



Quantification Of Favipiravir By Analytical Methods In Bulk And Capsule Dosage Form

¹*Mule, SR. & Hingane LD

¹Department of pharmaceutical analysis, Aditya College Of Pharmacy, Beed, Maharashtra, India

ABSTRACT:

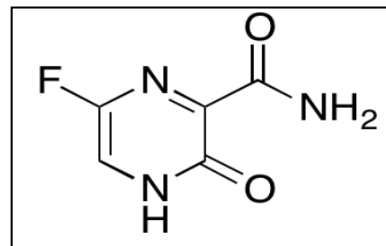
Favipiravir is an antiviral drug currently under development for the treatment of severe viral infections, including the coronavirus responsible for causing chronic diseases like COVID-19. In this study, we have developed and validated three different analytical methods, namely UV-Spectrophotometry (Method 1), NP-HPTLC (Method 2), and RP-HPLC (Method 3), for the quantitative estimation of Favipiravir in bulk and capsule dosage forms. These methods were found to be specific, sensitive, rapid, and cost-effective, making them suitable for routine analysis in pharmaceutical laboratories. Following the ICH guidelines, the developed methods were validated for linearity, accuracy, precision, ruggedness, and sensitivity.

KEYWORDS

Favipiravir, Covid-19, UV-Spectrophotometry, NP-HPTLC, RP-HPLC, Chronic & Antiviral.

INTRODUCTION

Favipiravir has emerged as a promising antiviral drug candidate in the treatment of COVID-19 and other viral infections. It is an isopropyl ester prodrug, which converts to an active nucleoside analogue (T-705-RTP or EIDD-1931) analogue of tent antiviral activity against various RNA viruses when metabolized in the plasma. The urgency of effective treatment options for COVID-19 has led to the Emergency Use Authorization (EUA) of Favipiravir by the U.S. Food and Drug Administration (FDA) for high-risk patients. Therefore, the development of reliable and efficient analytical methods for the quantification of Favipiravir in pharmaceutical formulations is crucial for quality control and therapeutic monitoring.



Favipiravir (DB12466) Structure:

| | |
|--------------------------|--|
| Molecular Formula | C ₅ H ₄ FN ₃ O ₂ |
| Molecular Weight | 157.104 g·mol ⁻¹ |
| Chemical Name | 6-fluoro-3-oxo-3,4-dihydropyridine-2-carboxamide |
| Description | Light yellow to yellow solid |
| Melting Point | 187–193 °C |
| Solubility | Slightly soluble in water |

Following the ICH guidelines, the developed methods were validated for linearity, accuracy, precision, ruggedness, and sensitivity agents

All chemicals and reagents were purchased from Merck Chemicals India's analytical grade, including Favipiravir standard, solvents, mobile phase components, and any other necessary materials.

2. Instrumentation

Mention the specific instruments used in each analytical method:

- UV-Spectrophotometry: SHIMADZU AUX – 120 (Weighing Balance) UV Shimadzu 2450 (PC Series) UV-visible double beam spectrophotometer.
- NP-HPTLC: HPTLC instrument and Camag TLC System with Linomat 5 Applicator used for the NP-HPTLC method.
- RP-HPLC: HPLC instrument (UFLC-LC 20 AD) equipped with a PDA detector used for the RP-HPLC method.

3. Standard Preparation

The stock standard solution was prepared by weighing 10 mg of Favipiravir. The weighed powder was transferred into a volumetric flask of 10 mL and dissolved and diluted to mark with methanol to obtain a concentration of n 1 mg/mL.

4. Method 1: Quantitative Estimation of Favipiravir by UV-Spectrophotometric Method

a. Zero Order UV-Spectrophotometric Method (Method A)

- The wavelength (235 nm) used for the analysis.
- The linearity of an analytical method is its ability to elicit test results that are directly, or by a well-defined mathematical transformation, proportional to the concentration of analyte in the sample within the range. Appr volumese volumes in the range of 0.5-3.0 mL were transferred from the stock solution into a series of 10 mL volumetric flasks and volumes were made up to mark with the Methanol: Water (60:40 v/v) mobile phase to the concentration in the range of 5-30 µg/mL. A constant volume of 20 µL for each sample was injected with the help of Hamilton Syringe. All measurements were repeated five times for each concentration and a calibration curve was constructed by plotting the peak area versus the drug concentration
- To assess the accuracy of the proposed method, recovery studies were carried out at three different levels i.e. 80, 100 and 120%. To pre-analyse sample solution a known amount standard rd drug solution was added at three different levels, and absorbance was recorded.

b. UV-Spectrophotometry using AUC (Method B)

- The wavelength range (228.00 – 243.40 nm) was used for AUC calculations.
- From the stock solutions, 1 mL of FVP was transferred to a 10 mL volumetric flask and the volume was adjusted to the mark with the same solvent to obtain concentrate ion 10 µg/mL. The solution was scanned in the UUV of ange 400 - 200 nm. AUC was selected in the wavelength range of 228.00 – 243.40 nm.

5. Method 2: Development and Validation of NP-HPTLC Method for Quantitative Estimation of Favipiravir in Bulk and Capsule Dosage Form

- NP-HPTLC aluminium plates precoated with silica gel 60-F254 TLC plates used.
- Mobile phase composition (Acetone: Chloroform: Formic acid, 4:6:0.1 v/v).
- The densitometric quantification for these drugs was carried out at 235 nm. Favipiravir obeyed linearity in the range of 500 – 3000 ng/band. The R_f of Favipiravir was found to be 0.55.

6. Method 3: RP-HPLC Method Development and Validation for Estimation of Favipiravir in Bulk and Capsule Dosage Form

- LC-GC Qualisil BDS C8 column (250 mm x 4.6 mm, 5 µm) used for separation.
- Mobile phase composition (Methanol: Water, 60:40 v/v) and the flow rate (1 ml/min).
- Wavelength (235 nm) at which the analyte was monitored.
- Retention time (4.1 min) of Favipiravir and the range of concentrations used for linearity (5 - 30 µg/mL).

7. Validation of Methods

Accuracy

To assess the accuracy of the proposed method, recovery studies were carried out a three different levels i.e. 80, 100 and 120%. Absorbapre-analyzed to the pre - analyzed sample solution, a known amount standard drug solution was added at three different levels The results are reported in **Tables 4.1.4 and 4.1.5.**

Table 4.1.4: % Recovery Studies (Method A)

| Drug | Initial Amount [µg/mL] | Amount added [µg/mL] | Amount Recovered [µg/mL, n=3] | % Recovered | % RSD |
|------|------------------------|----------------------|-------------------------------|-------------|-------|
| FVP | 15 | 12 | 11.99 | 99.97 | 0.23 |
| | 15 | 15 | 14.90 | 99.34 | 0.23 |
| | 15 | 18 | 17.84 | 99.15 | 0.17 |

Table 4.1.5: % Recovery Studies (Method B)

| Drug | Initial Amount [µg/mL] | Amount added [µg/mL] | Amount Recovered [µg/mL, n=3] | % Recovered | % RSD |
|------|------------------------|----------------------|-------------------------------|-------------|-------|
| FVP | 15 | 12 | 11.94 | 99.50 | 0.14 |
| | 15 | 15 | 14.93 | 99.56 | 0.13 |
| | 15 | 18 | 17.93 | 98.85 | 0.37 |

Precision

Precision of the method is studied as repeatability, intra-day and inter-day precision. Repeatability was determined by analyzing FVP (15 µg/mL) for six times and the results are reported in **Table 4.1.6**. Intra-day precision was determined by analyzing the 10, 15 and 20 µg/mL of FVP for three times in the same day.

Inter-day precision was determined by analyzing the same concentration of the solutions daily for three days, results are reported in **Table 4.1.7**.

Table 4.1.6: Repeatability Studies

| Drug | Amount Taken [µg/mL] | Method A | Method B | | |
|--------------|-------------------------|-------------------------|-------------------------|-------------------------|----------------------|
| | | Amount found [µg/mL] | % Amount found [n=6] | Amount found [µg/mL] | % Amount found |
| FVP | 15 | 15.06 | 100.46 | 15.09 | 100.65 |
| | 15 | 15.04 | 100.30 | 15.02 | 100.19 |
| | 15 | 14.88 | 99.21 | 15.07 | 100.52 |
| | 15 | 15.21 | 101.42 | 15.06 | 100.46 |
| | 15 | 15.09 | 100.66 | 15.12 | 100.84 |
| | 15 | 14.91 | 99.46 | 15.14 | 100.95 |
| | Mean ± SD | 15.03 ± 0.12 | 100.2 ± 0.81 | 15.09 ± 0.03 | 100.60 ± 0.24 |
| % RSD | 0.80 | 0.80 | 0.24 | 0.24 | |

n-number of determinations

Table 4.1.7: Precision Studies [Intra-day and Inter-day]

| Standard Concentration (µg/mL) | Method A | | | Method B | | |
|-----------------------------------|-------------------------|----------------|-------|-------------------------|----------------|-------|
| | Amount Found [µg/mL] | % Amount found | % RSD | Amount found [µg/mL] | % Amount found | % RSD |
| Intra-day Precision | | | | | | |
| 10 | 10.1154 | 101.15 | 1.84 | 10.0432 | 100.43 | 0.073 |
| 15 | 14.7642 | 98.42 | 1.43 | 14.8212 | 98.80 | 1.30 |
| 20 | 19.9926 | 99.96 | 0.45 | 19.7255 | 98.62 | 1.068 |
| Inter-day Precision | | | | | | |
| 10 | 9.7073 | 97.07 | 1.31 | 9.8297 | 98.29 | 1.69 |
| 15 | 15.0081 | 100.05 | 1.67 | 14.8921 | 99.28 | 0.71 |
| 20 | 19.1780 | 95.89 | 0.64 | 19.30 | 96.5 | 1.09 |

Sensitivity

Sensitivity of the proposed method was estimated in terms of Limit of Detection (LOD) and Limit of Quantification (LOQ). The LOD and LOQ were calculated by the use of the equation $LOD = 3.3 \times ASD/S$ and $LOQ = 10 \times ASD/S$; where, 'ASD' is Average standard deviation of the peak height and areas of the drug ($n = 3$), taken as a measure of noise, and 'S' is the slope of the corresponding calibration curve.

Different volume of stock solution in the range 5-10 $\mu\text{g/mL}$ was prepared. The procedure was repeated in triplicate. LOD and LOQ was found to be **1.1 μg** and **3.3 μg** (Method A), and **1.2 μg** and **3.6 μg** (Method B), respectively.

Ruggedness

Ruggedness of the proposed method is determined by analysis of aliquots from homogenous slot by two analysts using same operational and environmental conditions and the results are reported in **Table 4.1.8.**

Table 4.1.8: Ruggedness Studies

| Method A | | Method B | | |
|----------|---|----------|--|-------|
| Analysts | [%] Amount found \pm SD [n= 6] | % RSD | [%] Amount found \pm SD [= 6] | % RSD |
| I | 99.85 \pm 0.40 | 0.30 | 100.36 \pm 0.97 | 0.16 |
| II | 99.76 \pm 0.43 | 0.34 | 100.20 \pm 0.95 | 0.35 |

n-number of determinations

Method 2

Precision studies

Repeatability, Intra-day and Inter-day precisions were perceived using six repetitive measurements in target concentration level.

The precision of the proposed method was ascertained by actual determination of three replicates of 1000, 1500 and 2000 ng/band concentration of Favipiravir finding out the result by the proposed method. Repeatability expresses the precision under the same operating conditions over a short interval of time.

Repeatability is also termed intra-assay precision. So, the intra assay precision has been executed by analyzing samples 1500 ng/band of Favipiravir for six times. The precision of developed method was evaluated in terms of % RSD.

The results are shown in **Table 4.2.4 and 4.2.5.**

Table 4.2.4: Precision Studies [Intra and Inter-day]

| Standard Concentration (ng/band) | Amount Found [ng/band] [n=3] | % Amount found | % RSD |
|----------------------------------|------------------------------|----------------|-------|
| Intra-day Precision | | | |
| 1000 | 999.44 | 99.94 | 0.67 |
| 1500 | 1483.37 | 98.89 | 0.10 |
| 2000 | 1984.42 | 99.22 | 1.94 |
| Inter-day Precision | | | |
| 1000 | 990.84 | 99.08 | 1.58 |
| 1500 | 1459.03 | 97.26 | 1.89 |
| 2000 | 1957.99 | 97.89 | 0.69 |

n- number of determinations **Table 4.2.5:**

Repeatability Studies

| Drug | Amount taken [ng/band] | Amount found [ng/band] | % Amount found |
|--------------|------------------------|------------------------|----------------|
| FVP | 1500 | 1493.32 | 99.55 |
| | 1500 | 1493.10 | 99.54 |
| | 1500 | 1492.19 | 99.47 |
| | 1500 | 1492.67 | 99.51 |
| | 1500 | 1492.94 | 99.52 |
| | 1500 | 1492.04 | 99.46 |
| | Mean ± SD | | 1492.71 ± 0.51 |
| % RSD | | 0.034 | 0.034 |

Accuracy

Accuracy study was executed by standard addition method using three different levels. Recovery experiment was evaluated by over spotting the drug standard at 80 %, 100 % and 120 % to the pre-analyzed sample and the results were re-analyzed by proposed HPTLC method. The experiment was repeated three times.

The results are shown in **Table 4.2.6.**

Table 4.2.6: Recovery Studies

| Drug | Initial Amount [ng/band] | Excess drug added to the analyte | Total amount found \pm S.D. [$\mu\text{g/mL}$] | Recovery [%] [n=3] | %RSD [n = 3] |
|------|--------------------------|----------------------------------|--|--------------------|--------------|
| FVP | 1500 | 1200 | 2683.76 \pm 48.27 | 98.64 | 0.38 |
| | 1500 | 1500 | 2993.03 \pm 53.48 | 99.53 | 0.38 |
| | 1500 | 1800 | 3268.61 \pm 50.28 | 98.25 | 0.33 |

n- number of determinations Ruggedness

The ruggedness of the method was performed by two different analysts using same operational and environmental conditions. The ruggedness of the proposed method was determined by 1500 ng/band concentration of Favipiravir.

The results are shown in Table 4.2.7.

Table 4.2.7: Ruggedness studies

| Drug | Concentration n[ng/band] | Amount Found (%) \pm S.D. | |
|------|--------------------------|-----------------------------|--------------------|
| | | Analysts- I [n=6] | Analysts- II [n=6] |
| FVP | 1500 | 99.05 \pm 0.59 | 98.76 \pm 0.81 |

n-number of determinations

Method 3

Accuracy

To assess the accuracy of the proposed method, recovery studies were carried out three different levels i.e. 80, 100 and 120%. To the pre - analyzed sample solution a known amount standard drug solution was added at three different levels, absorbance was recorded. The results are reported in Table 4.1.5.

| Drug | Initial Amount [$\mu\text{g/mL}$] | Excess Drug Added to the Analyte | Total Amount Found \pm SD [$\mu\text{g/mL}$] | Recovery [%] [n=3] | % RSD [n=3] |
|------|-------------------------------------|----------------------------------|--|--------------------|-------------|
| FVP | 15 | 12 | 26.77 \pm 3169.56 | 98.09 | 0.18 |
| | 15 | 15 | 29.73 \pm 2158.95 | 98.25 | 0.11 |
| | 15 | 18 | 32.86 \pm 1089.69 | 99.24 | 0.05 |

Precision

The precision of the method was studied as repeatability, intra-day and inter-day variations. The precision of the proposed method was ascertained by actual determination of three replicates of 10, 15 and 20 µg/mL concentrations of FVP. The precision of the developed HPLC method was found to be precise as the RSD values for repeatability and intra-day and inter-day precision studies were < 2%, respectively as recommended by ICH guidelines.

Repeatability was measured by analyzing 15 µg/mL of Favipiravir for six times. The results are shown in Table 4.3.6 and Table 4.3.7.

Table 4.3.6: Precision Studies [Intra and Inter-Day]

| Standard Concentration [µg/mL] | Amount Found [µg/mL] | % Amount Found [µg/mL] [n=3] | % RSD |
|--------------------------------|----------------------|------------------------------|-------|
| Intra-day Precision | | | |
| 10 | 9.72 | 97.24 | 0.92 |
| 15 | 14.88 | 99.21 | 0.24 |
| 20 | 19.42 | 97.10 | 1.50 |
| Inter-day Precision | | | |
| 10 | 9.76 | 97.69 | 0.81 |
| 15 | 14.86 | 99.10 | 0.17 |
| 20 | 19.25 | 96.28 | 1.55 |

n=number of determinations

Table 4.3.7: Repeatability Studies

| Drug | Amount Taken [µg/mL] | Amount Found [µg/mL] [n=6] | % Amount Found |
|------|----------------------|----------------------------|---------------------|
| FVP | 15 | 14.91 | 99.44 |
| | 15 | 14.85 | 99.02 |
| | 15 | 14.85 | 99.02 |
| | 15 | 14.84 | 98.95 |
| | 15 | 14.84 | 98.95 |
| | 15 | 14.99 | 99.99 |
| | Mean ± SD | 14.88 ± 0.06 | 99.23 ± 0.41 |

Sensitivity

The sensitivity measurements of Favipiravir by use of the proposed methods were estimated in terms of the Limit of Detection (LOD) and Limit of Quantification (LOQ). The LOD and LOQ were calculated using equation $LOD = 3.3 \times N/B$ and $LOQ = 10 \times N/B$, where 'N' is the standard deviation of the results of the drugs (n=3), taken as a measure of noise, and 'B' is the slope of the corresponding calibration curve. Different concentrations i.e. 5, 6, 7, 8, 9 and 10 µg/mL were selected for the method. The Average Standard Deviation (A.S.D.) of peak areas was determined. For determination LOD and LOQ slope of the corresponding calibration curve was considered. LOD and LOQ were found to be **0.44 µg** and **1.34 µg** for Favipiravir.

Ruggedness

An appropriate concentration of Favipiravir (15 µg/mL) were prepared and analyzed by two different analysts using similar operational and environmental conditions.

Peak area was measured for same concentration solutions, six times. The results are shown in **Table 4.3.8.**

Table 4.3.8: Ruggedness Studies

| Drug | Concentration [µg/mL] | Amount Found (%) ± SD | |
|------|--------------------------|-----------------------|---------------------|
| | | Analyst-I [n=6] | Analyst-II [n=6] |
| FVP | 15 | 98.87 ± 0.179 | 98.18 ± 1.294 |

n= number of determinations

10. Data Analysis

Method 1: UV-Spectrophotometry

Data Analysis:

- The data obtained from the UV-Spectrophotometer involved measuring the absorbance of Favipiravir at a specific wavelength (235 nm).
- Linearity was established by plotting a calibration curve using different concentrations of Favipiravir standard solutions. The data points were fitted to a linear regression equation to determine the relationship between concentration and absorbance.
- The correlation coefficient (r^2) was calculated to assess the goodness of fit of the calibration curve.

Software Used for Data Processing and Calculations:

- Microsoft Excel: Excel was used to organize the raw data, perform calculations for the calibration curve, and plot the graphical representation of the calibration curve.

Method 2: NP-HPTLC**Data Analysis:**

- The data obtained from NP-HPTLC involved recording the peak areas of separated spots for Favipiravir in the samples.
- Linearity was established by plotting a calibration curve using different concentrations of Favipiravir standard solutions. The data points were fitted to a linear regression equation to determine the relationship between concentration and peak area.
- The correlation coefficient (r^2) was calculated to assess the goodness of fit of the calibration curve.

Software Used for Data Processing and Calculations:

- Camag HPTLC Software: The Camag HPTLC Software was used to analyze the raw data obtained from NP-HPTLC, calculate peak areas, construct the calibration curve, and visualize the data graphically.

Method 3: RP-HPLC**Data Analysis:**

- The data obtained from RP-HPLC involved chromatographic peaks corresponding to Favipiravir at a specific retention time (4.1 min).
- Linearity was established by plotting a calibration curve using different concentrations of Favipiravir standard solutions. The data points were fitted to a linear regression equation to determine the relationship between concentration and peak area or height.
- The correlation coefficient (r^2) was calculated to assess the goodness of fit of the calibration curve.

Software Used for Data Processing and Calculations:

- HPLC Data Processing Software: The HPLC Data Processing Software associated with the UFLC-LC 20 AD HPLC system was utilized for data processing, peak integration, calibration curve construction, and visualization of the results.

Overall Data Interpretation:

For all methods, the results obtained from the data analysis were interpreted to validate the linearity of the calibration curves and the accuracy of the methods for quantifying Favipiravir in bulk and capsule dosage forms. The correlation coefficients were used to assess the goodness of fit, and high correlation values close to 1 indicated excellent linearity between the concentrations and the corresponding response (absorbance, peak area, or height).

RESULTS**Method 1****Quantitative Estimation Of Favipiravir By UV- Spectrophotometric Method**

Two simple, specific, sensitive, rapid and economical UV-Spectrophotometric methods have been established for the determination of Favipiravir in bulk and capsules. **Method A** is zero order UV-Spectrophotometry using absorbance, and **Method B** is zero order UV-Spectrophotometry using AUC. The anticipated methods were effectively applied for the determination of Favipiravir in bulk and capsules. In water, Favipiravir showed maximum absorbance at **235 nm**. In **Method A**, the absorbance was recorded at 235 nm, while in **Method B**, AUC was selected in the wavelength range of **228.00 – 243.40 nm**. In all methods, Favipiravir followed linearity in the concentration range of 5-30 µg/mL with ($r^2 > 0.999$). All these developed methods were applied for the estimation of Favipiravir capsules. All these methods were validated for linearity and range, accuracy, precision, ruggedness and sensitivity.

The summary of all these methods are given in **Table 5.1 Table 5.1:**

Summary of developed method

| Parameters | | Methods | |
|----------------------|--------------------|---------|-------|
| | | A | B |
| Accuracy [n=3] | 80% | 99.97 | 99.50 |
| | 100% | 99.34 | 99.56 |
| | 120% | 99.15 | 98.85 |
| Precision (% RSD) | Intra-day (n=3) | 1.24 | 0.81 |
| | Inter-day (n=3) | 1.20 | 1.16 |
| Repeatability (%RSD) | | 0.80 | 0.24 |
| Ruggedness (%RSD) | Analyst-I | 0.30 | 0.16 |
| | Analyst-II | 0.34 | 0.35 |
| LOD (µg) | | 1.1 | 3.3 |
| LOQ (µg) | | 1.2 | 3.6 |

Method 2

Development and Validation of NP-HPTLC Method for Quantitative Estimation of Favipiravir in Bulk and Capsule Dosage Form

A NP-HPTLC method has been developed for the determination of Favipiravir pharmaceutical formulation. The separation of Favipiravir was performed on NP- HPTLC aluminum plates precoated with silica gel 60-F254 TLC plates using **Acetone:Chloroform: Formic acid (4:6:0.1 v/v)** as mobile phase. The densitometric quantification for these drugs was carried out at 235 nm. Favipiravir obeyed linearity in the range of 500 – 3000 ng/band. The R_f of Favipiravir was found to be 0.55. The proposed method was applied for pharmaceutical formulation and % amount found for Favipiravir was found to be 99.45 ± 0.042 . The method was validated for accuracy, precision, and ruggedness. Accuracy of the method was checked by recovery studies at three different levels i.e. 80 %, 100 %, and 120 %. The % recovery of Favipiravir was found to be in the range of 98.25-99.53 %; the % RSD values were less than 2 indicate the accuracy of the method. The method was found to be precise as indicated by the inter-day, intra-day and repeatability analysis; showing % RSD less than 2. The results did not show any statistical difference between operators suggesting that method developed was rugged. The results of the developed method are shown in **Table 5.2**.

Method 3

The RP-HPLC method was developed and validated for the estimation of Favipiravir in bulk and capsule dosage form. The separation was achieved using a LC-GC Qualisil BDS C8 column (250 mm x 4.6 mm, 5 µm). During separation mobile phase consist of Methanol and Water (60:40 v/v) was delivered at a rate of 1 ml/min. The analyte was monitored with PDA detector at 235 nm. The method was found to be linear in the range of 5 to 30 µg/ml. The retention time for analyte was found 4.1 min. The linear response is observed in the range of 5 - 30 µg/mL. A same optimized method has successively been applied for the determination of Favipiravir in the capsules formulation. The drug content for Favipiravir was found to be $99.25 \pm 0.18\%$. Accuracy of the method was studied by the recovery studies at three different levels i.e. 80 %, 100 %, and 120 % level. The % recovery was found to be within the limits of the acceptance criteria within range of 98.09– 99.24%. The precision of the method was studied in terms of repeatability, intra-day, and inter-day precision. The results were examined as % RSD values of concentration of drug determined. The low value of %RSD (less than 2) indicates high precision of the method. The method proved to be adequately sensitive as indicated by low values of DL and QL. The Summary of the developed Method shown in **Table 5.2**.

Table 5.2: Summary of developed methods

| Parameters | NP-HPTLC | RP-HPLC |
|-------------------------|-------------------------|-------------------|
| Drug | FVP | FVP |
| Linearity range | 500 – 3000 (ng/band) | 5 - 30 (µg/ml) |
| Correlation coefficient | 0.999 | 0.999 |
| LOD | 47.51 ng/band | 0.44 µg |
| LOQ | 143.99 ng/band | 1.34 µg |

| | | |
|----------------------------|-------|-------|
| % Recovery (n=3) | 98.80 | 98.52 |
| Precision (%RSD) | | |
| Intra-Day(n = 3) | 0.90 | 0.88 |
| Inter-Day(n = 3) | 1.38 | 0.84 |
| Repeatability (n=6) | 0.034 | 0.41 |
| Ruggedness (%Amount Found) | | |
| Analyst-I | 99.05 | 98.87 |
| Analyst-II | 98.76 | 98.18 |

CONCLUSION:

Three methods RP-HPLC, NP-HPTLC and UV-Spectrophotometry (Zero order spectroscopy and zero order AUC) have been developed for Favipiravir in bulk and capsules. RP-HPLC and HPTLC methods are found to be accurate, precise, rugged and robust. Both these methods are adequately sensitive. Two UV-Spectrophotometric methods using absorbance and Area Under Curve techniques has been developed for estimation of Favipiravir in bulk and capsules. UV-Spectrophotometry methods are simple, accurate and economical and least calculations are involved for estimation of concentrations of Favipiravir in bulk and capsules. All these methods may regularly be used for estimation of Favipiravir in its pharmaceutical formulation

REFERENCES:

1. D. A. Skoog, D. M. West, F. J. Holle, and S. R. Crouch.,(2014). Fundamentals of Analytical Chemistry,9th ed., Brooks/Cole, Cengage Learning, pp.1-6
2. D. A. Skoog, T. A. Nieman, F. J. Holler,(1998).Principles of instrumental analysis, 5th ed., London Stanford University, Saunders College Publications, pp. 1-4.
3. S. Ahuja, N. Jespersen.,2006). Comprehensive analytical chemistry, pp.1-2.
4. G. D. Christian.,(2001). Analytical Chemistry, 5th ed., John Wiley and Sons, pp. 1-3.
5. A. H. Beckett, J. B. Stenlake.,(2002).Practical Pharmaceutical Chemistry, 4th ed., New Delhi, CBS Publishers and Distributors, pp 345-347
6. C.H. Bryant, A. Adam, D.R.Taylor and R.C. Rowe,(1994). A review of expert systems for chromatography. *Analytica chimica acta*, 297(3), p.317-347.
7. E. Kennedler,(2004).Introduction to chromatography. Institute for Analytical Chemistry, University of Vienna, p. 46.
8. PD, Shethi, (1997).HPLC-Quantitative analysis of pharmaceutical formulations, 3rd edition, CBS publishers & distributors, p.182.
9. S.R. Abbott, (2001). Sample preparation for normal and reversed phase analysis, *Journal of Chromatography A*, pp. 203-206.
10. J Swadesh,(1997). HPLC –Practical and Industrial Applications, CRC Press, Boca Raton, pp. 20-25.
11. A.V. Kasture, S.G. Wadodkar, K.R. Mahadik, & H.M More.,(2007). Pharmaceutical analysis, volume II, 17th edition, pp.48-57.
12. B. K Sharma, (2002).Instrumental Method of Chemical Analysis, 21th ed., Goel Publishing Housing, p. 3.
13. A. K. Connors, (1999). Textbook of Pharmaceutical Analysis, 3 rd ed., A Wiley Inter sciences Publication, p. 616.
14. H. H Willard, L. L. Metritt, J. A. Dean, and F. A. Settal, (1986). Instrumental method of analysis, 7 th edn ., CBS Publishers and Distributors, p. 59.
15. A.T. Giese and C.S French.,(1955).The analysis of overlapping spectral absorption bands by derivative spectrophotometry. *Applied Spectroscopy*, 9 (2), pp.78-96
16. J. A. Hanley and B. J. McNeil, (1983). A method of comparing the areas under receiver operating characteristic curves derived from the same cases. *Radiology*, 148(3), pp.839-843.
17. J.Kenkel.,(2003).Analytical Chemistry for Technicians, 3rd ed., Lewis Publishers, pp. 1- 16.
18. J. Myerson, L. Green and Warusa with arana M., (2001). Area under the curve as a measure of discounting. *Journal of the experimental analysis of behavior*, 76(2), pp.235-243.
19. G. R. Chatwal, S. K. Anand, (2003). Instrumental method of chemical analysis, 5th ed., Mumbai, Himalaya Publishing House, pp. 234-239.
20. M. W. Raynor, (1994). Liquid Chromatography for the Analyst, Marcel Dekker Inc. New York, pp. 2-3.
21. L.R. Snyder, M. A. Stadalius,(1983).High-Performance Liquid Chromatography: Advances and Perspectives, C. Horvath, 2nd ed., Academic Press, San Diego,3, 157-158.
22. M. Willard, S. Dean, (1986).Instrumental method of analysis,7th ed., New Delhi, CBS publication and distributors, pp.3-5, 684-689.
23. A. R. Gennaro, Remington: The Science and Practice of Practice of Pharmacy, 21st ed., volume II, Lippincott, Williams, and Welkins Mack Publishing Company, pp. 1680-1682.
24. R. A. Lodder, Analytical Spectroscopy Research Group, Advanced Science & Technology Center, University of Kentucky available from site: www.pharm.uky.edu/ASRG/HPLC/Histry.html.
25. S. Lindsay, (1997). High Performance Liquid Chromatography,John Wiley and Sons Publication, pp. 14-15.
26. J. W. Munson, (2001).Pharmaceutical Analysis modern methods, 1st ed., Marcel Dekker series of books, pp. 30-50.
27. M. W. Dong, (2006), Modern HPLC for Practicing Scientist, John Wiley, and Sons, pp.1-13.
28. P. D. Sethi,(1996), HPTLC: Quantitative Analysis of Pharmaceutical Formulation, CBS Publications, New Delhi, pp. 162-165.
29. E Stahl, Thin Layer Chromatography: A Laboratory Handbook, 2nd ed., Springer International Edition, pp. 1-30
30. P. E. Wall, (2005).Thin-layer Chromatography (A Modern Practical Approach), VWR International Ltd., Poole, Dorset, pp. 115-118.
31. www.camag.com

32. B. K. Sharma, (2002). Instrumental Method of Chemical Analysis, 21th ed., Goel Publishing Housing, p. 3.
33. H. H Willard, L. L. Metritt, J. A. Dean and F. A. Settal, (1986), Instrumental methods of Analysis, 7 th edn ., CBS Publishers and Distributers, p. 118.
34. A.T. Giese and C. S. French, (1955). The analysis of overlapping spectral absorption bands by derivative spectrophotometry. Applied Spectroscopy, 9 (2), pp.78-96.
35. G. Talsk, L. Mayring, and H. Kreuzer,(1978). High-Resolution, Higher-Order UV/VIS Derivative Spectrophotometry. Angewandte Chemie International Edition in English, 17(11), pp.785-799.
36. J .A. Hanley, and B.J McNeil, (1983). A method of comparing the areas under receiver operating characteristic curves derived from the same cases. Radiology, 148(3), pp.839-843.
37. J. Myerson, L. Green and Warusa with arana, M., (2001). Area under the curve as a measure of discounting. Journal of the experimental analysis of behavior, 76(2), pp.235-243
38. US pharmacopoeia,(1999).Validation of compendia methods, section, united states pharmacopeial convention, Rockville, MD, p. 2149
39. International Conference on Harmonization (ICH), (1997) Q2b: Validation of analytical procedure: Methodology, USFDA federal register, Vol. 62, p. 27463
40. US pharmacopoeia 23, (1994).Chromatography section, united states Pharmacopeial Convention, Rockville, MD, p. 1776
41. International Conference on Harmonization (ICH),(1995). Q2A: Text on Validation of Analytical Procedure USFDA federal register, vol 60, p. 11260
42. G. A. Shabir, (2003).Validation of high-performance liquid chromatography methods for pharmaceutical analysis: Understanding the differences and similarities between validation requirements of the US Food and Drug Administration, the US Pharmacopeia, and the International Conference on Harmonization. Journal of Chromatography A, 987(1), pp.57-66

