosive and cholesterol. Cholesterol in mix with C16 G2 and DCP then yields C16 G2 niosomes.

Factors Affecting Physico-Chemical Properties of Niosomes

Different variables that influence the physico-synthetic properties of niosomes are examined further.

Selection of surfactants and fundamental added substances A surfactant utilized for planning of niosomes should have a hydrophilic head and a hydrophobic tail. One or two alkyl or perfluoroalkyl groups or, in some cases, a single steroidal group may make up the hydrophobic tail.[35] The ether-type surfactants with a single-chain alkyl tail are more harmful than those with a dialkyl ether chain. The ester-type surfactants are synthetically less steady than ether-type surfactants and the previous is less poisonous than the last option because of ester-connected surfactant debased by esterases to fatty substances and unsaturated fat in vivo.[36] The surfactants with alkyl tie lengths from C12 to C18 are reasonable for planning pernicious. Range series surfactants having HLB number somewhere in the

range of 4 and 8 can frame vesicles.[37] Various sorts of non-ionic surfactants with models are given in Table 1.[38]

Type of non-ionic surfactant	Examples
Fatty alcohol	Cetyl alcohol, stearyl alcohol, cetostearyl alcohol, oleyl alcohol
Ethers	Brij, Decyl glucoside, Lauryl glucoside, Octyl glucoside, Triton X-100, Nonoxynol-9
Esters	Glyceryl laurate, Polysorbates, Spans
Block copolymers	Poloxamers

The stable niosomes can be ready with the expansion of various added substances alongside surfactants and medications. Various additives can alter the morphologies of the formed niosomes and alter their permeability and stability properties by modifying membrane characteristics. In the event of polyhedral niosomes framed from C16G2, the state of these polyhedral niosomes stays unaffected just barely by solution C24 (cholesteryl poly-24-oxyethylene ether), which forestalls accumulation because of improvement of steric deterrent. C16G2's addition, on the other hand: cholesterol: The formation of spherical niosomes by solulan (49:49:2) is influenced by the composition of the membrane.[39] Expansion of cholesterol atom to niosomal situation makes the layer unbending and lessens spillage of medication from the niosome.[40]

The temperature of Hydration temperature impacts the shape and size of the niosome. For ideal conditions, it ought to be over the gel to fluid stage change temperature of the framework. Temperature change in the niosomal framework influences the gathering of surfactants into vesicles and furthermore prompts vesicle shape transformation.[35,39] A polyhedral vesicle shaped by C16G2: When solution C24 (91:9) is heated to 25°C, it becomes a spherical vesicle at 48°C. However, when it cools to 49°C from 55°C, the vesicle becomes a cluster of smaller spherical niosomes before becoming polyhedral structures at 35°C. Conversely, the vesicle framed by C16G2: cholesterol: solution C24 (49:49:2) shows no shape change on warming or cooling.[27] Alongside the previously mentioned factors, the volume of hydration medium and season of the hydration of niosomes are additionally basic variables. Ill-advised determination of these variables might bring about the development of delicate niosomes or the production of medication spillage issues.

The nature of the encapsulated drug Charge and rigidity of the niosome bilayer are influenced by the physicochemical properties of the encapsulated drug. The medication cooperates with surfactant head gatherings and fosters the charge that makes shared shock between surfactant bilayers, and subsequently increments vesicle size.[29] The conglomeration of vesicles is forestalled because of the charge improvement on bilayer. Table 2 displays the effect of the drug's nature on the formation vesicle.

Table 2: The nature of the drug's effect on niosome formation

Nature of the drug	Leakage from the vesicle	Stability	Other properties
Hydrophobic drug	Decreased	Increased	Improved trans-dermal delivery
Hydrophilic drug	Increased	Decreased	–
Amphiphilic drug	Decreased	–	Increased encapsulation, altered electrophoretic mobility
Macromolecule	Decreased	Increased	–

Drug Entrapment in Niosomes Increases Vesicle Size Drug Entrapment in Niosomes Increases Vesicle Size Drug Entrapment in Niosomes Increases Vesicle Size Drug Entrapment in Niosomes Increases Vesicle Size In polyoxyethylene glycol (Stake)-covered vesicles, some medication is ensnared in the long Stake chains, hence lessening the propensity to expand the size. The hydrophilic-lipophilic equilibrium of the medication influences the level of entanglement.

The surface free energy decreases with an increase in hydrophobicity of surfactants, so the mean size of niosomes increases proportionally with the hydrophilic-lipophilic balance (HLB) of surfactants such as Span 85 (HLB 1.8) to Span 20 (HLB 8.6).[41] The temperature, the type of lipid or surfactant, and the presence of other components such as cholesterol determine whether the bilayers of the vesicles are in the gel state, alkyl chains are available in a very much arranged structure, and in the fluid express, the construction of the bilayers is more scattered. The surfactants and lipids are portrayed by the gel-fluid stage change temperature (TC). Stage change temperature (TC) of surfactants likewise influences ensnarement effectiveness, ie, Length 60 having higher TC gives better entanglement. Cholesterol content and charge Consideration of cholesterol in niosomes builds its hydrodynamic width and capture productivity. Cholesterol typically performs two functions. Cholesterol alters the chain order of gel state bilayers while simultaneously enhancing the chain order of liquid state bilayers. The gel state is transformed into a liquid-ordered phase at high cholesterol levels. The release rate of the encapsulated material decreased as the cholesterol content of the bilayers increased, and as a result, the rigidity of the resulting bilayers increased. In multilamellar vesicle structure, the presence of charge tends to increase the overall entrapped volume by increasing the interlamellar distance between successive bilayers.

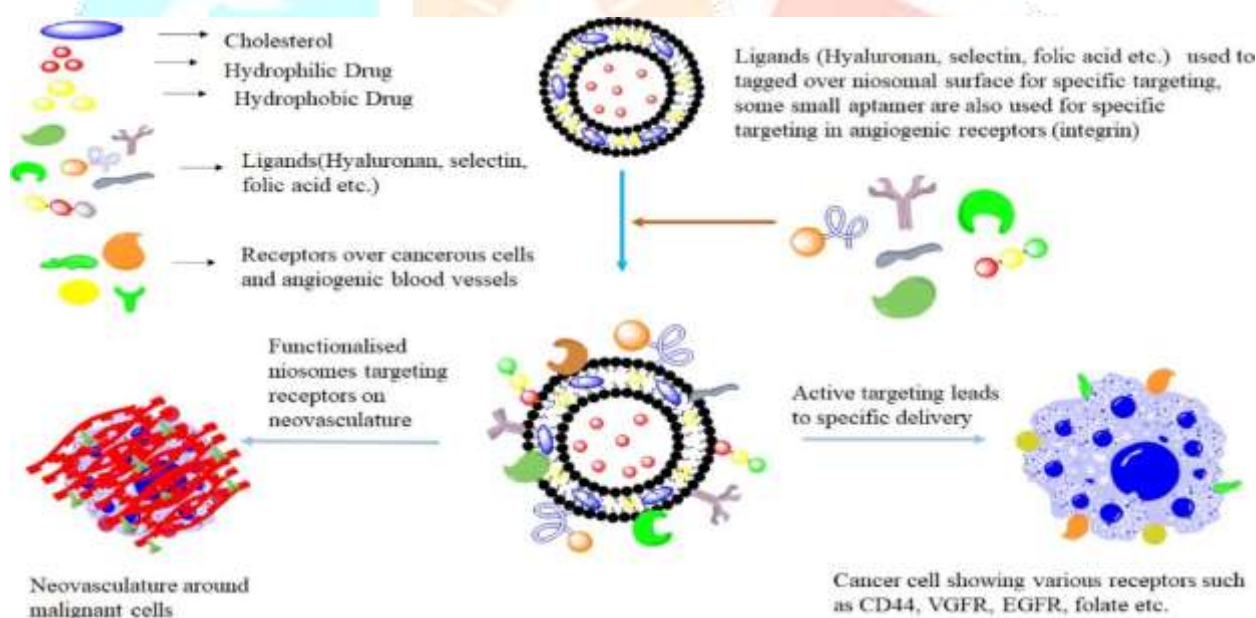
Clinical application of niosomes in cancer

Nearly 9.6 million people worldwide die of cancer, making it one of the deadliest diseases [41]. According to the contemporary examination connected with malignant growth therapy, notwithstanding the accessible definition and absence of helpful moiety, disease treatment is as yet a critical reason for worry for scientists [42]. In the number of patients, the treatment couldn't be finished because of patient rebelliousness and the different aftereffects like sickness, heaving, etc.[43] without designated drugs, these diseases restorative moiety additionally influences the typical cells rather than harmful cells and hence prompt quick and long-haul secondary effects [43]. Various receptors are primarily expressed on the surface of cancer cells, and appropriate ligand-coated nanoformulations may be used to target these receptors for cancer treatment [44]. Further, these particular designated treatments decline the dosing recurrence and lessen the poison levels and incidental effects [45],[46],[47]. In recent years, numerous studies have been conducted to successfully target cancer cell surface receptors such as CD44 [49], epidermal growth factor receptor (EGFR) [50], vascular growth factor receptor (VGFR) [51], folate, and integrin, among others, in order to achieve time-dependent targeting, minimize systemic toxicity, and

retain in the system for an extended period [48]. An abundant and adequate supply of oxygen, vascularization, and nutrients are essentially required to grow malignant cells [49]. A couple of receptors help in angiogenesis, even as an assistance in offering nourishment to growth cells. We might check the cancer cell's food supply by repressing the surface receptors like folate or transferrin, while explicit receptors like EGFR might be designated to restrain the neovasculature around the harmful cells. A few significant receptors connected with the focusing of malignant growth cells have been illustrated in ensuing segments

Nontargeted and vague treatment might cause serious entanglements in the term of secondary effects and fundamental poisonousness. In such situation, conveyance of remedial particles in a particular and designated way is especially required. Numerous ligands have been identified for various receptors. Coupling these ligands to the niosomal surface utilizing a spacer particle (EDC-NHS) direct connection of these ligands will explore the niosomal detailing toward the specific receptor and advance explicit focusing on and successful conveyance of helpful moieties. As a result, the sections that come after this one have gone over a few key receptors that are necessary for an active targeting strategy. Further, a diagrammatic portrayal of niosomes showing dynamic focusing of different receptors communicated near threatening cell surface has been portrayed the Fig.1

CD44: It is a multifunctional primary protein that aids in different cell working like cell expansion, cell bond, cell relocation, and angiogenesis, which is pivotal for danger. It has been accounted for that CD44 protein is communicated in different assortments of malignant growths like lungs, bosoms, renal, while certain creators have likewise revealed the over expression of this protein in leukemia.



Different ligands like Ezrin, Fibrin, or immobilized fibrinogen, selectin, and hyaluronan have been utilized lately to focus on the CD44 protein communicated on the outer layer of dangerous cells. Among them, hyaluronan assortments are the most common and tie explicitly with 90 kDa CD44 receptors. Penno et al., Mooney and other, furthermore, Herrlich et al. had discussed the binding of CD44 and hyaluronan in a variety of cancers, including glioblastoma, non-small cell lung cancer, and the majority of colon cancers, respectively.[52] EGFR: The epidermal growth factor receptor is a transmembrane protein receptor that is commonly referred to as ErbB-1 and HER1. These tyrosine kinase receptors are abundantly expressed in a variety of malignant cells. These receptors are also known as ErbB-1 and HER1. The EGFR receptor can impact different cell pathways (Rodent sarcoma (RAS)/Quickly sped up fibrosarcoma (RAF)/MEK/ERK, Janus kinases/signal transducer and activator of record proteins (JAK/Detail), and P13K/AKT, which at last actuates the phone motioning for multiplication, relocation, and intrusion of dangerous tissues. It has likewise been accounted for the overexpression of EGFR in non-little cell cellular breakdown in the lungs, colon, and bosom diseases might happen because of a continuous transformation by the erasure of exon 19 or point change at L858R quality. In addition, 85

percent of nasopharyngeal carcinoma patients express these receptors primarily. Additionally, monoclonal antibodies that target the EGFR protein that is expressed over the malignant cells have been clinically approved in a variety of formulations, including cetuximab and nimotuzumab.[52] VGFR: Vascular development factor receptor is a flagging protein alluded to as vascular penetrability factor, empowering vein development like vasculogenesis (development of the undeveloped circulatory framework) and angiogenesis (the development of veins from the previous vasculature). Among its five individuals, VEGF-A works with an angiogenesis and is otherwise called vascular penetrability factor (VPF). VGFR is overexpressed in bosom disease, which prompts a high metastasis rate followed by quick movement and mitosis of endothelial cells and exorbitant metalloproteinase action For focusing of VGFR, we regularly utilize monoclonal antibodies that go about as a homing device, and successive restricting will prompt restraint of angiogenesis . This technology has primarily been utilized in recent years to inhibit angiogenesis and reduce solid tumors. Tivozanib, Bevacizumab (Avastin®), Cabozantinib (Cometriq®), Everolimus (Afinitor®), and others are VEGF receptor inhibitors. [52] Folate receptor: A normally happening receptor assists cell digestion with enjoying nucleotide combination for quickly isolating cells. Additionally, these receptors are overexpressed in numerous growth cells like ovaries, neck, bosom, colon, constant myeloid leukemia, and so on. These receptor focusing can be accomplished by forming favored definitions with folate ligands and collecting in the cytosol through endocytosis. Enormous quantities of definitions are now evolved, such as vintafolide (designated folic corrosive formed framework with microtubule undermining specialist as an anticancer moiety) to target threatening cells to diminish their size and to repress metastasis .[52]Number of niosomal details are arranged utilizing different anticancer medications to improve their viability and diminish poisonousness. Different creators have created different ligand-beautified niosomes to work on their adequacy and decrease their dosing recurrence to limit their side effects[52]. A rundown of some anticancer medication stacked niosomal definitions, their trademark, and delivery profile has been summed up in Table

Routes of drug administration	Examples of Drugs
Intravenous route	Doxorubicin, methotrexate, sodium stibogluconate, iopromide, vincristine, diclofenac sodium, flurbiprofen, centchroman, indomethacin, colchicine, rifampicin, tretinoin, transferrin and glucose ligands, zidovudine, insulin, cisplatin, amarogentin, daunorubicin, amphotericin B, 5-fluorouracil, camptothecin, adriamycin, cytarabine hydrochloride
Peroral route	DNA vaccines, proteins, peptides, ergot alkaloids, ciprofloxacin, norfloxacin, insulin
Transdermal route	Flurbiprofen, piroxicam, estradiol, levonorgestrol, nimesulide, dithranol, ketoconazole, enoxacin, ketorolac
Ocular route	Timolol maleate, cyclopentolate
Nasal route	Sumatriptan, influenza viral vaccine
Inhalation	All-trans retinoic acids

Mechanisms of Niosomes Penetration through the Skin

For dermatological issues, niosome are testing drug transporters. Niosome have been utilized not only in the cosmetics industry but also for the delivery of peptide drugs. The home time frame of drug in the layer corneum and epidermis can be expanded by skin utilization of niosomes while limiting the foundational assimilation of medications. They are accepted to improve the horny layer's qualities by diminishing transepidermal water misfortune and by reestablishing lost skin lipids, which increments smoothness.[53], [54] After the layer corneum layer of skin, niosomes diffuses all in all and in the skin new more modest vesicles are shaped (re-development of niosome vesicles).[55],[56] On the outer layer of the skin, adsorption, and combination of niosomes drive a high thermodynamic action slope at the connection point for pervasion of lipophilic medications which fills in as a driving force.[57] The boundary of the layer corneum is defeacted by the impact of vesicles as entrance enhancers.

Methods to improve drug skin penetration A variety of strategies have been developed to increase skin permeability. Surfactants, which are the components of niosomes, increase transdermal permeation and percutaneous absorption by decreasing surface tension.[58] This incorporated; for instance, the use of an electric field; iontophoresis and electroporation, the utilization of compound enhancers, and the utilization of nanocarriers like liposomes, polymeric, and stable lipid nanoparticles (Table 3). The infiltration through layer corneum and focusing of growth cells can be further developed utilizing these methods.[60]

Table 3: Penetration of enhancers for anti-cancer delivery.

Anticancer Drug	Penetration methods	Penetration Enhancer/ Technique used/ Model	Highlights
Aminolevulinic acid	Physical Method	Iontophoresis/ <i>In vivo</i> : human	Significantly increased drug penetration
5-Aminolevulinic acid (5-ALA) and ALA derivatives	Chemical Method	DMSO and DMSO with EDTA/ Mouse Skin	Increased drug penetration
5-Aminolevulinic acid (5-ALA) and ALA derivatives	Chemical Method	Oleic acid/Mouse skin	Increased penetration of ALA in the skin
Atenolol	Chemical Method	Iontophoresis/ <i>in vitro</i> : pig ear	The highest permeation was attained when oleic acid was combined with iontophoresis
Cisplatin	Chemical Method	Monoolein and propylene Glycol/ <i>In vitro</i> : pig ear	Increased absorbance to the skin
Doxorubicin	Chemical Method	Monoolein and propylene Glycol	Improved drug retention in the Skin.
Doxorubicin	Physical Method	Iontophoresis/ <i>In vitro</i> : pig ear	Increased drug penetration
5-fluorouracil	Chemical Method	Azone, lauryl alcohol and isopropyl myristate Azone/ <i>In vitro</i> : pig ear	It improved drug flux through the Skin.
Meso-tetra-(N-methyl pyridinium-4-yl)-porphyrin and meso-tetra-(4-sulfonatophenyl)-porphyrin	Physical Method	Iontophoresis/ <i>In vitro</i> : pig ear	Increased drug skin penetration
Metoprolol	Chemical Method	Iontophoresis	Combination of iontophoresis with SLS further enhanced the drug delivery
Photosensitizer chlorin and phthalocyanine	Physical Method	Electroporation/ <i>In vitro</i> : pig ear	Increased drug skin penetration
Ruthenium Complex	Physical Method	Electroporation/ <i>In vitro</i> : melanoma cell line	Increased drug skin penetration
Tretinoin	Chemical Method	Liposomes combined with decylpolyglucosid, 8-glyceride, ethoxydiglycol, caprylocaproyl macrogol and propylene glycol	Improved drug Retention
Zinc phthalocyanine tetrasulfonic acid	Physical Method	Iontophoresis/ <i>In vitro</i> : pig ear	Significantly increased drug penetration

Conclusion

Niosomes, innovative drug carriers, show promise in dermatological applications, enhancing drug delivery efficiency while minimizing systemic absorption. They improve skin characteristics, such as reducing transepidermal water loss and restoring lost skin lipids. Niosomes diffuse through the stratum corneum and can reform as smaller vesicles within the skin, aiding in drug absorption. Techniques like iontophoresis, electroporation, and nanocarriers like liposomes and lipid nanoparticles, along with surfactants present in niosomes, enhance skin permeability and drug penetration. These methods offer the potential for targeted delivery and improved efficacy in treating skin disorders, including cancer.

Reference :

1. Allen TM. Liposomal drug formulations: Rationale for development and what we can expect for the future. *Drugs*. 1998;56:747–56. [[PubMed](#)] [[Google Scholar](#)]
2. Malhotra M, Jain NK. Niosomes as drug carriers. *Indian Drugs*. 1994;31:81–6. [[Google Scholar](#)]
3. Udupa N. Niosomes as drug carriers. In: Jain NK, editor. *Controlled and novel drug delivery*. 1st edition. New Delhi: CBS Publishers and Distributors; 2002. [[Google Scholar](#)]
4. Baillie AJ, Florence AT, Hume LR, Muirhead GT, Rogerson A. The Preparation and properties of Niosomes-Non ionic surfactant vesicles. *J Pharm Pharmacol*. 1985;37:863–8. [[PubMed](#)] [[Google Scholar](#)]
5. Kaur IP, Garg A, Singla AK, Aggarwal D. Vesicular systems in ocular drug delivery: An overview. *Int J Pharm*. 2004;269:1–14. [[PubMed](#)] [[Google Scholar](#)]
6. Hu C, Rhodes DG. Proniosomes: A Novel Drug Carrier Preparation. *Int J Pharm*. 1999;185:23–35. [[PubMed](#)] [[Google Scholar](#)]
7. Azmin MN, Florence AT, Handjani-Vila RM, Stuart JF, Vanlerberghe G, Whittaker JS. The effect of non-ionic surfactant vesicle (niosome) entrapment on the absorption and distribution of methotrexate in mice. *J Pharm Pharmacol*. 1985;37:237–42. [[PubMed](#)] [[Google Scholar](#)]
8. Szoka F, Jr, Papahadjopoulos D. Comparative properties and methods of preparation of lipid vesicles (liposomes) *Annu Rev Biophys Bioeng*. 1980;9:467–508. [[PubMed](#)] [[Google Scholar](#)]
9. Jadon PS, Gajbhiye V, Jadon RS, Gajbhiye KR, Ganesh N. Enhanced oral bioavailability of griseofulvin via niosomes. *AAPS PharmSciTech*. 2009;10:1186–92. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
10. Sheena IP, Singh UV, Kamath R, Uma Devi P, Udupa N. Niosomal withaferin A, with better tumor efficiency. *Indian J Pharm Sci*. 1998;60:45–8. [[Google Scholar](#)]
11. Baillie AJ, Coombs GH, Dolan TF, Laurie J. Non-ionic surfactant vesicles, niosomes, as delivery system for the anti-leishmanial drug, sodium stibogluconate. *J Pharm Pharmacol*. 1986;38:502–5. [[PubMed](#)] [[Google Scholar](#)]
12. Gregoriadis G. Targeting of drugs: Implications in medicine. *Lancet*. 1981;2:241–6. [[PubMed](#)] [[Google Scholar](#)]
13. Hunter CA, Dolan TF, Coombs GH, Baillie AJ. Vesicular systems (Niosome and Liposomes) for delivery of sodium stibogluconate in experimental murine visceral leishmaniasis. *J Pharm Pharmacol*. 1988;40:161–5. [[PubMed](#)] [[Google Scholar](#)]
14. Cummings J, Stuart JF, Calman KC. Determination of adriamycin, adriamycinol and their 7-deoxyaglycones in human serum by high-performance liquid chromatography. *J Chromatogr*. 1984;311:125–33. [[PubMed](#)] [[Google Scholar](#)]

15. Suzuki K, Soka K. The Application of Liposomes to Cosmetics. *Cosmetic and Toiletries*. 1990;105:65–78. [[Google Scholar](#)]
16. Alcantar N, Dearborn K, VanAuker M, Toomey R, Hood E. Niosome-hydrogel drug delivery. *US 2008/0050445A1*. 2008 [[Google Scholar](#)]
17. Brewer JM, Alexander J. The adjuvant activity of non-ionic surfactant vesicles (niosomes) on the BALB/c humoral response to bovine serum albumin. *Immunology*. 1992;75:570–5. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
18. Moser P, Marchand-Arvier M, Labrude P, Handjani -Vila RM, Vignerson C. Hemoglobin niosomes. I. Preparation, functional and physico-chemical properties, and stability. *Pharma Acta Helv*. 1989;64:192–202. [[PubMed](#)] [[Google Scholar](#)]
19. Moser P, Arvier MM, Labrude P, Vignerson C. Niosomes of hemoglobine. II. *Vitro* interactions with plasma proteins and phagocytes. *Pharm Acta Helv*. 1990;65:82–92. [[PubMed](#)] [[Google Scholar](#)]
20. Jayaraman SC, Ramachandran C, Weiner N. Topical delivery of erythromycin from various formulations: An *in vivo* hairless mouse study. *J Pharm Sci*. 1996;85:1082–4. [[PubMed](#)] [[Google Scholar](#)]
21. Khandare JN, Madhavi G, Tamhankar BM. Niosomes novel drug delivery system. *East Pharmacist*. 1994;37:61–4. [[Google Scholar](#)]
22. Mayer LD, Bally MB, Hope MJ, Cullis PR. Uptake of antineoplastic agents into large unilamellar vesicles in response to a membrane potential. *Biochem Biophys Acta*. 1985;816:294–302. [[PubMed](#)] [[Google Scholar](#)]
23. Naresh RA, Chandrashekhar G, Pillai GK, Udupa N. Antiinflammatory activity of Niosome encapsulated diclofenac sodium with Tween-85 in Arthritic rats. *Ind J Pharmacol*. 1994;26:46–8. [[Google Scholar](#)]
24. Rogerson A, Cummings J, Willmott N, Florence AT. The distribution of doxorubicin in mice following administration in niosomes. *J Pharm Pharmacol*. 1988;40:337–42. [[PubMed](#)] [[Google Scholar](#)]
25. Pardakhty A, Varshosaz J, Rouholamini A. *In vitro* study of polyoxyethylene alkyl ether niosomes for delivery of insulin. *Int J Pharm*. 2007;328:130–41. [[PubMed](#)] [[Google Scholar](#)]
26. Hao Y, Zhao F, Li N, Yang Y, Li K. Studies on a high encapsulation of colchicines by a niosome system. *Int J Pharm*. 2002;244:73–80. [[PubMed](#)] [[Google Scholar](#)]
27. Uchegbu IF, Vyas SP. Non-ionic surfactant based vesicles (niosomes) in drug delivery. *Int J Pharm*. 1998;172:33–70. [[Google Scholar](#)]
28. Almira I, Blazek-welsh IA, Rhodes GD. Maltodextrin – Based proniosomes. *AAPS PharmSciTech*. 2001;3:1–8. [[Google Scholar](#)]
29. Biswal S, Murthy PN, Sahu J, Sahoo P, Amir F. Vesicles of Non-ionic Surfactants (Niosomes) and Drug Delivery Potential. *Int J Pharm Sci Nanotech*. 2008;1:1–8. [[Google Scholar](#)]
30. Azmin MN, Florence AT, Handjani-Vila RM, Stuart JF, Vanlerberghe G, Whittaker JS. The effect of non-ionic surfactant vesicle (niosome) entrapment on the absorption and distribution of methotrexate in mice. *J Pharm Pharmacol*. 1985;37:237–42. [[PubMed](#)] [[Google Scholar](#)]
31. Manosroi A, Wongtrakul P, Manosroi J, Sakai H, Sugawara F, Yuasa M, et al. Characterization of vesicles prepared with various non-ionic surfactants mixed with cholesterol. *Colloids Surf B*. 2003;30:129–38. [[Google Scholar](#)]
32. Balasubramaniam A, Kumar VA, Pillai KS. Formulation and *in-vivo* evaluation of niosome encapsulated daunorubicin hydrochloride. *Drug Dev Ind Pharm*. 2002;28:1181–93. [[PubMed](#)] [[Google Scholar](#)]

33. Yoshioka T, Stermberg B, Florence AT. Preparation and properties of vesicles (niosomes) of sobitan monoesters (Span 20, 40, 60, and 80) and a sorbitan triester (Span 85) *Int J Pharm.* 1994;105:1–6. [[Google Scholar](#)]
34. Karki R, Mamatha GC, Subramanya G, Udupa N. Preparation, characterization and tissue disposition of niosomes containing isoniazid. *Rasayan J Chem.* 2008;1:224–7. [[Google Scholar](#)]
35. Uchegbu FI, Vyas PS. Non-ionic surfactant based vesicles (niosomes) in drug delivery. *Int J Pharm.* 1998;172:33–70. [[Google Scholar](#)]
36. Hunter CA, Dolan TF, Coombs GH, Baillie AJ. Vesicular system (niosomes and liposomes) for delivery of sodium stibogluconate in experimental murine visceral leishmaniasis. *J Pharm Pharmacol.* 1988;40:161–5. [[PubMed](#)] [[Google Scholar](#)]
37. Yoshioka T, Florence AT. Vesicle (niosome)-in-water-in-oil (v/w/o) emulsion an *in-vitro* study. *Int J Pharm.* 1994;108:117–23. [[Google Scholar](#)]
38. Cooper, Gunn . Emulsions and cream. In: Carter SJ, editor. *Dispensing for Pharmaceutical Students*. 12th edition. New York: CBS Publishers and Distributors; 2000. p. 128. [[Google Scholar](#)]
39. Arunothayanun P, Bernard MS, Craig DQ, Uchegbu IF, Florence AT. The effect of processing variables on the physical characteristics of nonionic surfactant vesicles (niosomes) formed from hexadecyl diglycerol ether. *Int J Pharm.* 2000;201:7–14. [[PubMed](#)] [[Google Scholar](#)]
40. Rogerson A. Adriamycin-loaded niosomes–drug entrapment, stability and release. *J Microencap.* 1987;4:321–8. [[PubMed](#)] [[Google Scholar](#)]
41. [https://www.thelancet.com/journals/lancet/article/PIIS0140-6736\(15\)60692-4/fulltext?elsca4=Public+Health%7CInfectious+Diseases%7CHealth+Policy%7CInternal%2FFamily+Medicine%7CGeneral+Surgery%7CLancet&code=lancet-site](https://www.thelancet.com/journals/lancet/article/PIIS0140-6736(15)60692-4/fulltext?elsca4=Public+Health%7CInfectious+Diseases%7CHealth+Policy%7CInternal%2FFamily+Medicine%7CGeneral+Surgery%7CLancet&code=lancet-site)
42. <https://academic.oup.com/chemlett/article-abstract/45/3/242/7337206>
43. <https://www.sciencedirect.com/science/article/pii/S0145212698000368>
44. <https://advances.umw.edu.pl/pl/article/2015/24/1/103/>
45. <https://www.nature.com/articles/nrc903>
46. <https://link.springer.com/article/10.1186/s12951-018-0351-4>
47. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6827193/>
48. <https://europepmc.org/article/med/8961159>
49. <https://www.sciencedirect.com/science/article/pii/S0169409X17300388>
50. <https://link.springer.com/article/10.1007/s12282-019-01014-w>
51. <https://www.sciencedirect.com/science/article/pii/S1742706115301227>
52. https://link.springer.com/chapter/10.1007/978-94-017-7215-0_20
53. https://link.springer.com/chapter/10.1007/978-94-017-7215-0_20
54. <https://onlinelibrary.wiley.com/doi/abs/10.1111/jcmm.14943>
55. <https://www.sciencedirect.com/science/article/pii/S0928098721003547#bib0014>
56. Effect of helper lipids on the interaction of DNA with cationic lipid monolayers studied by specular neutron reflection
- <https://pubs.acs.org/doi/abs/10.1021/bm300639n>

57.The role of the helper lipid dioleoylphosphatidylethanolamine (DOPE) for DNA transfection cooperating with a cationic lipid bearing ethylenediamine

<https://www.sciencedirect.com/science/article/pii/S0005273612003665>

58..Transdermal drug delivery: Innovative pharmaceutical developments based on disruption of the barrier properties of the stratum corneum

<https://www.mdpi.com/1999-4923/7/4/438>

59.,Synthesis and potential applications of lipid nanoparticles in medicine

<https://www.mdpi.com/1996-1944/15/2/682>

60.Emerging role of hydrogels in drug delivery systems, tissue engineering and wound management

<https://www.mdpi.com/1999-4923/13/3/357>

61.Iontophoretic transport of zinc phthalocyanine tetrasulfonic acid as a tool to improve drug topical delivery

https://journals.lww.com/anti-cancerdrugs/Fulltext/2011/09000/Iontophoretic_transport_of_zinc_phthalocyanine.9.aspx

62.Penetration enhancer containing vesicles as carriers for dermal delivery of tretinoin

<https://www.sciencedirect.com/science/article/pii/S0378517311003243>

63.The influence of positive or negative charges in the passive and iontophoretic skin penetration of porphyrins used in photodynamic therapy <https://www.sciencedirect.com/science/article/pii/S0939641110003127>

64.Doxorubicin skin penetration from monoolein-containing propylene glycol formulations

<https://www.sciencedirect.com/science/article/pii/S0378517306006946>

65.Delivery of a classical antihypertensive agent through the skin by chemical enhancers and iontophoresis

<https://onlinelibrary.wiley.com/doi/abs/10.1111/j.1600-0846.2009.00350.x>

66.A vehicle for photodynamic therapy of skin cancer: influence of dimethylsulphoxide on 5-aminolevulinic acid in vitro cutaneous permeation and in vivo protoporphyrin ...

<https://www.sciencedirect.com/science/article/pii/S0168365999002138>

67.Combined therapy of the antimetastatic compound NAMI-A and electroporation on B16F1 tumour cells in vitro

<https://www.sciencedirect.com/science/article/pii/S1567539407000424>

68.Effects of ionization and penetration enhancers on the transdermal delivery of 5-fluorouracil through excised human stratum corneum

<https://www.sciencedirect.com/science/article/pii/S0378517305002279>

69.Effect of the iontophoresis of a chitosan gel on doxorubicin skin penetration and cytotoxicity

<https://www.sciencedirect.com/science/article/pii/S0168365908007086>

70.Effect of permeation enhancers on the iontophoretic transport of metoprolol tartrate and the drug retention in skin

<https://www.tandfonline.com/doi/abs/10.3109/10717544.2010.509361>

71..Effect of permeation enhancers on the iontophoretic transport of metoprolol tartrate and the drug retention in skin

<https://www.tandfonline.com/doi/abs/10.3109/10717544.2010.509361>

72..Monoolein: a review of the pharmaceutical applications

<https://www.tandfonline.com/doi/full/10.1081/DDC-100101304>

73.Preparation and evaluation of reverse-phase evaporation and multilamellar niosomes as ophthalmic carriers of acetazolamide

<https://www.sciencedirect.com/science/article/pii/S0378517305006022>

74..A review on novel vesicular drug delivery: proniosomes

<https://www.tandfonline.com/doi/abs/10.3109/10717544.2013.841783>

75.Non-ionic surfactant vesicles, niosomes, as a delivery system for the anti-leishmanial drug, sodium stibogluconate

<https://academic.oup.com/jpp/article-abstract/38/7/502/6176315>

