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A Concise Review On Copper Nanoparticles For Drug Delivery Against Luliconazole Resistant Bacterial Strains

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ABSTRACT: In present investigation, luliconazole coupled copper nanoparticles were synthesized to overwhelm drug resistance in Staphylococcus aurous, responsible for dermal skin infections. Luliconazole, copper sulphate, Tri sodium citrate are used. Copper nanoparticles were produced by size reduction method by using Copper Sulphate and Tri sodium citrate. CuNPs were merged into gel base of carbopol which was formed by hot method. Carbapol gel shows no phase separation. The particle size of CuNPs was found to be 413.0± 30.2nm. The % EE was found to be 65.2%. On description unilamellar, sphere-shaped vesicles with soft surface were detected under transmission electron microscopy. XRD of CuNPs was found out crystalline in nature. The zeta potential of nanoparticle shows less aggregation of particle with 15.1 ± 3.69 mV value. The amount of copper content was measured 5.9 microgram/10 mg of nanoparticles. In vitro release study of Cu NPs shows 96.5% release of drug and show effective antibacterial activity against Staphylococcus aureus. Conclusion: Luliconazole copper nanoparticles were produced and show excellent antibacterial activity upon S. aureus.

KEYWORDS: Copper Nanoparticle, Luliconazole, Carbopol gel, Antibacterial activity, Chemical reduction method, Copper sulphate.

INTRODUCTION

In the past decade, the cure of sickness has been consummate by administrating drugs to human body through different routes likes parental, oral, topical, sublingual, inhalation, rectal etc. The delivery of drug through topical denotes the application of drug onto the body employing vaginal, rectal, ophthalmic and skin as the route of administration. On human body, skin is widely used and accepted route for local application and constitutes the principal administration for local application. 1

The term topical drug delivery means administration for medicament containing formulation to the skin to openly care for the cutaneous manifestations of a common illness (e.g. psoriasis) or cutaneous disorders with the purpose of confining the pharmacological or other effect of the medicament inside or surface of skin.2 During recent years, a report will show interest in the synthesis and applications of various metallic nanoparticles due to their outstanding optical and electronic properties, especially copper, gold and silver nanoparticles. Copper nanoparticles (CuNPs) have gradually become an active area of research because of unique chemical, physical, electrical and optical properties, low cost, ease availability and exhibit good antibacterial properties. The prime advantage of CuNPs is their low cost and its availability compared to gold and silver nanoparticles, resulting in the sample synthesis and various applications of CuNPs.3, 4

Copper is easily available metal and one of the vital trace elements for mainly living creature. Copper was used as potential antimicrobial agent from ancient times.[5] Copper and its complexes used as a disinfectants, antiviral as well as antibacterial from centuries. It is said that the enhanced antimicrobial activity of Cu-NPs due to their crystallographic surface structure and large surface to volume ratio compared with copper salts. $Cu(OH)_2$ and $CuSO_4$ are used as the conventional inorganic antibacterial agents. Also, complex copper species, aqueous copper solutions or copper containing polymers are used as antifungal compounds as well as antibacterial. At current, advancement in new antibacterial agents is essential due to steady raise of new bacterial strains resistant to the potent antibiotics. Substances with low molecular weight like copper nanoparticles generally inhibit the growth or kill a wide range of bacterium bacteria. Copper ions shows antimicrobial activity across a wide range of microorganisms, such as Salmonella enteric, Staphylococcus aureus, Campylobacter jejuni, Listeria monocytogenes and Escherichia coli. The surfaces of copper can be used to kill viruses, yeasts and bacteria hence copper known as “contact killing”.6,7 The Cu particles in nano range have been shown a antibacterial effect on the microbial cell functions in numerous ways, including electrostatic interaction between particles and gram negative bacteria cell wall, denaturation of the intracellular proteins and interaction with phosphorus- and sulfur containing compounds like DNA. The nanoparticles passed through bacteria cell membrane and then injurious for the vital enzymes of bacteria can be the primary mechanism of anti- microbial action in CuNPs.8,9

MATERIALS AND METHODS

The following chemicals were used: Luliconazole (Combiotic Pvt. Ltd. India), copper sulphate (Thermo Fisher Scientific, India), Tri sodium citrate (Nice chemicals, India.) Propylene glycol (S.D. Fine Chem. India), Methanol (Loba Chemie, India), Ethanol (Loba Chemie, India), Carbopol 940 (Lubrizol advanced material, Belgium), Methyl paraben (Lubrizol advanced material, Belgium), Propyl paraben (Lubrizol advanced material, Belgium), Triethanolamine (Nice chemicals, India).

Methods

Determination of melting point: In capillary tube, small amount of drug was added, and tube is sealed. The sealed tube was located in the melting point apparatus. The heat in the apparatus was slowly increased and the temperature at which whole drug gets melted was noted. DSC study of pure drug was conceded out on Shimadzu thermal analyzer DSC TA 60. The apparatus was calibrated using standard metal like high purity indium metal. The scans were conducted at heating rate of 10°C/min in nitrogen environment.

Solubility studies: The solubility study of Luliconazole was performed in methanol, ethanol, chloroform, acetone, distilled water, 0.1 N HCl, phosphate buffer solution pH 6.8, 7.4, individually by keeping the drug containing test tube on vortex mixture.

Preparation of standard curve in methanol: Accurately weighed 100 mg of luliconazole and transferred into 100 ml volumetric flask, make the volume up to 100 ml using methanol. From the above solution 10 ml was pipette out and transferred into 100 ml volumetric flask. The volume is made up with methanol in order to get standard stock solution containing 100 ppm. Form the above solution; a sequence of dilution (2, 4, 6, 8, 10 ppm) was diluted with the help of methanol. All dilutions were measured using UV spectrophotometrically against blank of methanol at 220 nm for luliconazole. Absorbance of drug at different concentrations was calculated and graph was plotted.10

- **Infrared spectroscopic analysis:** The FTIR spectrums of luliconazole, carbopol and mixture of luliconazole, carbopol were recorded on IR spectrophotometer. All the samples are free from moisture. Infrared spectrum was recorded in the 4000-400 cm^{-1} regions (Bruker).

COPPER NANOPARTICLES SYNTHESIS: Cu nanoparticles were synthesized using size reducing agent like tri sodium citrate. Copper sulphate and trisodium citrate employed as initial substances in the development of copper NPs. All solutions were prepared in distilled water. Make 0.001 M CuSO4 solution with distilled water, take 40 ml from this solution in beaker and heated the solution to boil. In above solution, 10mL of 1% trisodium citrate was mixed drop wise. The mixture was heated under continuous magnetic stirring for 30 minutes. The mix solution was them cooled near room temperature. The reaction was allowed to take place for 24 hr. Accurately weighed 2g luliconazole was dissolved in methanol and added to copper nanoparticles.11

PREPARATION OF THE CARBOPOL GEL: At low concentration, carbopol 940 forms very good flexible transparent gel. The gel base of 2% was prepared by scattering 2 g carbopol 940 in 86 ml warm distilled water. Accurately weighed 0.6 g propyl paraben and dissolved in ethanol. Accurately weighed 0.3 g methyl paraben and dissolved in 15 ml of propylene glycol. Stirred the mixture unless gelling occurred and then mixture was neutralized with the help of 50% (w/w) triethanolamine. Triethanolamine was added drop by drop to maintain the pH between 6-7.12

The nanoparticle formulation containing drug was slowly added in carbopol 940 gel base and mixed with the help of stirrer for 5 min continue stirring.

EVALUATION OF NANOPARTICLES

- **Drug entrapment efficiency**: Take 5 ml formulation and diluted the formulation up to 8 ml with distilled water and centrifuged the diluted formulation at 15,000 rpm at 4°C for 45 min using a cooling centrifuge. The sediment and supernatant were restored after centrifugation, their volume was calculated. Then sediment was break down through n-propanol and filtered using a 0.45 µm nylon filter. The concentration of luliconazole in the sediment and supernatant was examined by UV- spectrophotometer at 220 nm. The % entrapment efficiency was estimated.

$$\text{Percentage Entrapment efficiency} = \frac{\text{Amount of entrapped drug recovered}}{\text{Total amount of drug}}$$

- **Nanoparticle shape**: Transmission electron microscopy (Philips Technai electron microscope, Netherlands) was used for the forecast of nanoparticle. At room temperature, sample was dried and vesicular were forecast under microscopy working at an acceleration voltage of 200 KV for 5 min.
- **Particle size estimation:** Dynamic light scattering method was used for the determination of copper nanoparticles, using a computerized inspection system (Malvern Zetasizer Nano-ZS, Malvern, U.K.). For the measurement of size, copper nanoparticle solution was attenuated with distilled water and implement in cuvettes of zetasizer.13
- **Zeta potential measurement:** Physical property like zeta potential which describe the net surface charge of copper nanoparticles. The stability criteria of CuNPs are measured when the zeta potential values ranges

from higher than +30 mV to lower than 30 mV.[14]

- **X-ray diffraction:** 1 ml of the copper nanoparticle solution was extend on a glass slide and dried at 40°C in an oven. The Phillips Xpert a glass slide and dried at 40°C in an oven. The Phillips Xpert Pro Diffractometer were recorded the spectra running at 40 kV and 30 mA.15
- **Copper content determination:** Determination the copper (II) ions, take 200 ml of tap water in the beaker. Water is evaporated up to 50 ml Solution is transferred into a volumetric flask and the determination is performed. To different volumes of water, the solution containing copper are added and the solution is was brought up to the mark by mixture of acetate buffer. The absorbance is measured at a wavelength of 520 nm.16

Physical evaluation of nanoparticle gel pH measurement of the nanoparticle gel: 1 gm luliconazole coupled nanoparticle gel base was mixed in 100 ml beaker containing distilled water. After that pH electrode was deep in beaker and readings were reported from digital pH meter.

- **Viscosity study**: The viscosity of copper nanoparticle was measured in Brookfield instrumentation by selecting appropriate spindle and rpm. In 50 ml beaker, 50 g of formulation was added which was set till spindle channel was drenched and set rpm. Reading pointed out over three minutes.
- **Spreadability:** Spreadability term denote a area is required to which gel willingly fall on appliance to skin or affected part.17,18 It was calculated through formulation:

$$S = M \times \frac{L}{T}$$

Where T: time taken to separate the slides L: length of slides
M: wt. tied to upper slide

- **Extrudability study:** The extrudability of luliconazole coupled nanoparticle gel was considered by stuffing nanoparticle gel in the foldable tubes. Determination in words of weight in grams, 10 sec required to extrude a 0.5 cm ribbon of gel.
- **Percentage yield:** Percentage yield was calculated by the formula.

$$\text{Percentage yield} = \frac{\text{Practicle yield}}{\text{Theortical yield}} X\ 100$$

- **Grittiness and homogeneity:** A tiny amount of nanoparticle gel was squeezed in the middle of index finger and the thumb. Uniformity of the nanoparticle gel is observed, any crude particles visible on fingers.[19,20]
- ***In vitro* release studies:** Vertical Franz diffusion cell apparatus was employed for the in vitro absorption studies. It contain donor as well as receptor chamber that is filled with PBS. The donor chamber is filled and the permeation of solute through the membrane is monitored at different interval of time. Episodic sampling from the receptor chamber was collected and measured. The jacketed cell personified is stirred during experiment at 500 rpm using a magnetic agitator.21

Drug release kinetics: The kinetic of drug release was calculated by various kinetic models as zero order release kinetics plot, first order release kinetics plot, korsmeyer-peppas release kinetics plot and higuchi release kinetics plot. To study profile release kinetics of the copper nanoparticle figures obtained from in-vitro release profile were plotted for various kinetic models. The finest fit model was set by the value of R2 close to 1.22

Antimicrobial activity studies: Antimicrobial activity has been assayed against bacteria by using agar diffusion method. The antibiotics action of drug is as oleic of its capability to growth inhibition of bacto nutrient agar or broth. Cup-plate method shall be used for the consideration of bacterial inhibition. In experiment, discs of average diameter were prepared in the bacto agar nutrient medium, containing standard bacterial inoculums. The test samples are injected in the disc and the diameter of the zone of inhibition was evaluated. All the test samples were evaluated for antibiotics activity against Staphylococcus aureus (gram positive).23

REFERENCES

1. Verma A, Singh S. Topical Gels as Drug Delivery Systems: A Review, Int J Pharm Sci Rev Res. 2013;23(2):374-82
2. Kaur J. Recent Advances in Topical Drug Delivery System. Indo American Journal of Pharmaceutical Research. 2016;6(07).
3. Mahmoodi S, Elmi A. Copper Nanoparticles as Antibacterial Agents,J Mol Pharm Org Process Res. 2018;6:1.
4. Dizaj S, Lotfipour F. Antimicrobial activity of the metals and metal oxide nanoparticles. Materials Science and Engineering C 44 (2014): 278–284
5. Santo C, Taudte N. Contribution of Copper Ion Resistance to Survival of *Escherichia coli* on Metallic Copper Surfaces. Applied And Environmental Microbiology. 2008;74(4):977-86.
6. Grass G, Rensing C. Metallic Copper as an Antimicrobial Surface. Applied And Environmental Microbiology. Mar. 2011, p. 1541–1547, Vol. 77, No. 5
7. Santo C, Lam E. Bacterial Killing by Dry Metallic Copper Surfaces, Applied And Environmental Microbiology, Feb. 2011, p. 794–802, Vol. 77, No. 3.
8. Mehtar S, Wiid I. The antimicrobial activity of copper and copper alloys against nosocomial pathogens and Mycobacterium tuberculosis isolated from healthcare facilities in the Western Cape: an *in-vitro* study. Journal of Hospital Infection (2008) 68, 45e51
9. Casey A, Adams D. Role of copper in reducing hospital environment Contamination, Journal of Hospital Infection (2010) 74, 72e77
10. Anuradha Y, Venkateswara Rao P. Method Development And Validation For Simultaneous Estimation Of Luliconazole And Metronidazole In Combined Dosage Form By Rp-Hplc, World Journal of Pharmacy and Pharmaceutical Sciences, Vol 4, Issue 07, 2015
11. Saravanam B, Mannivanam V. Synthesis of copper nanoparticle using trisodium citrate and evaluation of antibacterial activity, Life Science Informatics Publications, 4(5) Page No.842, 2018 Sept – Oct
12. Devi M, Senthil Kumar M, Mahadevan N. Amphotericin-B loaded vesicular systems for the treatment of topical fungal infection, International Journal of Recent Advances in Pharmaceutical Research, 2011; (4): 37-46.
13. Sivashankar N, Karthick D. Particle Characterization of Copper Nanoparticles by Electrochemical Method, IJSTE - International Journal of Science Technology and Engineering, Volume 3 ,Issue 01, July 2016
14. Mohammed J. Haider*, Mohammed S. Mehdi. Study of morphology and Zeta Potential analyzer for the Silver Nanoparticles International Journal of Scientific and Engineering Research, Volume 5, 2014; 63-69
15. Usha S, Ramappa KT, Hiregoudar S, Vasanthkumar G and Aswatha narayana D.S. Biosynthesis and Characterization of Copper Nanoparticles from Tulasi (Ocimum sanctum L.) Leaves. Int.J.Curr.Microbiol.App.Sci. 6(11): 2219-2228.
16. Zagurskaya-Sharaevskaya O and Povar I.Determination of Cu (II) ions using sodiumsalt of 4-phenylsemicarbazone 1,2-naphthoquinone-4-sulfonic acid in natural and industrial environments, Zagurskaya-Sharaevskaya and Povar Ecological Processes (2015) 4:16
17. Prasanthi D, Lakshmi PK. Development of ethosomes with taguchi robust design-based studies for

transdermal delivery of alfuzosin hydrochloride, Prasanthi and Lakshmi, International Current Pharmaceutical Journal 2012, 1(11): 370-375

18. Thakur V, Prashar B, Arora S. Formulation and *in vitro* Evaluation of Gel for Topical Delivery of Antifungal Agent Fluconazole Using Different Penetration Enhancers, Drug Invention Today, 2012, 4(8), 414-419

19. Niyaz Basha B, Prakasam K, Goli D. "Formulation and evaluation of Gel containing Fluconazole-Antifungal Agent", Int. J. Drug Dev. and Res., Oct-Dec 2011, 3(4): 109-128

20. Saroha1 K, Singh S. Transdermal Gels - An Alternative Vehicle For Drug Delivery, IJPCBS 2013, 3(3), 495-503

21. Barupal AK, Gupta V, Ramteke S. Preparation and Characterization of Nanoparticles for Topical delivery of Aceclofenac, Indian Journal of Pharmaceutical Sciences, 2010; 72(5): 582-86.

22. Saiesh P, Shabaraya AR, Shripathy D, Leyana Soman, Kinetic Modeling of Drug Release from Topical Gel, International Journal of Universal Pharmacy and Bio Sciences, 2014; 3(3): 92-104.

23. Ghatage SL, Navale SS, Mujawar NK, Patil S, Patil V. Antimicrobial screening. Indian Journal of Drugs. 2014, 2(3), 84-8.

24. Hamelian, M., Zangeneh, M.M., Amisama, A., Varmira, K., Veisi, H. Green synthesis of silver nanoparticles using *Thymus kotschyanus* extract and evaluation of their antioxidant, antibacterial and cytotoxic effects. Appl Organometal Chem. 2018; 32: e4458.

25. Seydi, N., Mahdavi, B., Paydarfard, S., Zangeneh, A., Zangeneh, M.M., Najafi, F., Jalalvand, A.R., Pirabbasi, E. Preparation, characterization, and assessment of cytotoxicity, antioxidant, antibacterial, antifungal, and cutaneous wound healing properties of titanium nanoparticles using aqueous extract of Ziziphora clinopodioides Lam leaves. Appl Organometal Chem. 2019; 33:e5009.

26. Zangeneh, M.M., Joshani, Z., Zangeneh, A., Miri, E., Green synthesis of silver nanoparticles using aqueous extract of Stachys lavandulifolia flower, and their cytotoxicity, antioxidant, antibacterial and cutaneous wound-healing properties. Appl Organometal Chem. 2019; 33: e5016.

27. Hemmati S., Rashtiani A., Zangeneh P.M.A., Veisi, H. Green synthesis and characterization of silver nanoparticles using Fritillaria flower extract and their antibacterial activity against some human pathogens. Polyhedron. 2019; 158; 8-14.

28. Zangeneh, M.M. Green synthesis and chemical characterization of silver nanoparticles from aqueous extract of *Falcaria vulgaris* leaves and assessment of their cytotoxicity and antioxidant, antibacterial, antifungal and cutaneous wound healing properties. Appl Organometal Chem. 2019;33:e4963.

29. Mahdavi, B., Saneei, S., Qorbani, M., Zhaleh, M., Zangeneh, A., Zangeneh, M.M., Pirabbasi, E., Abbasi, N. *Ziziphora clinopodioides* Lam leaves aqueous extract mediated synthesis of zinc nanoparticles and their antibacterial, antifungal, cytotoxicity, antioxidant, and cutaneous wound healing properties under *in vitro* and *in vivo* conditions. Appl Organometal Chem. 2019;e5164.

30. Seydi, N., Saneei, S., Jalalvand, A.R., Zangeneh, M.M., Zangeneh, A., Tahvilian, R. Pirabbasi, E. Synthesis of titanium nanoparticles using *Allium eriophyllum* Boiss aqueous extract by green synthesis method and evaluation of their remedial properties. Appl Organometal Chem. 2019; e5191.