



STUDY OF TOLERANCE IN SOME SPECIES OF PLANTS NEAR COPPER MINES IN KHETRI, JHUNJHUNU, RAJASTHAN

Dr. Anita

Assistant Professor, Department of Botany
R.K.J.K. Barasia P.G. College, Surajgarh, JJN. (Raj.)

Abstract

Bioaccumulation of copper in soil increases toxicity and this causes further phytotoxicity ie. Plant toxicity and environmental pollution risk increases. Some plant species are weakened due to copper toxicity whereas some plant species can tolerate it, due to high mineral content in soil as compared to copper, more absorption facilities promoted by VAM fungi and high nutrient supply by external sources. Water provision also washes away the effect of copper toxicity. The fertilizers can also subdue the copper toxicity effect ie. NPK fertilizers.

Keywords: tolerance, plants, species, copper mines, Khetri, Jhunjhunu, Rajasthan, mineral, toxicity, pollution

Introduction

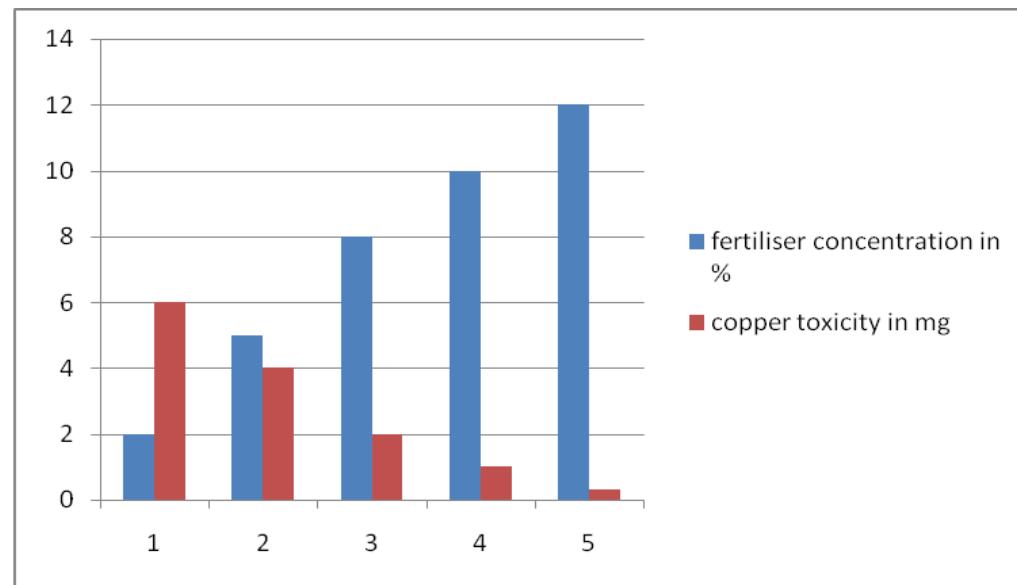
Copper tolerance was found in plant species colonizing soils with high concentrations of soil copper in Khetri, Jhunjhunu, Rajasthan. Seven of the eight plant species tested were found at more than one copper mine in the present study.[1,2] The mines are geographically isolated, which makes dispersal of seeds from one mine to another unlikely. Tolerance has probably evolved independently at each site in this area. The plant in this region namely *Vulpia microstachya* displays significantly higher tolerance to copper at all copper concentration levels. The shoot tissue of the copper mine plants of *Arenaria douglasii*, *Bromous mollis*, and *V. microstachya* accumulated less copper in drought region of Khetri due to stress conditions and more water supply externally. [3,4]The root tissue of these mine plants contain more copper than the roots of the non mine plants in the drought region due to presence in soil. No difference in the tissue copper concentration was detected between tolerant and non tolerant plants of *Lotus purshianus*, *Lupinus bicolor*, and *Trifolium pratense* even though the root tissue had more copper than the leaves.

This is a fact that copper (Cu) is an essential element for humans and plants when present in lesser amount, while in excessive amounts it exerts detrimental effects. There subsists a narrow difference amid the indispensable, positive and detrimental concentration of Cu in plants, which substantially alters with Cu speciation,[5,6] and form in plants. Mechanisms related to detoxification strategies like antioxidative response and generation of glutathione and phytochelatins to combat Cu-induced toxicity in plants can be done due to which copper tolerance increases in plants and plants can survive. [7,8]

Cu concentration in plants beyond critical limits affects the plant growth, promotes leaf chlorosis and causes cytotoxicity as seen in drought region in Khetri, Jhunjhunu, Rajasthan. Phytoremediation is an evolving approach which uses plants to decontaminate the polluted soil/water especially near copper mines in Khetri region. There are many plants which have phytoremediation potential and are best accrual of heavy metals like copper. The tolerance, accrual and transport of metals to the aerial tissues of plants are vital features which are taken into consideration

during species choice in phytoremediation strategy[9,10] . The application of Cu-amended fertilizers is usual in nation's deficient of Cu in the soils. Certain NPK fertilizers increase soil minerals and mineral content in plants thus decreasing copper toxicity in plants and increasing copper tolerance in stress conditions of Khetri region[11,12].

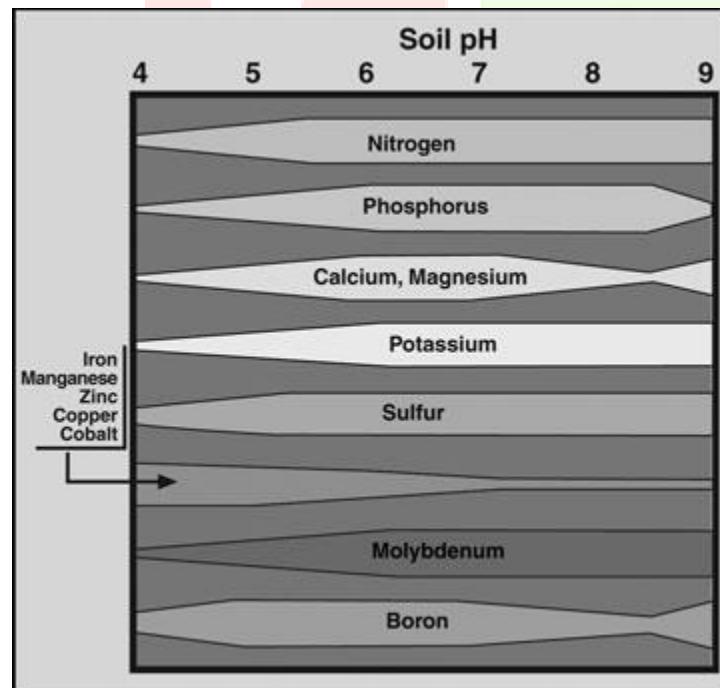
Discussion



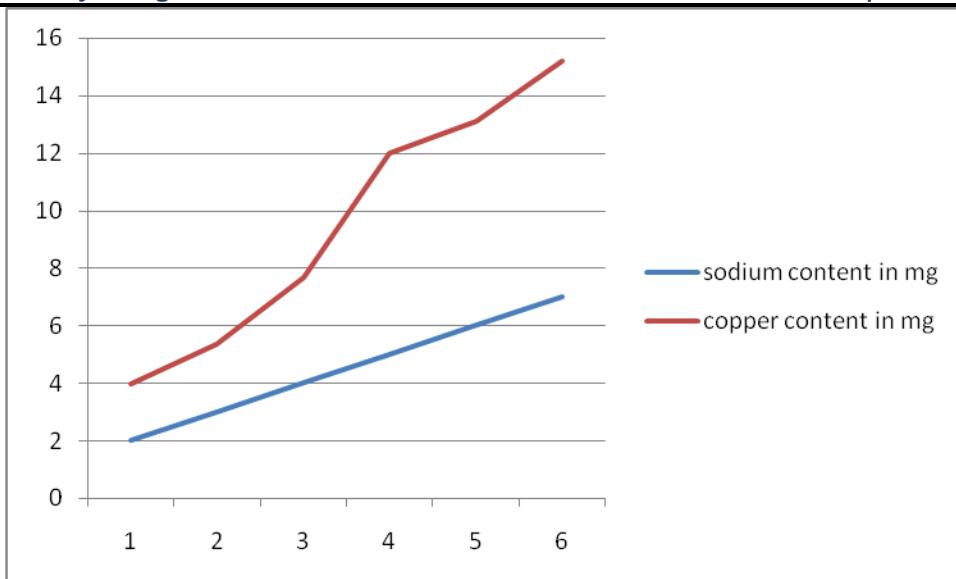
Graph1 -showing decrease in copper toxicity in plants of Khetri by increasing fertiliser concentration

However, excessive sodium concentration in soil of drought region of Khetri , Rajasthan, increases the copper toxicity due to more stress in soil which is usually dry and naturally there is no provision of water (graph-2).

Fig. Soil pH affects nutrient availability to plants. The width of the band indicates the relative availability of each plant nutrient at various pH levels.



Graph-2: At high sodium concentrations the toxicity of copper increases in plants



Results

Among different plant species in Khetri, Jhunjhunu, Rajasthan, only few plant species got germinated at 300 mg Cu kg⁻¹. The growth parameters were significantly reduced under high Cu stress (from 25 to 100 mg Cu kg⁻¹) in and drought conditions, also the sodium content increased in this drought region.[13,14] The chlorophyll content found decreased in copper toxic regions of mine sites, but the supply of NPK fertilizers could increase chlorophyll content. Significantly, high Cu accumulation was found in roots and shoots of drought plants in Khetri and few were only tolerant due to external provision of water and minerals. [15,16]

Conclusions

Copper is an essential microelement, for plants is nutritive for plant growth but under stress conditions as in Khetri, Jhunjhunu its high prevalence in soil causes copper phytotoxicity.[17,18] We can reduce this by providing fertilizers externally and water containing minerals. The copper tolerance increases in plants and plants survive more in comparison to common conditions of drought. [19,20]

References

1. Adhikari, T., Kundu, S., Biswas, A. K., Tarafdar, J. C., & Rao, A. S. (2012). Effect of copper oxide nano particle on seed germination of selected crops. *Journal of Agricultural Science and Technology*, A, 2(6A), 815. [Google Scholar]
2. Ahsan, N., Lee, D. G., Lee, S. H., Kang, K. Y., Lee, J. J., Kim, P. J., & Lee, B. H. (2007). Excess copper induced physiological and proteomic changes in germinating rice seeds. *Chemosphere*, 67, 1182–1193. [Crossref], [Web of Science ®], [Google Scholar]
3. Ali, S., Shahbaz, M., Shahzad, A. N., Fatima, A., Khan, H. A., Anees, M., & Haider, M. S. (2015). Impact of copper toxicity on stone-head cabbage (*Brassica oleracea* var. *capitata*) in hydroponics. *PeerJ*, 3, e1119. [Crossref], [Web of Science ®], [Google Scholar]
4. Ansari, M. K. A., Oztekin, E., Ahmad, A., Umar, S., Iqbal, M., & Owens, G. (2013). Identification of the phytoremediation potential of Indian mustard genotypes for copper, evaluated from a hydroponic experiment. *Clean: Soil Air Water*, 41, 789–796. [Crossref], [Web of Science ®], [Google Scholar]
5. Arnon, D. I. (1949). Copper enzymes in isolated chloroplasts: Polyphenol oxidase in *beta vulgaris*. *Plant Physiology*, 24, 1–15. [Crossref], [Web of Science ®], [Google Scholar]
6. Audet, P., & Charest, C. (2007). Heavy metal phytoremediation from a meta-analytical perspective. *Environmental Pollution*, 147, 231–237. [Crossref], [Web of Science ®], [Google Scholar]
7. Awokunmi, E. E. (2016). The potential of *abelmoschus esculentus* in EDTA-assisted phytoextraction of heavy metals from soil of Bashiri Dumpsite, Ado Ekiti, Nigeria. *International Journal of Environmental Protection*, 6, 9–14. [Google Scholar]

8. Azooz, M. M., Abou-Elhamd, M. F., & Al-Fredan, M. A. (2012). Biphasic effect of copper on growth, proline, lipid peroxidation and antioxidant enzyme activities of wheat (*Triticum aestivum*'cv. Hasaawi) at early growing stage. *Australian Journal of Crop Science*, 6, 688–694. [Google Scholar]
9. Barbosa, R. H., Tabaldi, L. A., Miyazaki, F. R., Pilecco, M., Kassab, S. O., & Bigaton, D. (2013). Foliar copper uptake by maize plants: Effects on growth and yield. *Ciencia Rural*, 43, 1561–1568. [Crossref], [Web of Science ®], [Google Scholar]
10. Carolin, C. F., Kumar, P. S., Saravanan, A., Joshua, G. J., & Naushad, M. (2017). Efficient techniques for the removal of toxic heavy metals from aquatic environment: A review. *Journal of Environmental Chemical Engineering*, 5(3), 2782–2799. [Crossref], [Web of Science ®], [Google Scholar]
11. Ciura, J., Poniedziałek, M., Sękara, A., & Jędrzczak, E. (2005). The possibility of using crops as metal phytoremediants. *Polish Journal of Environmental Studies*, 14, 17–22. [Web of Science ®], [Google Scholar]
12. Cui, S., Zhou, Q., & Chao, L. (2007). Potential hyperaccumulation of Pb, Zn, Cu and Cd in endurant plants distributed in an old smeltery, northeast China. *Environmental Geology*, 51, 1043–1048. [Crossref], [Web of Science ®], [Google Scholar]
13. Dresler, S., Hanaka, A., Bednarek, W., & Maksymiec, W. (2014). Accumulation of low-molecular-weight organic acids in roots and leaf segments of *Zea mays* plants treated with cadmium and copper. *Acta Physiologiae Plantarum*, 36, 1565–1575. [Crossref], [Web of Science ®], [Google Scholar]
14. Fanrong, Z., Shafaqat, A., Haitao, Z., Younan, O., Boyin, Q., Feibo, W., & Guoping, Z. (2011). The influence of pH and organic matter content in paddy soil on heavy metal availability and their uptake by rice plants. *Environmental Pollution*, 159, 84–91. [Crossref], [Web of Science ®], [Google Scholar]
15. Fitz, W. J., & Wenzel, W. W. (2002). Arsenic transformation in the soil rhizosphere plant system, fundamentals and potential application of phytoremediation. *Journal of Biotechnology*, 99, 259–278. [Crossref], [Web of Science ®], [Google Scholar]
16. Gopal, R., & Rizvi, A. H. (2008). Excess lead alters growth, metabolism and translocation of certain nutrients in radish. *Chemosphere*, 70(9), 1539–1544. [Crossref], [Web of Science ®], [Google Scholar]
17. Hanen, Z., Tahar, G., Abelbasset, L., Rawdha, B., Rim, G., Majda, M., ... Chedly, A. (2010). Comparative study of Pb-phytoextraction potential in *Sesuvium portulacastrum* and *Brassica juncea*: Tolerance and accumulation. *Journal of Hazardous Materials*, 183, 609–615. [Crossref], [Web of Science ®], [Google Scholar]
18. Hegedus, A., Erdei, S., & Horvath, G. (2001). Comparative studies of H₂O₂ detoxifying enzymes in green and greening barley seedlings under cadmium stress. *Plant Science*, 160, 1085–1093. [Crossref], [Web of Science ®], [Google Scholar]
19. Herawati, N., Suzuki, S., Hayashi, K., Rivai, I. F., & Koyoma, H. (2000). Cadmium, copper and zinc levels in rice and soil of Japan, Indonesia and China by soil type. *Bulletin of Environmental Contamination and Toxicology*, 64, 33–39. [Crossref], [Web of Science ®], [Google Scholar]
20. Kabata-Pendias, A., & Pendias, H. (2001). *Trace elements in soils and plants* (3rd ed.). Boca Raton: CRC Press. [Google Scholar]