



# GREEN SYNTHESIS OF SILVER NANOPARTICLES BY USING AZADIRACHTA INDICA AQUEOUS LEAF EXTRACT

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**ABSTRACT;** In this study, we employed a fast and straightforward approach to synthesize silver nanoparticles using an aqueous leaf extract of Azadirachta indica. The plant extract serves as both a reducing agent and a capping agent.

To identify the specific compounds responsible for the reduction of silver ions, we conducted an investigation of the functional groups present in the plant extract using FTIR analysis. Several techniques were employed to characterize the synthesized nanoparticles, including DLS, Photoluminescence, TEM, and UV-Visible spectrophotometry. The UV-visible spectrophotometer revealed an absorbance peak within the range of 436-446 nm.

The silver nanoparticles exhibited antibacterial properties against both gram-positive microorganism Staphylococcus aureus and gram-negative microorganism Escherichia coli. Additionally, we evaluated the photoluminescence properties of the synthesized silver nanoparticles.

The results confirmed that this protocol is a simple, rapid, one-step, eco-friendly, and non-toxic alternative to conventional physical/chemical methods. Remarkably, the conversion of ions into silver nanoparticles was accomplished within only 15 minutes at room temperature, without the use of any hazardous chemicals.

**Keywords:** Azadirachta indica, silver nanoparticles, FTIR, TEM and UV-Visible spectrophotometry,

## 1 INTRODUCTION

The 'green' environment friendly processes in chemistry and chemical technologies are becoming increasingly popular and are much needed as a result of worldwide problems associated with environmental concerns.

Silver is the one of the most commercialised nano-material with five hundred tons of silver nanoparticles production per year and is estimated to increase in next few years.

Including its profound role in field of high sensitivity bio-molecular detection, catalysis, biosensors and medicine; it is been acknowledged to have strong inhibitory and bactericidal effects along with the anti-fungal, anti-inflammatory and anti-angiogenesis activities.

A number of techniques are available for the syntheses of silver nanoparticles like ion sputtering, chemical reduction, sol gel, etc.

Unfortunately many of the nanoparticle syntheses methods involve the use of hazardous chemicals or high energy requirements, which are rather difficult and including wasteful

purifications. Thus; scenario is that whatever the method followed, will always leading to the chemical contaminations during their syntheses procedures or in later applications with associated limitations. Yet; one cannot deny their ever growing applications in daily life. For instances; "The Noble Silver Nanoparticles" are striving towards the edge-level utilities in every aspect of science and technology including the medical fields; thus cannot be neglected just because of their source of generation.

Hence, it is becoming a responsibility to emphasise on an alternate as the synthetic route which is not only cost effective but should be environment friendly in parallel. Keeping in view of the aesthetic sense, the green syntheses are rendering themselves as key procedure and proving their potential at the top.

The techniques for obtaining nanoparticles using naturally occurring reagents such as sugars, biodegradable polymers (chitosan, etc.), plant extracts, and microorganisms as reductants and capping agents could be considered attractive for nanotechnology

Greener syntheses of nanoparticles also provides advancement over other methods as they are simple, one step, cost-effective, environment friendly and relatively reproducible and often results in more stable materials

Microorganisms can also be utilized to produce nanoparticles but the rate of syntheses are slow compared to routes involving plants mediated synthesis. Although, the potential of higher plants as source for this purpose is still largely unexplored. Very recently plant extract of marigold flower, *Abutilon indicum*, *Solanum tuberosum*, *Erythrina indica*, beet root, mangosteen, *Ocimum tenuiflorum*, *Spirogyra varians*, *Melia dubia*, leaf extract of *Acalypha indica* with high anti-bacterial activities and of *Sesuvium portulacastrum* also reported in literature with nanoparticle size ranging from 5 to 20 nm are brimming in literature as a source for the synthesis of silver nanoparticles as an alternative to the conventional methods.

Considering the vast potentiality of plants as sources this work aims to apply a biological green technique for the synthesis of silver nanoparticles as an alternative to conventional methods.

In this regard, leaf extract of *Azadirachta indica* (commonly known as neem) a species of family Meliaceae was used for bioconversion of silver ions to nanoparticles. This plant is commonly available in India and each part of this tree has been used as a household remedy against various human ailments from antiquity and for treatment against viral, bacterial and fungal infections.

Silver nanoparticles can be produced at low concentration of leaf extract without using any additional harmful chemical/physical methods.

The effect of concentration of metal ions and concentration of leaf extract quantity were also evaluated to optimize route to synthesize silver nanoparticle. The method applied here is simple, cost effective, easy to perform and sustainable.

## 2 EXPERIMENTAL

Typically, a plant extract-mediated bioreduction involves mixing the aqueous extract with an aqueous solution of the appropriate metal salt. The synthesis of nanoparticle occurs at room temperature and completes within a few minutes.

### • Preparation of plant extract

*A. indica* leaf extract was used to prepare silver nanoparticles on the basis of cost effectiveness, ease of availability and its medicinal property. Fresh leaves were collected from university campus in month of February. They were surface cleaned with running tapwater to remove debris and other contaminated organic contents, followed by double

distilled water and air dried at room temperature. About 20 gm of finely cut leaves were kept in a beaker containing 200 mL double distilled water and boiled for 30 min. The extract was cooled down and filtered with Whatman filter paper no.1 and extract was stored at 4 °C for further use.

- **Green synthesis of silver nanoparticles**

Silver nitrate GR used as such (purchased from Merck, India). 100 mL, 1 mM solution of silver nitrate was prepared in an Erlenmeyer flask. Then 1, 2, 3, 4 and 5 mL of plant extract was added separately to 10 mL of silver nitrate solution keeping its concentration at 1 mM. Silver nanoparticles were also synthesized by varying concentration of AgNO<sub>3</sub> (1 mM to 5 mM) keeping extract concentration constant (1 mL). This setup was incubated in a dark chamber to minimize photo-activation of silver nitrate at room temperature. Reduction of Ag<sup>+</sup> to Ag<sup>0</sup> was confirmed by the colour change of solution from colour-less to brown. Its formation was also confirmed by using UV-Visible spectroscopy.

- **Characterization of synthesised silver nanoparticles**

UV-Vis spectral analysis was done by using Shimadzu UV-Visible spectrophotometer (UV-1800, Japan). UV-Visible absorption spectrophotometer with a resolution of 1 nm between 200 and 800 nm was used. One millilitre of the sample was pipetted into a test tube and subsequently analysed at room temperature. Dynamic light scattering (Spectroscatter 201) was used to determine the average size of synthesized silver nanoparticles. FTIR spectra were recorded on Perkin Elmer 1750 FTIR Spectrophotometer. The particle size and surface morphology was analysed using Transmission electron microscopy (TEM), operated at an accelerated voltage of 120 kV. Photoluminescence studies were evaluated by using eclipse Fluorescence spectrophotometer (Agilent technologies).

- **Fixation of different parameters**

The reaction was monitored at different time intervals. The reaction was monitored using different concentration of silver nitrate (1 mM, 2 mM, 3 mM, 4 mM and 5 mM) and also by varying leaf extract solution (1e5 mL) and their absorbance was measured.

- **Assessment of antimicrobial assay**

The antibacterial assays were done on human pathogenic *Escherichia coli* and *Staphylococcus aureus* by using standard disc diffusion method. MacConkey broth (HiMedia) medium was used to sub culture bacteria and were incubated at 37 °C for 24 h. Fresh overnight cultures were taken and spread on the MacConkey agar plates to cultivate bacteria. Sterile paper discs of 5 mm diameter saturated with plant extract, silver nanoparticle and double distilled water (as control) were placed in each plate and incubated again at 37 °C for 24 h and the antibacterial activity was measured based on the inhibition zone around the disc impregnated with plant extract and synthesized silver nanoparticle.

### 3 RESULTS AND DISCUSSION

- **Visual observation and UV-Vis spectroscopy**

In all experiments, addition of plant extract of *A. indica* into the beakers containing aqueous solution of silver nitrate led to the change in the colour of the solution to yellowish to reddish-brown (shown in Fig. 1) within reaction duration due to excitation of surface plasmon

vibrations in silver nanoparticles (Veerasamy et al., 2011). On addition of different concentration (1e5 mL) of leaf extracts to aqueous silver nitrate solution keeping its concentration 10 mL (1 mM) constant, the colour of the solution changed from faint light to yellowish brown and finally to colloidal brown indicating formation of silver nanoparticles. Different parameters were optimized including concentration of silver nitrate and *A. indica* leaf extract, and time which had been identified as factors affecting the yields of silver nanoparticles.

Silver nanoparticles were synthesized at different concentrations of leaf extract such as 1e5 mL using 1 mM of silver nitrate were analysed by UV spectra of Plasmon resonance band observed at 436-446 nm similar to those reported in literature.



**Fig. 1 e Digital optical images of synthesized silver nanoparticles with different conc (1e5 mM) of AgNO<sub>3</sub>.**

If we increase the leaf extract concentration to 4 mL, there is increase in wavelength up to 448 nm as pre-sented in Fig. 2a. The slight variations in the values of absor-bance signifiesthat the changes are the particle size.

On increasing concentration of extract there is increase in in-tensity of absorption. The UVVisible spectra recorded after different time intervals of 1 h, 2 h, 3 h, 4 h, 18 h and 24 h from the initiation of reaction with varying amount of plant extract Fig 3ae. It is generally recognize that UVeVis spectroscopy could be used to examine size and shape-controlled nano-particles in aqueous suspensions.

Parallel changes in colour have been observed when different concentrations (1 mM to 5 mM) of silver nitrate was used by keeping plant extract (1 mL) constant. The appearance of the brown colour was due to the excitation of the Surface Plasmon Resonance (SPR), typical of silver nanoparticles having absorbance values which were reported earlier in the visible range of 446-448 nm.

There is increase in intensity of absorption peaks after regular in- tervals of time and the colour intensity increased with the duration of incubation. It was also observed from Fig. 2b

that the intensity of absorption peaks increases with increase in the concentration of the silver nitrate salt. All the results are very close already reported in literature showing absorbance at 445 nm of silver nanoparticles synthesized by *Cochlospermum religiosum* extract and by

*Pithecolobium dogonia* extract .

The UVVis spectra recorded, implied that most rapid bioreduction was achieved using *A. indica* spectra and visual observation revealed that formation of silver nanoparticles occurred rapidly within 15 min.

#### • Particle size and distribution

The size distribution histogram of dynamic light scattering (DLS) indicates that the size of these silver nanoparticles is 34 nm. Some distribution at lower range of particle size indicates that the synthesized particles are also in lower range of particle size. Fig. 4 shows the DLS pattern of the suspension of Ag nanoparticles synthesized using *A. Indica* aqueous leaf extract.

#### • FTIR analysis

The dual role of the plant extract as a reducing and capping agent and presence of some functional groups was confirmed by FTIR analysis of silver nanoparticle. A broad band between 3454  $\text{cm}^{-1}$  is due to the N-H stretching vibration of group  $\text{NH}_2$  and OH the overlapping of the stretching vibration of attributed for water and *A. indica* leaf extract molecules. The band at 1636  $\text{cm}^{-1}$  corresponds to amide C=O stretching and a peak at 2083  $\text{cm}^{-1}$  can be assigned to alkyne group present in phyto-constituents of extract Fig. 5. The observed peaks at 1113  $\text{cm}^{-1}$  denote eCeOC- linkages, or eCeO- bonds. The observed peaks are mainly attributed to flavanoids and terpenoids excessively present in plants extract. On the other hand, the extract sample prepared shows a wide and strong peak with maximum intensity at 553  $\text{cm}^{-1}$ . The results are in good agreement with those found in literature From FTIR results, it can be concluded that some of the bioorganics compounds from *A. indica* extract formed a strong coating/capping on the nanoparticles.

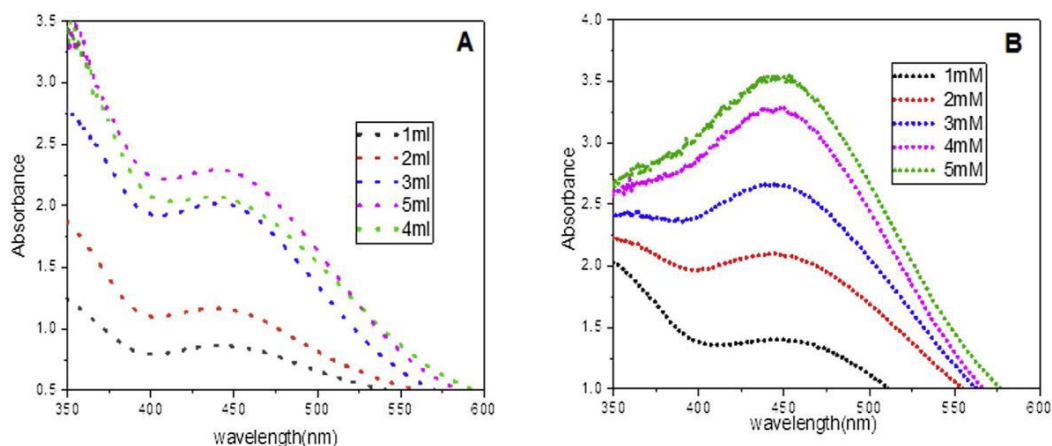


Fig.2 UV-Vis- spectra showing absorbance with different conc. of (a) plant extract (1e5mL) and (b) AgNO3 (1e5 mM)



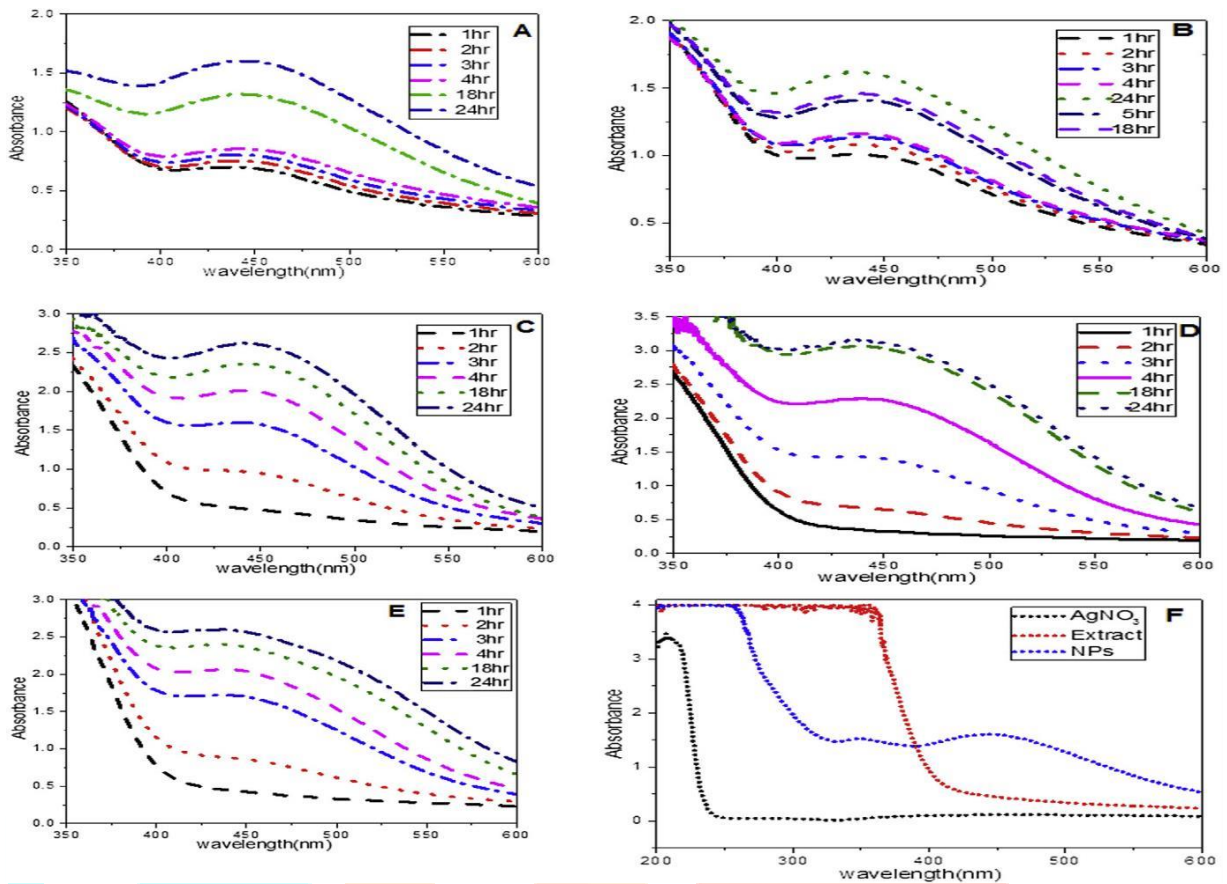


Fig. 3 e UV-Vis spectra showing absorbance with (different time intervals) with conc. of (a) 1 mL extract (b) 2 mL extract (c) 3 mL extract (d) 4 mL extract (e) 5 mL extract and (f) extract, AgNO<sub>3</sub> solution and silver nanoparticle.

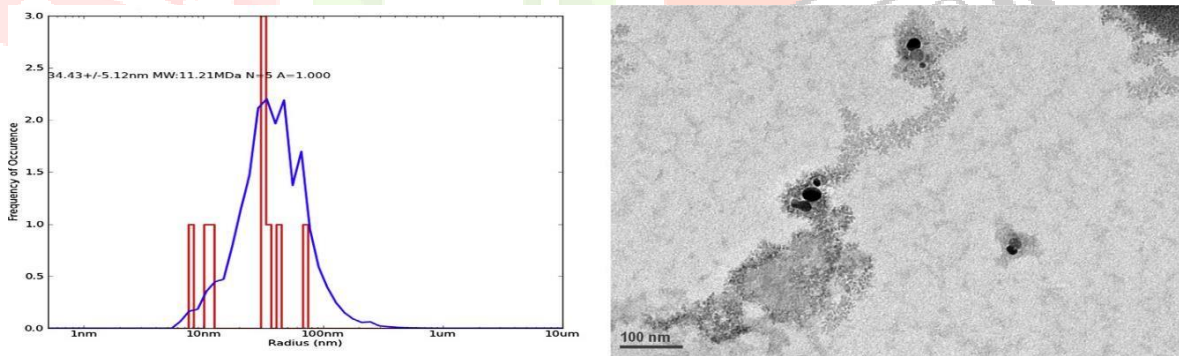


Fig. 4 e DLS histogram and TEM image of synthesised silver nanoparticles.

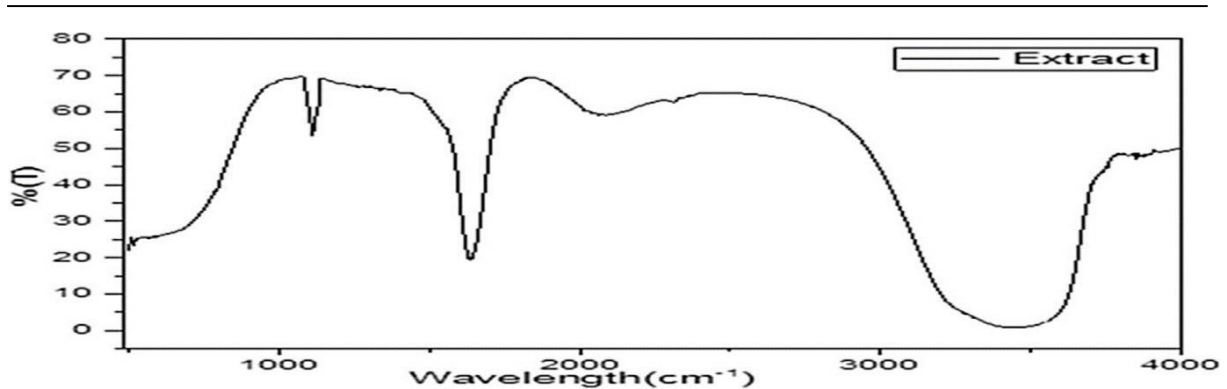


Fig. 5 e FTIR spectra of plant extract and synthesised silver nanoparticle.

## TEM analysis

Transmission electron microscopy (TEM) has been used to identify the size, shape and morphology of nanoparticles. It reveals that the silver nanoparticles are well dispersed and predominantly spherical in shape, while some of the NPs were found to be having structures of irregular shape as shown in Fig. 4. The nanoparticles are homogeneous and spherical which conforms to the shape of SPR band in the UV-visible spectrum. The particle size agrees with that calculated from DLS histogram with average diameter of around 34 nm.

## Antimicrobial activity

Silver nanoparticles, due to their antimicrobial properties have been used most widely in the health industry, medicine, textile coatings, food storage, dye reduction, wound dressing, antiseptic creams and a number of environmental applications. Since ancient times,

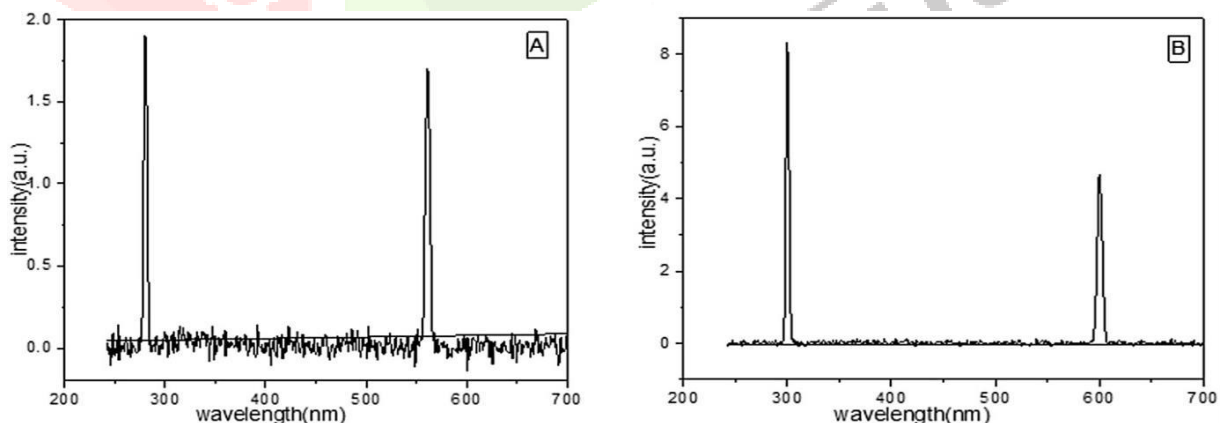
elemental silver and its compounds have been used as antimicrobial agents; and was used to preserve water in form of silver coins/silver vessels. We have examined *A. indica* extract

mediated silver nanoparticles as possible antibacterial agents. The plant extract and those

mediated silver nanoparticles were immediately tested for respective antimicrobial activities towards both gram positive (*S. aureus*) and gram negative (*E. coli*) bacterial strains showing the zones of inhibition. Based on the zone of inhibition produced, synthesized silver nanoparticles prove to exhibit good antibacterial activity against *E. coli* and *S. aureus*. On the other hand, control and plant extract alone did not exhibit any antibacterial activity. Although, it is to be presumed that the leaves extract of the plant used possess the antibacterial activities and must be reflected through greater inhibition zone but it alone shows very low activity due to its medium of extraction as well as lower concentration during experimentation.

**Table 1 – Zone of inhibition (mm) obtained by disc diffusion method.**

Components	Zone of inhibition (mm)	
	<i>E. coli</i>	<i>S. aureus</i>
Control	NZ	NZ
Plant extract	NZ	NZ
Silver nanoparticle	9	9



**Fig. 6 e Fluorescence spectra of silver nanoparticles formed with excitation at (a) 280nm and (b) 300 nm.**

The silver nanoparticles showed efficient antimicrobial property compared to other due to their extremely large surface area providing better contact with cell wall of microorganisms.

## Photoluminescence study

Silver nanoparticles are reported to exhibit visible photo-luminescence and their fluorescence spectra are shown in Fig. 6. The silver nanoparticles were found to be luminescent with two emissions at 280 and 561 nm for an excitation at 280 nm. When nanoparticles were excited at 300 nm, it showed two excitation at 300 and 600 nm, the excitation at 300 nm is of high intensity in comparison to other one. The luminescence at 280 and 300 nm may be due to presence of biochemical or antioxidants present in plant extract. The nanoparticles synthesised using olive leaf extract are also reported to be luminescent with emission band at 425 nm.

## 4 CONCLUSION

A simple one-pot green synthesis of stable silver nanoparticles using *A. indica* leaf extract at room temperature was reported in this study. Synthesis was found to be efficient in terms of reaction time as well as stability of the synthesized nanoparticles which exclude external stabilizers/reducing agents. It proves to be an eco-friendly, rapid green approach for the synthesis providing a cost effective and an efficient way for the synthesis of silver nanoparticles. Therefore, this reaction pathway satisfies all the conditions of a 100% green chemical process. The synthesized silver nanoparticles showed efficient antimicrobial activities against both *E. coli* and *S. aureus*. Benefits of using plant extract for synthesis is that it is energy efficient, cost effective, protecting human health and environment leading to lesser waste and safer products. This eco-friendly method could be a competitive alternative to the conventional physical/chemical methods used for synthesis of silver nanoparticle and thus has a potential to use in biomedical applications and will play an important role in opto-electronics and medical devices in near future.

## 5 REFERENCES

- 1) Ahmed, S., Ahmad, M., & Ikram, S. (2014). Chitosan: a natural antimicrobial agent a review. *Journal of Applicable Chemistry*, 3(2), 493e503.
- 2) Ahmed, S., Ahmad, M., Swami, B. L., & Ikram, S. (2015). Plants extract mediated synthesis of silver nanoparticles for antimicrobial applications: a green expertise. *Journal of Advance Research*. <http://dx.doi.org/10.1016/j.jare.2015.02.007>. Ahmed, S., & Ikram, S. (2015). Chitosan & its derivatives: a review in recent innovations. *International Journal of Pharmaceutical Sciences and Research*, 6(1), 14e30.
- 3) Ashokkumar, S., Ravi, S., & Velmurugan, S. (2013). Green synthesis of silver nanoparticles from *Gloriosa superba* L. leaf extract and their catalytic activity. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 115, 388e392.
- 4) Banerjee, P., Satapathy, M., Mukhopadhyay, A., & Das, P. (2014). Leaf extract mediated green synthesis of silver nanoparticles from widely available Indian plants: synthesis, characterization, antimicrobial property and toxicity analysis. *Bioresources and Bioprocessing*, 1(3), 1e10.
- 5) Bindhu, M. R., & Umadevi, M. (2015). Antibacterial and catalytic activities of green synthesized silver nanoparticles. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 135, 373e378.
- 6) Devi, L. S., & Joshi, S. R. (2015). Ultrastructures of silver nanoparticles biosynthesized using endophytic fungi. *Journal of Microscopy and Ultrastructure*, 3, 29e37.
- 7) El-Chaghaby, G. A., & Ahmad, A. F. (2011). Biosynthesis of silver nanoparticles using pistacia lentiscus leaves extract and investigation of their antimicrobial effect. *Oriental Journal of Chemistry*, 27, 929e936
- 8) Gao, X., Yourick, J. J., Topping, V. D., Black, T., Olejnik, N., Keltner, Z., et al. (2014). Toxicogenomic study in rat thymus of F1 generation offspring following maternal exposure to silver ion. *Toxicology Reports*. <http://dx.doi.org/10.1016/j.toxrep.2014.12.008>.
- 9) Ibrahim, H. M. M. (2015). Green synthesis and characterization of silver nanoparticles using banana peel extract and their antimicrobial activity against representative microorganisms. *Journal of Radiation Research and Applied Sciences*. <http://dx.doi.org/10.1016/j.jrras.2015.01.007>.
- 10) Khalil, M. M. H. (2013). Green synthesis of silver nanoparticles using olive leaf extract and its antibacterial activity. *Arabian Journal of Chemistry*, 1131e1139.
- 11) Khalil, M. M. H., Ismail, E. H., & El-Magdoub, F. (2012). Biosynthesis of Au nanoparticles using olive leaf extract. 1<sup>st</sup> nano updates. *Arabian Journal of Chemistry*, 5, 431e437.

- 12) Kharissova, O. V., Dias, H. V. R., Kharisov, B. I., Perez, B. O., & Perez, V. M. J. (2013). The greener synthesis of nanoparticles. *Trends in Biotechnology*, 31, 240e248.
- 13) Krishnaraj, C., Jagan, E. G., Rajasekar, S., Selvakumar, P., Kalaichelvan, P. T., & Mohan, N. (2010). Synthesis of silver nanoparticles using *Acalypha indica* leaf extracts and its antibacterial activity against water borne pathogens. *Colloids and Surfaces B: Biointerfaces*, 76, 50e56.
- 14) Larue, C., Castillo-Michel, H., Sobanska, S., Cecillon, L., Bureau, S., Barthès, V., et al. (2014). Foliar exposure of the crop *Lactuca sativa* to silver nanoparticles: evidence for internalization and changes in Ag speciation. *Journal of Hazardous Materials*, 264, 98e106.
- 15) Logeswari, P., Silambarasan, S., & Abraham, J. (2012). Synthesis of silver nanoparticles using plants extract and analysis of their antimicrobial property. *Journal of Saudi Chemical Society*, 19(3), 311e317.
- 16) Logeswari, P., Silambarasan, S., & Abraham, J. (2013). Ecofriendly synthesis of silver nanoparticles from commercially available plant powders and their antibacterial properties. *Scientia Iranica*, 20, 1049e1054.
- 17) Mahdi, S., Taghdiri, M., Makari, V., & Rahimi-Nasrabadi, M. (2015). Procedure optimization for green synthesis of silver nanoparticles by aqueous extract of *Eucalyptus oleosa*. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 136, 1249e1254.
- 18) Mittal, J., Batra, A., Singh, A., & Sharma, M. M. (2014). Phytofabrication of nanoparticles through plant as nanofactories. *Advances in Natural Sciences: Nanoscience and Nanotechnology*, 5. <http://dx.doi.org/10.1088/2043-6262/5/4/043002>, 043002, 043002.
- 19) Nabikhan, A., Kandasamy, K., Raj, A., & Alikunhi, N. M. (2010). Synthesis of antimicrobial silver nanoparticles by callus and leaf extracts from saltmarsh plant, *Sesuvium portulacastrum* L. *Colloids and Surfaces B: Biointerfaces*, 79, 488e493.
- 20) Obaid, A. Y., Al-Thabaiti, S. A., Al-Harbi, L. M., & Khan, Z. (2015). Extracellular bio-synthesis of silver nanoparticles. *Global Advanced Research Journal of Microbiology*, 3(8), 119e126.
- 20) Omoja, V. U., Anaga, A. O., Obidike, I. R., Ihedioha, T. E., Umeakuana, P. U., Mhomga, and diminazene diacetate in the treatment of experimental *Trypanosoma brucei brucei* infection in rats. *Asian Pacific Journal of Tropical Medicine*, 4, 337e341. L. I., et al. (2011). The effects of combination of methanolic leaf extract of *Azadirachta indica*.
- 21) Padalia, H., Moteriya, P., & Chanda, S. (2014). Green synthesis of silver nanoparticles from marigold flower and its synergistic antimicrobial potential. *Arabian Journal of Chemistry*. <http://dx.doi.org/10.1016/j.arabjc.2014.11.015>.
- 22) Prathna, T. C., Chandrasekaran, N., Raichur, A. M., & Mukherjee, A. (2011). Biomimetic synthesis of silver nanoparticles by *Citrus limon* (lemon) aqueous extract and theoretical prediction of particle size. *Colloids and Surfaces B: Biointerfaces*, 82, 152e159.
- 23) Sadeghi, B., & Gholamhoseinpoor, F. (2015). A study on the stability and green synthesis of silver nanoparticles using *Ziziphora tenuior* (Zt) extract at room temperature. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 134, 310e315.
- 24) Salari, Z., Danafar, F., Dabaghi, S., & Ataei, S. A. (2014). Sustainable synthesis of silver nanoparticles using macroalgae *Spirogyra varians* and analysis of their antibacterial activity. *Journal of Saudi Chemical Society*. <http://dx.doi.org/10.1016/j.jscs.2014.10.004>.
- 25) Sasikala, A., Linga Rao, M., Savithamma, N., & Prasad, T. N. V. K. V. (2014). Synthesis of silver nanoparticles from stem bark of *Cochlospermum religiosum* (L.) Alston: an important medicinal plant and evaluation of their antimicrobial efficacy. *Applied Nanoscience*. <http://dx.doi.org/10.1007/s13204-014-0380-8>.
- 26) Sinha, S. N., Paul, D., Halder, N., Sengupta, D., & Patra, S. K. (2014). Green synthesis of silver nanoparticles using fresh water green alga *Pithophora oedogonia* (Mont.) Wittrock and evaluation of their antibacterial activity. *Applied Nanoscience*. <http://dx.doi.org/10.1007/s13204-014-0366-6>.
- 27) Sre, P. R. R., Reka, M., Poovazhagi, R., Kumar, M. A., & Murugesan, K. (2015). Antibacterial and cytotoxic effect of biologically synthesized silver nanoparticles using aqueous root extract of *Erythrina indica* lam. *Spectrochimica Acta Part A:*



- 28) Thuesombat, P., Hannongbua, S., Akasit, S., & Chadchawan, S. (2014). Ecotoxicology and environmental safety effect of silver nanoparticles on rice (*Oryza sativa* L. cv. KDML 105) seed germination and seedling growth. *Ecotoxicology and Environmental Safety*, 104, 302e309.
- 29) Tripathy, A., Raichur, A. M., Chandrasekaran, N., Prathna, T. C., & Mukherjee, A. (2010). Process variables in biomimetic synthesis of silver nanoparticles by aqueous extract of *Azadirachta indica* (Neem) leaves. *Journal of Nanoparticle Research*, 12, 237e246.
- 30) Veerasamy, R., Xin, T. Z., Gunasagaran, S., Xiang, T. F. W., Yang, E. F. C., Jeyakumar, N., et al. (2011). Biosynthesis of silver nanoparticles using mangosteen leaf extract and evaluation of their antimicrobial activities. *Journal of Saudi Chemical Society*, 15, 113e120.

