



A REVIEW ARTICLE ON MICROSPHERE PREPARATION AND EVALUATION

Sampada G. Shrawankar*, Dr. Manjeet Singh, Dr. Rajesh Z. Mujariya, Lokesh I. Patle.

Student, Executive Director, Principal, HOD Pharmaceutics.

Department of Pharmaceutics,
Sardar Patel University, Balaghat

Abstract: The goal of targeted drug delivery is to concentrate the medication in the tissues of interest while reducing the relative concentration of the medication in the other tissues. As a consequence, the drug is directed to the desired location. As a result, the drug has no impact on the tissues around it. As a result, carrier technology offers an intelligent form of drug administration by attaching the drug to a carrier particle such as microspheres, nanoparticles, liposomes, niosomes, and so on, which modulates the drug's release and absorption qualities. Microspheres are free-flowing powders comprised of biodegradable proteins or synthetic polymers with particle sizes smaller than 200 m. It is the most dependable means of delivering medication to the intended recipient. With an emphasis on preparation, application, biocompatibility, and stability, it becomes necessary to deliver that specific agent to the target tissues in optimal amount at the right time with minimal side effects in order to achieve the required or enhanced therapeutic efficacy of a given drug effect. The controlled drug delivery system is concerned with the systemic release of a pharmacological agent in order to maintain a therapeutic level of medication in the body for an extended length of time. This can be accomplished by integrating the therapeutic ingredient into biodegradable polymers and constantly releasing the substance as the matrix erodes.

Key Words: *Microsphere, Types of microspheres, Methods of preparation, characterization of microspheres, applications.*

INTRODUCTION: Microspheres are characterized as free-flowing powders made of protein or synthetic polymers that are biodegradable in nature and have diameters ranging from 1 m to 1000 m. These are free-flowing spherical particles made of proteins or synthetic polymers. They are biodegradable in nature. Microspheres are classified into two types: microcapsules and micrometrics. Microcapsules are those in which the entrapped material is surrounded by a distinct capsule wall and are also known as micro particles. Microspheres may be made from a variety of natural and synthetic materials. Microspheres serve a significant function in increasing the bioavailability of conventional medications while reducing negative effects.

IDEAL CHARACTERISTICS OF MICROSPHERES

- The capacity to include moderately high quantities of the medication.
- After-synthesis stability with a therapeutically acceptable shelf life. Controlled particle size and dispersibility in aqueous injection vehicles.
- Active reagent release with good control over a large time scale. Biocompatibility with regulated biodegradability.
- Chemical modification susceptibility.

ADVANTAGES OF MICROSPHERES

- Decrease the frequency of dose and thereby enhance patient compliance. Improved medication use will increase bioavailability and lessen the occurrence or severity of side effects.
- Microsphere shape enables regulated variations in drug breakdown and release. Turn liquid to solid and conceal harsh flavour. Protects the GIT from the drug's irritating effects.
- Biodegradable microspheres offer the benefit of not requiring surgical procedures for insertion and removal over big polymer implants.

- Particle size reduction to improve the solubility of poorly soluble drugs. Consistent and prolonged therapeutic effect.
- Increase patent compliance by providing constant drug concentration in blood.
- Reduce dosage and toxicity. Shield the medication against enzymatic and photolytic cleavage, making it ideal for protein drug delivery.

TYPES OF MICROSPHERES

- Bioadhesive microspheres
 - Magnetic microspheres
 - Floating microspheres
 - Radioactive microspheres
 - Polymeric microspheres
- i) Biodegradable polymeric microsphere ii) Synthetic polymeric microsphere

METHOD OF PREPARATION

➤ **Spray Drying-** First, the polymer is dissolved in a suitably volatile organic solvent, such as Acetone, dichloromethane, and so forth. The solid medication is then distributed in the polymer solution using high-speed homogenization. This dispersion is then atomized in a hot air stream. Atomization produces microscopic droplets or fine mists from which the solvent evaporates instantly, resulting in the creation of microspheres with sizes ranging from 1 to 100m. The cyclone separator separates micro particles from hot air, while vacuum drying removes any trace of solvent. One of the most significant advantages of the method is its ability to operate in aseptic conditions. The process is fast, which results in the creation of porous micro particles. A liquid manufacturing vehicle is used to carry out the procedures. The microcapsule coating is disseminated in a volatile solvent that is incompatible with the liquid phase of the production vehicle. In the coating polymer solution, a core material to be microencapsulated is dissolved or distributed. To acquire the proper size microcapsule, the core material combination is distributed in the liquid production vehicle phase by agitation. If required, the mixture is heated to evaporate the solvent so that the polymer of the core material can be dispersed in the polymer. Polymer shrinks around the core as a result of the solution. Matrix-type microcapsules are formed when the core material is dissolved in the coated polymer solution are created.

➤ **Single emulsion techniques-** This approach is used to create a variety of proteins and carbs. In which natural polymers are dissolved in an aqueous solution and then Dispersion in the oil phase, i.e. in a non-aqueous medium. It is the initial step. The next stage is cross linking, which is done in two ways.

(1) **Cross linking by heat:** this is done by placing the dispersion in hot oil, but it is not suited for Thermolabile medicines.
 (2) **Cross-linking chemicals:** - by employing substances such as formaldehyde, di acid chloride, glutaraldehyde, and others, but with the drawback of high exposure. If the active component is introduced during the preparation process and subsequently exposed to centrifugation, washing, and separation. By adding chitosan solution (in acetic acid) to liquid paraffin containing a surfactant, a w/o emulsion is formed. Metformin hydrochloride microspheres are created by utilising a 25% solution of gluteraldehyde as a cross linking agent.

➤ **Double emulsion technique-** It is the manufacture of multiple emulsions, namely W/O/W, by pouring the main w/o emulsion into an aqueous solution of poly vinyl alcohol. For 30 minutes, this w/o/w emulsion was constantly stirred. Over a 30-minute period, gradually add some water to the emulsion. Filter the microcapsules and dry them under vacuum. It works well with water-soluble medicines, peptides, proteins, and vaccinations. This approach may be used with both natural and synthetic polymers. A lipophilic organic continuous phase disperses the aqueous protein solution.

➤ **Polymerization-** The two main procedures for preparing microspheres are classified as follows:

(1) **Standard polymerization-** To commence polymerization in bulk polymerization, a monomer or a combination of monomers, together with the initiator or catalyst, is commonly heated. The resulting polymer may be moulded into microspheres. Drug loading can be accomplished by including the medication during the polymerization process. It is a pure polymer production technology, however dissipating the heat of reaction is quite challenging. This has an impact on the thermo labile active substances. Suspension polymerization, also known as pearl polymerization, is performed at a lower temperature by heating the monomer combination with active medication as droplets dispersion in a continuous aqueous phase. The size of microspheres created via suspension methods is less than 100 m. Emulsion polymerization differs from suspension polymerization because of the existence of an initiator in the aqueous phase, but it is also performed at a low temperature since the exterior phase of the suspension is generally water in the previous two procedures, allowing heat to disperse readily.

(2) **Interfacial Phenomenon-** It entails the interaction of different monomers at the interface of two immiscible liquid phases to generate a polymer film that effectively envelops the liquid phase. The scattered phase. In this approach, two reactive monomers are used; one is dissolved in continuous phase, while the other is dispersed in continuous phase (aqueous in nature), and the second monomer is emulsified throughout. Because of the solubility of the produced polymer in the emulsion droplet, two circumstances exist. If the polymer is soluble in droplet, the carrier will be monolithic. If the polymer is insoluble in a droplet, the capsular type is generated. If the active component is introduced during the preparation process and subsequently exposed to centrifugation, washing, and separation. By adding chitosan solution (in acetic acid) to liquid paraffin containing a surfactant, a w/o emulsion is formed. Metformin hydrochloride microspheres are created by utilising a 25% solution of gluteraldehyde as a cross linking agent.

PHYSICO-CHEMICAL EVALUATION

- **Characterization**-Characterization of the microparticulate carrier is an essential phenomena that aids in the development of an appropriate carrier for protein, medication, or antigen delivery. The microstructures of these microspheres vary. These microstructures govern release and stability of the carrier.
- **Size and form of particles**-The most common methods for seeing microparticles are conventional light microscopy (LM) and scanning electron microscopy (SEM) (SEM). Both may be used to determine microparticle shape and exterior structure. In the case of double-walled microspheres, LM allows for control over coating settings. The architecture of the microspheres may be seen before and after coating, and the difference can be assessed microscopically. In comparison to the LM, the SEM gives better resolution. SEM enables analyses of the surfaces of microspheres and after particles are added.
- **Density estimation**-A multi volume pycnometer may be used to determine the density of the microspheres. A cup of accurately weighed sample is inserted in the multi volume pycnometer. Helium is a gas. The gas is fed into the chamber at a steady pressure and allowed to expand. As a result of this expansion, the pressure within the chamber decreases. Two successive values of pressure decrease at various beginning pressures are recorded. The volume and hence density of the microsphere carrier are calculated using two pressure measurements.
- **Point isoelectric**-The micro electrophoresis device is used to estimate the isoelectric point by measuring the electrophoretic mobility of microspheres. The mobility of microspheres can be connected to their surface charge, ionisable behaviour, or ion absorption nature.
- **Carboxylic acid residue on the surface**-Radioactive glycine is used to assess the surface carboxylic acid residue. The radioactive glycine conjugates are made by combining C14-glycine ethyl ester hydro chloride with microspheres. The conjugate's radioactivity is then determined using a liquid scintillation counter. As a result, the carboxylic acid residue may be compared and connected.
- **Amino acid residue on the surface**-The radioactive C14-acetic acid conjugate determines the surface associated amino acid residue. The amino acid residue can be estimated indirectly by measuring the carboxylic acid residue using a liquid scintillation counter.
- **Capture effectiveness**-Allowing washed microspheres to lyses can be used to estimate the capture efficiency or percent entrapment. The lysate is then tested for active components in accordance with the monograph requirements. The following equation is used to compute the percent encapsulation efficiency:

$$\% \text{ Entrapment} = \text{Real content/Theoretical content multiplied by } 100.$$

- **The contact angle**- The wetting property of a micro particle carrier is determined by measuring the angle of contact. It influences whether microspheres are hydrophilic or hydrophobic in nature at the solid/air/water interface, the angle of contact is measured.

➤ **Drug release**

- **In vitro Drug release** -In vitro drug release studies have been used as a quality control method in the manufacturing of pharmaceuticals, in the development of new products, etc. Data on releases that are sensitive and repeatable and that are drawn from physicochemical and hydrodynamically defined conditions essential, but no accepted in vitro technique has yet been created. Depending on the design and use of the developed dosage form, various employees have employed apparatus in a variety of settings.
- **Beaker technique**- Using an overhead stirrer, the dosage form is uniformly stirred while being made to adhere to the bottom of the beaker holding the medium. The amount of medium used in the research described in the literature ranges from 50 to 500 ml, and the stirrer's speed ranges from 60 to 300 rpm.
- **Mechanism of interface diffusion**-The authors of this approach are Dearden & Tomlinson. There are four sections in it. A originally held an appropriate drug concentration in a buffer and is a representation of the oral cavity. 1-octanol was present in compartment B, which represented the buccal membrane, and 0.2 M HCL was present in compartment C, which represented bodily secretions. 1-Octanol was also found in compartment D, which represented protein binding. The aqueous component and 1-octanol were saturated with one another prior to use. Samples were removed.
- **In vivo procedures**-Techniques that take advantage of the biological reaction of the organism locally or systemically and those that involve direct contact with the intact mucosa are methods for examining its permeability. local assessment of surface penetration uptake or accumulation. The most popular techniques include perfusion chambers for investigating drug permeability, in vivo studies using animal models, and buccal absorption tests.
- **Animal studies**- Animal models are primarily used for screening compound sequences, examining the processes and utility of permeation enhancers, or assessing a collection of formulations. There have been reports of using animal models of the canine, rat, rabbit, cat, hamster, pig, and sheep. In general, the process starts with anesthesia of the animal before administering the medication. In

rodents, the esophagus is strangulated to block absorption routes other than the oral mucosa. The blood is taken out and examined at various times.

- **In vivo In vitro Correlations-** The phrase "in vitro-in vivo correlations" refers to relationships between in vitro dissolution rates and the rate and degree of availability as assessed by blood concentration and/or urinary excretion of drug or metabolites. Such connections enable the development of products requirements for absorption.
- **Peak Plasma Concentration vs. Percent of Drug Dissolved in Vitro-** Measuring the percentage of the drug released from various dosage forms, as well as estimating the highest plasma concentrations attained by them, and then examining the correlation between them, are two methods of determining whether in vitro and in vivo results are consistent.

APPLICATION OF MICROSPHERE

1. **Ophthalmic Drug Delivery-** Microspheres made of polymer display positive biological behaviour such as bioadhesion, permeability increasing qualities, and intriguing physicochemical features, making it a unique material for the field.
2. **Oral Drug Delivery-** The ability of polymer-containing microspheres to form films allows for their use in the formulation of film dosage forms as an alternative to pharmaceutical tablets. Because of the pH sensitivity and reactivity of the primary amine groups Microspheres are more suited for oral medication delivery applications.
3. **Nasal drug delivery-** Polymer-based drug delivery systems, such as microspheres, liposomes, and gels, have been shown to have high bioadhesive properties and expand quickly when in contact with the nasal mucosa, improving drug bioavailability and residence duration. For example, starch, dextran, albumin, chitosan, and gelatin.
4. **Gene delivery-** Because of their adhesion and transport capabilities in the GI tract, microspheres might be a good oral gene carrier. Chitosan, gelatin, viral vectors, cationic liposomes, and polycation complexes are a few examples.
5. **Intratumoral and local drug delivery-** Polymer films are used to deliver paclitaxel to the tumour site at therapeutically relevant concentrations. The medication mixture offers considerable promise for regulated distribution in the oral cavity. For example, PLGA, Chitosan and PCL are both natural fibres.
6. **Drug delivery in the gastrointestinal tract-** When polymer granules with interior voids generated by deacidification are introduced to acidic and neutral environments, they float and give a controlled release of the medicine. Eudragit, Ethyl cellulose+Carbopol BSA, Gelatin, for example.
7. **Transdermal medication delivery-** Polymers have excellent film-forming characteristics. The drug release from the devices is influenced by the thickness of the membrane and the cross-linking of the film. Chitosan and alginate are two examples.
8. **Colonic medication delivery-** Polymers have been utilised to transport insulin to the colon. Chitosan, for example.
9. **Vaginal medication delivery-** A polymer modified by the addition of thioglycolic acid to the polymer's main amino groups is commonly used to treat mycotic infections of the genitourinary tract. Chitosan, gelatin, and PLGA are a few examples.
10. **Targeting using microparticulate carriers-** Polymer pellets are manufactured using extrusion/spheronization technique. Chitosan and microcrystalline cellulose are two examples.
11. **Medicine-** Prolonged release of proteins, hormones, and peptides; gene therapy utilising DNA plasmids; and insulin administration. Vaccine delivery for illnesses such as hepatitis, influenza, pertussis, ricin toxoid, diphtheria, and birth control. Passive targeting of leaky tumour vasculature, active targeting of tumour cells and antigens by intravascular/intravenous administration. Doxorubicin tumour targeting, as well as Leishmaniasis treatments. Magnetic microspheres can be used to collect stem cells and purge bone marrow. Used in antibody isolation, cell separation, and toxin extraction by affinity chromatography. Used in a variety of diagnostic tests for infectious disorders such as bacterial, viral, and fungal infections.

CONCLUSION

The present review article demonstrates that microsphere is a better option for novel drugs delivery system than other types of drugs delivery system. In this review, we take the position that improved therapeutic action, improved bioavailability, and decreased toxicity. Microspheres well discover that the significant role in novel drug delivery, namely in diseased cell sorting diagnosis, genetic material, safe targeted, and efficient in vivo administration system, in the future by combining a variety of other strategies. Compared to other delivery systems, microspheres have more potential to improve therapy today.

REFERENCES

1. Chowdary KPR, Yarraguntla SR. Mucoadhesive microsphere for controlled drug delivery. *Biol. Pharm. Bull.* 2004;1717-24.
2. Shanthi NC, Gupta R, Mahato KA, Traditional and Emerging Applications of Microspheres: A Review, *International Journal of Pharm. Tech Research.* 2010; 2(1):675-81.
3. Chandrawanshi P, Patidar H. Magnetic microsphere: As targeted drug delivery. *J. of Pharmacy Res.* 2009.
4. Najmuddin M., Ahmed A., Shelar S, Patel V, Khan T. Floating Microspheres Of Ketoprofen: Formulation and Evaluation, *International Journal Of Pharmacy and Pharmaceutical sciences.* 2010; 2(2):83-87.
5. Lopez CR, Portero A, Vila-Jato JC, Alonso MJ. "Design and Evaluation of Chitosan/Ethylcellulose mucoadhesive bilayered devices for buccal drug delivery." *J.control.Rel.* 1998;55: 143-52.
6. Alagusundaram M, Chetty MSC, Umashankari K, Badarinath AV, Lavanya C, Ramkanth S. Microspheres as a Novel Drug Delivery System- A Review. *Int. J. of Chem. Tech Res.*2009;1: 526-34
7. Chaturvedi G and Saha RN. A Review on Microsphere Technology And Its Application. *Birla institute of technology and sciences.* 2009:56-58.
8. Gholap SB, Banarjee SK, Gaikwad DD, Jadhav SL and Thorat RM. Hollow microsphere: a review, *International Journal of Pharmaceutical Sciences Review and Research.* ,2010;1:10-15.
9. Nachts S and Martin K. In: *The microsponges a novel topical programmable delivery formulation*, Marcel Dekker Inc. Newyork. 1990; 299.
10. Thanoo BC, Sunny MC and Jayakrishnan A. Cross-linked chitosan microspheres: Preparation and evaluation as a matrix for the controlled release of pharmaceuticals. *J Pharm Pharmacol.* 1992;44:283-286.
11. A Garg. S. Visht. P. K. Sharma and N. Kumar A review formulation charectrization and application on nanoparticals. *Der pharmasinica*, 2011; 17-26.
12. T. Lopez; JL Cuevas. G. Jardan. E. Gomez P. Ramirez and Tecaupetla, preparation and charectrization of anti-epileptic drug encapsulated in sol gel titaniannanoparticals as control release system, *medicinal chemistry*, 2015.
13. Gavini, A.B., Hegge, G. Rassu, V. Sanna, C., Testa as nasal administration of Carbamazepine using chitosan Microspere: In- vitro/ In-vivo studies. *International Journal of Pharmacy and Biological Sciences*, 2011; 121-129
14. Kosta A. K., Solakhia T. M., Dr. Agrawal S. Chitosan Nanoparticals- a drug delivery system. *Canadian Journal of Veterinary Research*, 2012; 737-743.
15. Pardesi C. V., Rajput P. V., Belgamvar V. S., Tekade A. R. Formulation , optimization and evaluation of spray-dried mucoadhesive microspheres as intranasal carriers for Valsartan. *Indian Journal of Pharmaceutical Sciences*, 2011; 64 (2)341-349.
16. Hafeli U. *Physics and Chemistry Basic of Biotechnology: Focus on biotechnology.* Review: Radioactive Microspheres for Medical Application, 7:213-248.
17. Parmar H., Bakliwal S., Gujarathi N., Rane B., Pawar S. Different methods of formulation and evaluation of mucoadhesive microsphere. *International Journal of PharmTech Research*, 2010; 1: 1157-1167.