



NATURAL MEDICINE FOR DIABETES AS DIPEPTIDYL PEPTIDASE-IV [DPP-IV] INHIBITOR

Dr. Avani N Doshi^{1*}, Bhumi M Shah^{2,3}, Nidhi H Jadav⁴, Geetika S Sharma⁵ & Dr. Dhruvo Jyoti Sen⁶

¹Department of Pharmaceutical Chemistry, K. B. Raval College of Pharmacy, Shertha, Post Kasturinagar, B/h IFFCO township, Gandhinagar, Gujarat, 382443, India.

²Department of Pharmaceutical Chemistry, K. B. Institute of Pharmaceutical Education and Research, Gandhinagar, Gujarat 382023, India.

³Department of Pharmaceutical Chemistry, L. J. Institutes of Pharmacy, L J University, Ahmedabad, Gujarat 382210, India.

^{4,5}Department of Pharmaceutical Chemistry SAL Institute of Pharmacy, Opp Science City, Ahmedabad, Gujarat 380060, India.

⁶School of Pharmacy, Techno India University, Salt Lake City, Sector-V, EM-4/1, Kolkata-700091, West Bengal, India.

ABSTRACT: DPP-IV is the main point of target for the treatment of type 2 diabetes. Plant products are always accessible for the attainable lead generation for various diseases. In this review, we tried to cover many natural sources which exhibit antidiabetic action mainly due to inhibition of DPP-IV. The most potent chemical constituents of the plant as DPP-IV inhibitors were resveratrol, luteolin, apigenin and flavone. Now a day, it is very important to identify DPP-IV inhibitors which can act as forthcoming antidiabetic agent. Already available synthetic inhibitors like sitagliptin, vildagliptin, saxagliptin showed unenviable side effects. Phenolic compound and flavonoids have antioxidant properties as well being present in many functional foods. So, the researchers try to uncover the natural sources for lead generation as DPP-IV inhibitors.

Keywords: Active constituents, Diabetes, DPP-IV inhibitors, DPP-IV, Natural products

INTRODUCTION: Diabetes is a disease that involve problems with the hormone insulin. Diabetes mellitus is a clinical condition which is characterized by an increase in plasma blood glucose. Normally, the pancreas releases insulin to help your body store and use the sugar and fat from the food you eat but in diabetes pancreas is unable to produce insulin or our body cannot utilize the insulin.¹

Diabetes is a chronic disorder of interference in metabolism characterized by increased fasting and post prandial blood sugar levels. The global preponderance of diabetes expected to be increase 5.4 % by the year 2025. It is estimated that there are approximately 33 million adults with diabetes in India which is going to be increase to 57.2 million by the year 2025. Type I diabetes (Insulin dependent) is caused due to insulin insufficiency because of lack of functional beta cells.² Patients suffering from type-I are therefore totally dependent on exogenous source of insulin while patients suffering from Type II diabetes (Insulin independent) are unable to respond to insulin and can be treated with changes in diet, exercise or medication.

Plants as DPP-IV inhibitors: *Momordica charantia* (Ayurvedic/Sanskrit name: Karavella, Kathilla; Common name: Karela, Bitter Gourd)

Momordica charantia commonly known as bitter gourd or karela or bitter melon, used for many medicinal purposes like constipation, inflammation, cough, microbial infection, and respiratory diseases, hyperglycaemia etc.⁶ Singh J. et al. have isolated an alkaloid, Momordicin (**Figure-3**) and a steroidal saponin, Charantin (momocharin) (**Figure-3**) from fruit of the plant and exhibit hypoglycaemic effect.⁷ Bhat G A. et al. have carried out a study effect of aqueous extract of plant. Aqueous extract increases tissue glycogen, serum insulin and GLP-1 non-significantly. The polar molecules of *M. charantia* depolarize the L-cell through elevation of intracellular Ca^{2+} concentration which releases GLP-1 leads to elevate beta-cell proliferation and insulin secretion.⁸ Methanolic extract (0.5 mg/ml) of *M. charantia* showed 53.25 % DPP-IV inhibition and the extract has phenolic content which increases DPPH free radical scavenging activities.⁹

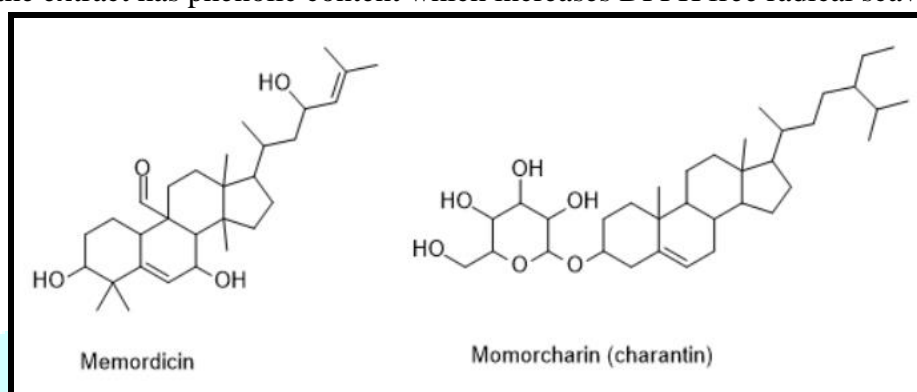


Figure-3: Active constituents of *Momordica Charantia* as DPP-IV inhibitors

Cassia nigricans (Ayurvedic/Sanskrit name: Aragvadhā; Common name: Senna)

Cassia nigricans, Senna is an especially important plant of Ayurvedic medicines which possesses anti-inflammatory, antioxidant, hypoglycemic and anticancer activities.¹⁰ Saidu Y. et al. have prepared methanolic, ethanolic and hexane fractions of the plant. All of them showed 63.1 % inhibition of DPP-IV which is less effective than standard, P32/98.¹¹ Further *in-vitro* enzyme inhibition was carried out by methanolic extract of *Cassia nigricans* ($57.0 \pm 1.91\%$) by Saidu y. et al. which serves as sources of inhibitors of the DPP-IV enzyme in the treatment of Type-2 diabetes mellitus.¹²

Ocimum sanctum (Ayurvedic/Sanskrit name: Gouri, Bhuteshta; Common name: Tulsi)

Inbaraj S. et al. has been reported that *O. sanctum* possess anti diabetic, antifertility, anti-cancer, anti-fungal, hepatoprotective and cardioprotective actions.¹³ Somasundaram G. et al. have reported that methanolic extract of *Ocimum sanctum* possesses good DPP-IV inhibition activity with % inhibition $66.81 \pm 0.05\%$. The chemical constituents like ursolic acid and oleanolic acid (**Figure-4**) (Triterpenoids) present in leaf extract showed DPP-IV inhibition action and potentiation of insulin release.¹⁴ Singh A K. et al. have showed α -glucosidase and DPP-IV enzyme inhibition effects by *in-vitro* method using leaf extract. *Ocimum basilicum* contains almost 10% eugenol (**Figure-4**) which is found to lower the blood glucose level.¹⁵

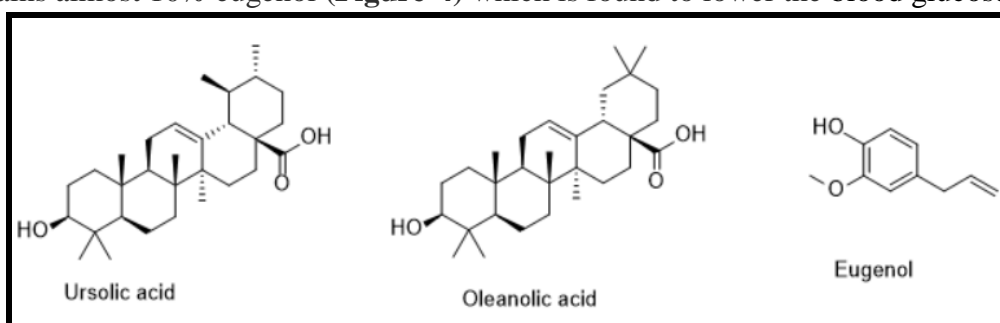


Figure-4: Active constituents of *Ocimum sanctum* as DPP-IV inhibitors

Gymnema sylvestre (Ayurvedic/Sanskrit name: Madhunashini; Common name: Gudmar)

Gymnema sylvestre is used as sugar destroyer because chewing of leaves destroys the ability to identify the sweet taste in case of glycosuria and other urinary diseases. The plant extract of *Gymnema sylvestre* [Gurmar] was showed better *in vitro* DPP-IV ($60.21 \pm 0.35\%$) inhibitory activity.¹⁶ This plant has the

potential to be developed as a natural alternative to synthetic DPP-IV inhibitors. In terms of utilization by the pharmaceutical companies, research outcomes of this study may play a key role in the development of natural indigenous DPP-IV inhibitors.¹⁷ Laha S. et al. have isolated primary chemical constituents of *Gymnema* as DPP-IV inhibitors include gymnemic acid (**Figure-5**) identified when leaves of this plant reduces urine glucose in diabetes. Also, Arun L. B. et al. have identified *G. sylvestre* as an alternative to synthetic DPP-IV inhibitors by screening their extract for invitro DPP-IV inhibitory activity.¹⁸

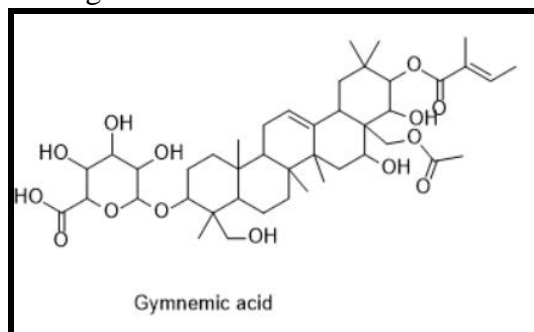


Figure-5: Active constituents of *Gymnema sylvestre* as DPP-IV inhibitors

***Mangifera indica* (Ayurvedic/Sanskrit name: Aamra, Sahakara; Common name: Mango)**

It is commonly known as mango, is a species of flowering plant in the sumac and poison family Anacardiaceae. Mangiferin is a pharmacologically active flavonoids, which having the anti-bacterial, antioxidant, anticancer, antidiabetic, hepatoprotective, anti-inflammatory activities.¹⁹ Suman R k et al. have isolated the chemical constituents from the barks of *Mangifera indica*. Their structures were identified as mangiferone, mangiferin, myricetin, myricitrin, rutin and quercetin. The active constituent quercetin (**Figure-6**) is act as DPP-IV inhibitors.²⁰ Methanolic extract of *Mangifera indica* leaves was prepared by Kalita P. et al. which showed potent DPP-IV inhibitory activity ($68.22 \pm 1.14\%$).²¹ Also, methanolic extract prepared by Yoshida S. et al. showed potent activity with an IC_{50} value of $182.7\mu\text{g/ml}$. *Mangifera indica* inhibits the DPP-IV and enhances the GLP-1 for type 2 diabetes. The peptide is rapidly inactivated by the enzyme dipeptidyl peptidase-IV (DPP-IV), resulting in a half-life of active GLP-1 of only approximately 1-2 minutes. Inhibition of DPP-IV increases the levels of endogenous active GLP-1 and prolongs its half-life.²²

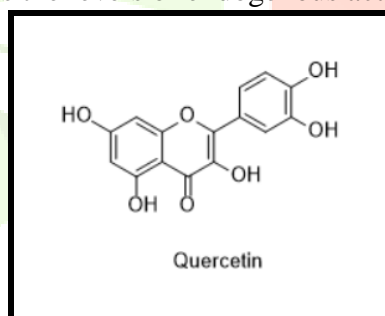


Figure-6: Active constituents of *Mangifera indica* as DPP-IV inhibitors

Bodhi Leaves (Ayurvedic/Sanskrit name: Bodhivriksha; Common name: Peepal)

It is known as peepal tree family (*F. religiosa* L.). Fruits are used as laxatives, latex is used as a tonic, and fruit powder is used to treat asthma. It is also used traditionally antiulcer, antibacterial, antidiabetic, in the treatment of gonorrhea and skin diseases²³. The ethanolic extract of Bodhi leaves showed highest (68.98 ± 1.95) inhibitory activity against DPP-IV enzyme that plays a role in improving diabetes²⁴. Plant extract of 100, 200 and 400 mg/kg reduced the blood glucose level by 33%, 53% and 54% after three weeks. Plectranthoic acid (**Figure-7**) showed inhibition of glucosidase, amylase and DPP-IV activities.²⁵

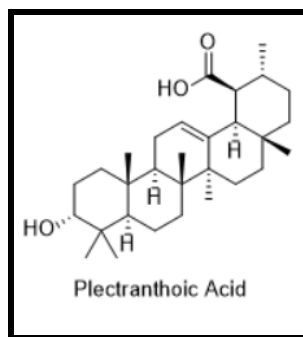


Figure-7: Active constituents of *Ficus religiosa* as DPP-IV inhibitors

Garlic (Ayurvedic/Sanskrit name: Lasunah, Rasonah; Common name: Lasun)

Garlic (*Allium sativum*) naturally belongs to a member of the Alliaceae family, is well recognized as a principal spice, and is additionally used as a remedy for specific ailments and physiological disorders. Allicin, (**Figure-8**) a sulfur-containing compound is responsible for its pungent odour and it has been shown to have significant hypoglycemic activity. This effect is thought to be due to increased hepatic metabolism, increased insulin release from pancreatic beta cells and/or insulin sparing effect.²⁶ Several other research including studies on effect of garlic extract on blood glucose levels and lipid profiles in streptozotocin/alloxan-induced diabetic rats, alloxan diabetic rabbits have exhibited its antidiabetic activity.²⁷⁻³⁰ Apart from this, *Allium sativum* exhibits antimicrobial, anticancer and cardio protective activities also. Ultrasonic-assisted garlic extract has been demonstrated as a promising anti-diabetic agent by inhibiting therapeutic drug target dipeptidyl peptidase IV, results showed that 70.9 µg/ml of garlic bulb extract inhibited 50% DPP-IV activity.³¹

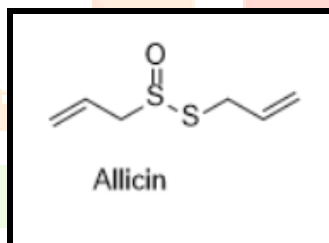


Figure-8: Active constituents of *Allium sativum* as DPP-IV inhibitors

***Caesalpinia sappan* (Ayurvedic/Sanskrit name: Patangah; Common name: Sappan wood)**

Sappan wood (*Caesalpinia sappan* L.) is a Brazilian plant that is referred to as Brazil wood. Brazilin (**Figure-9**) sappan wood major compound and important bioactive, has been used to treat diabetes through several biomolecular mechanisms. Previously 100 ppm of sappan wood extract had DPP-IV inhibition activity of 84.24%.³² Sappan wood extract with chlorine chloride-lactic acid of 0.60 ppm, 1.60 ppm and 2.03 ppm showed DPP-IV inhibition 4.54 %, 5.01%, and 5.72 %, respectively while Wood extract with Betaine-Lactic acid of 0.73 ppm, 1.61 ppm, 2.08 ppm showed DPP-IV inhibition with 5.70 %, 6.72 % and 7.74 %, respectively.³³

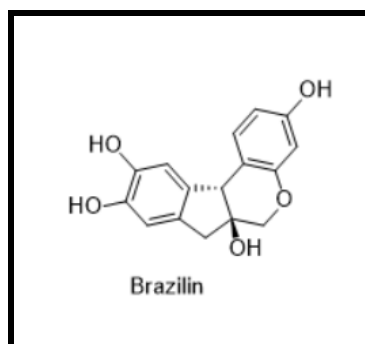


Figure-9: Active constituents of *Caesalpinia sappan* as DPP-IV inhibitors

***Camellia sinensis* (Ayurvedic/Sanskrit name: Syamaparni; Common name: Tea)**

Camellia sinensis (Tea) is a traditional health beverage in China and Indian medicine, and its antioxidant, antidiabetic, anti-hypertensive functions have been confirmed. For instance, tea polysaccharides can inhibit alpha-glucosidase, which is related to type 1 diabetes^{34,35} and tea polyphenol is one of the effective antidiabetic components in decreasing glucose levels and preventing complications.³⁶⁻³⁹

Chinese tea which is used as beverage and food supplements, is made from the leaves and leaf buds of the *Camellia sinensis* species and used for finding DPP-IV inhibitory effects. Aqueous extract of black tea after ultra-filtration showed very good DPP-IV inhibition *in-vitro*.⁴⁰

White tea (*Camellia sinensis* (L.) Kuntze) is a tea bud which is still roll processed without fermentation. The methanol fraction of the tea bud showed IC₅₀ value 227 µg /mL which is the most active inhibitor of DPP-IV enzyme. The inhibitory activity determined by using rat blood serum as DPP-IV enzyme source (*ex-vivo*).⁴¹

EGCG, a major flavonol (**Figure-10**) in tea, was shown to have antidiabetic activities in rodents. EGCG appears to have multiple antidiabetic actions including islet protection, increasing insulin secretion, decreasing insulin tolerance, and decreasing gluconeogenesis and insulin-mimetic action.^{42,43} The role of EGCG in islet protection was shown to protect against cell death mediated by islet amyloid polypeptide (IAP) *in-vitro*.^{44,45} EGCG was reported to activate AMPK in adipocytes.⁴⁶ *In-vitro*, the concentration of Epigallocatechin-3-gallate (EGCG) required to inhibit DPP-IV activity by 50% (the IC₅₀ value) was 28.42 µM. These data provide a theoretical basis for intervention in glucose metabolism with EGCG.⁴⁷

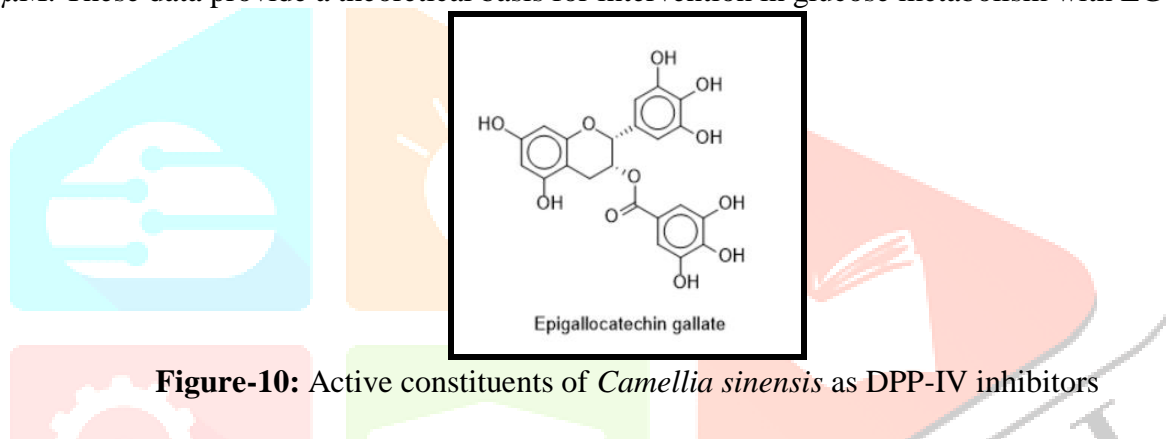


Figure-10: Active constituents of *Camellia sinensis* as DPP-IV inhibitors

***Berberine* (Ayurvedic/Sanskrit name: India barberry, Daruharidra; Common name: Berberina)**

Berberis aristate commonly known as “Dāruhaldhi” and “Citra” used in skin diseases, menorrhagia, diarrhea, jaundice, fever, diabetes, hepato-biliary disorder.⁴⁸ Phytochemical screening of methanolic extracts of leaves, stems and roots of *B. aristate* showed the presence of active compounds such as alkaloids (berberine) (**Figure-11**), reducing sugars, steroids, flavonoids, terpenoids, glycosides and saponins while tannins were found to be absent.⁴⁹ Chakrabarti R. et al. have found that methanolic extract of the bark of berberis aristate shown comparable DPP-IV inhibition (IC₅₀ - 14.4 µg/ml) with standard diprotin-A (IC₅₀ - 15 µg/ml).⁵⁰

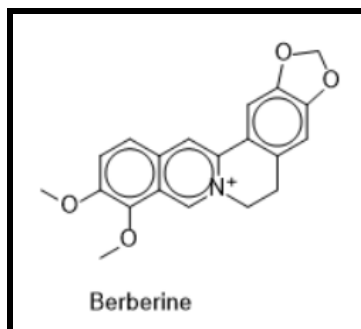


Figure-11: Active constituents of *Berberis aristate* as DPP-IV inhibitors

***Terminalia arjuna* (Ayurvedic/Sanskrit name: Arjuna; Common name: Arjuna tree)**

Terminalia arjuna is widely used in Indian system of medicine as cardioprotective, hypotensive, antidiabetic, hypolipidemic and wound healing activity.⁵¹ Borde M. K. et al. reported that extract of *Terminalia arjuna* exhibited good DPP-IV inhibition activity with the highest percentage of DPP-IV inhibition 83.39±7.58%.⁵² Ipseeta et al. concluded that the active ingredient of *Terminalia arjuna*; Arjunetin, Arjungenin, Ellagic acid

and Arjunic acid (**Figure-12**) showed superior DPP-IV Inhibitory activity as compared to synthetic DPP-IV inhibitors (Sitagliptin and Vildagliptin) based on results of docking studies.⁵³

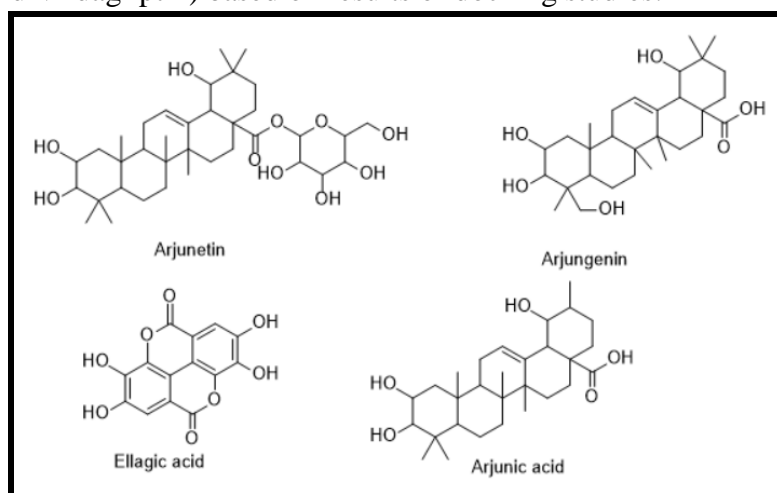


Figure-12: Active constituents of *Terminalia arjuna* as DPP-IV inhibitors

***Withania somnifera* (Ayurvedic/Sanskrit name: Kamrupini; Common name: Indian winter cherry)**

Withania somnifera (L.) is commonly known as Ashwagandha. In the traditional system of Ayurvedic medicine, this plant is claimed to have potent aphrodisiac rejuvenate and useful in the treatment of anti-inflammatory, antitumor, antistress, antidiabetic, antiaging, stimulant for neurotransmitter and life prolonging properties.^{54,55} Therefore, Kempegowda P K. et al. have tried to evaluate the extract of roots, leaves and fruits of *Withania somnifera* plant by *in-vitro* inhibitory assay. The methanolic extract of the root showed the maximum DPP-IV inhibition activity compared to other extract. Catechin (**Figure-13**) present in methanolic extract of the root showed the potent DPP-IV inhibition (>80 % inhibition at 125 µg/ml concentration) which is also proved by molecular docking study.⁵⁶

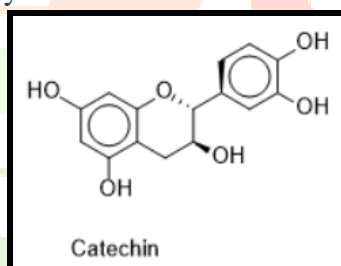


Figure-13: Active constituents of *Withania somnifera* as DPP-IV inhibitors

***Trigonella foenum-graecum* [Fenugreek] (Ayurvedic/Sanskrit name: Samudra methi; Common name: Methi)**

Trigonella foenum-graecum [Fenugreek] is used to treat numerous health problems, including insulin resistance, diabetes, poor appetite, inflammation, digestive problems and menopausal symptoms. 4-hydroxyisoleucine, a novel amino acid from fenugreek seeds increases glucose stimulated insulin release. In animal experiments, it has been shown that oral administration of plant extract decreased the blood glucose levels. Administration of fenugreek seeds improved glucose metabolism and reduced hepatic and renal glucose-6-phosphatase and fructose-1,6-biphosphatase activity.⁵⁷ Chemical constituents of the plant include saponins, many of which are glycosides of diosgenin. The seeds also contain the alkaloids trigonelline, gentianine, and carpaine (**Figure-14**) compounds. Seed constituents include 4-hydroxyisoleucine, an amino acid, and fenugreekine. It has been shown to increase erythrocyte insulin receptors and improve peripheral glucose utilization, thus showing potential pancreatic as well as extra pancreatic effects.⁵⁸ Fenugreekine, a steroidal saponogenic peptide ester, may have hypoglycemic properties. Trigonelline, another component, may exert hypoglycemic effects in healthy patients without diabetes, but other studies have shown that fenugreek has no effect on fasting or postprandial blood glucose levels in no diabetic subjects. Fenugreek seeds provided the highest inhibitory activity against DPP-IV enzyme *in-vitro* at a concentration of 2.5 µg/ml. That plays a role in improving diabetes. The highest inhibitory percentage was given by the ethanol extract of fenugreek seeds of *Trigonella foenum-graecum* L. is 71.29 ± 0.33 .⁵⁹

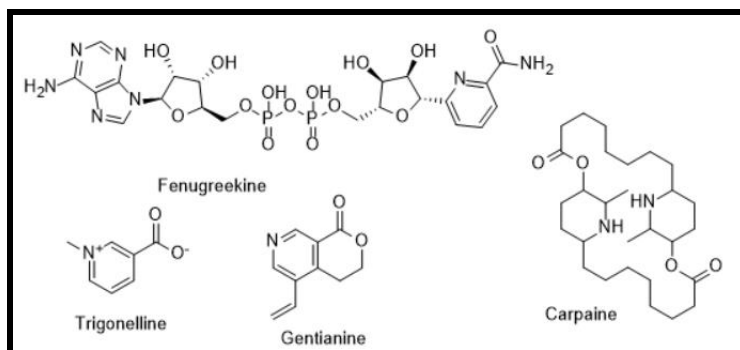


Figure-14: Active constituents of *Trigonella foenum* as DPP-IV inhibitors

***Eucalyptus globulus* (Ayurvedic/Sanskrit name: Tailapatra; Common name: Blue gum)**

Eucalyptus which is commonly known as gum tree. The bark and leaves of the eucalyptus tree have been used for many diseases. Dey B. et al. have evaluated leaf extract of *E. globulus*, *E. citriodora*, *E. camaldulensis* as natural enzyme inhibitors. Polyphenol and flavonoids are present in these variety. Through correlation study they came to know that the phenolic content has more correlation with the inhibitory potential of enzymes. Triterpenoids and Phenolic compounds (**Figure-15**) of the plant is mainly responsible for DPP-IV inhibition. The leaf extract of *E. globulus*, *E. citriodora*, *E. camaldulensis* showed DPP-IV inhibition with IC_{50} 3.098, 6.138, 3.99, respectively.⁶⁰ Kato E. et al. have isolated macrocarpals A–C from *Eucalyptus globulus* which is acting as DPP-IV inhibitors. Amongst them macrocarpals C (**Figure-15**) showed the most potent activity.⁶¹

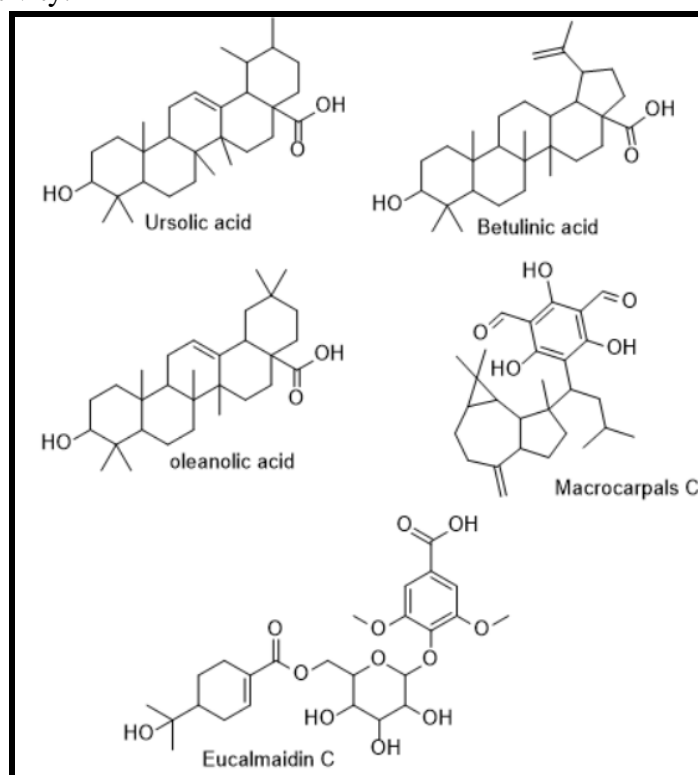


Figure-15: Active constituents of *Eucalyptus globulus* as DPP-IV inhibitors

***Emblica officinalis* (Ayurvedic/Sanskrit name: Amalki; Common name: Amla, Indian gooseberry)**

Majeed M. et al. extracted β -glucogallin (**Figure-16**) from the fruits and checked for antidiabetic activity. The fruit extract inhibited α -amylase and α -glucosidase enzyme with IC_{50} values of $135.70 \mu\text{g mL}^{-1}$ and $106.70 \mu\text{g mL}^{-1}$, respectively. Also, the fruit extract showed inhibition of dipeptidyl peptidase-IV enzyme with IC_{50} value $3770 \mu\text{g mL}^{-1}$.⁶²

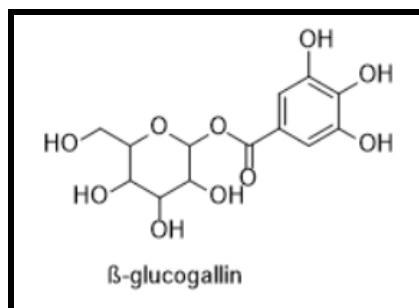


Figure-16: Active constituents of *Emblica officinalis* as DPP-IV inhibitors

***Rosa gallica* (Ayurvedic/Sanskrit name: Devataruni; Common name: Gulab)**

Kato E et al. have evaluated Ellagitannins were isolated as DPP-IV inhibitors from rose bud extract powder. Inhibiting DPP-IV protects GLP-1 from degradation, which enhances the secretion of insulin to help our body decrease the blood sugar level. Rugosin A and Rugosin B act as DPP-IV inhibition at 100μM (%).⁶³

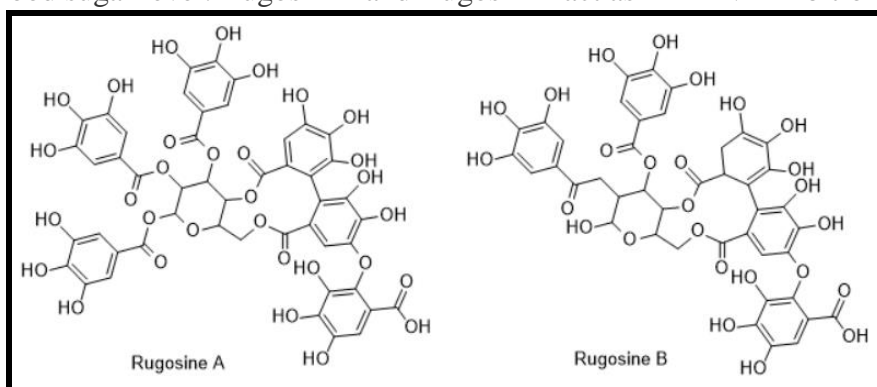


Figure-17: Active constituents of *Rosa gallica* as DPP-IV inhibitors

***Commiphora mukul* (Ayurvedic/Sanskrit name: Guggulu; Common name: Guggul)**

C. mukul is commonly known as Indian bdellium tree. Sharma D. et al. have isolated the percentage inhibition of *C. mukul* (92.97±8.45%) shows that it is a potent inhibitor of DPP-IV enzyme.⁶⁴ Sharma B. et al. have tried to evaluated *C. mukul* exerts anti-diabetic activity via DPP-IV inhibition. It guggulsterone act as DPP-IV inhibit enzyme, which can help to cure type II diabetes.⁶⁵

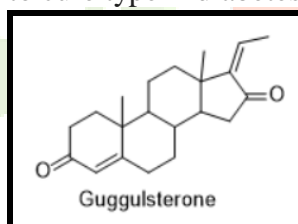


Figure-18: Active constituents of *Commiphora mukul* as DPP-IV inhibitors

***Punica granatum* (Ayurvedic/Sanskrit name: Dadima; Common name: Pomegranate)**

Punica granatum L. (Leguminosae) it is also known as Pomegranate, fruit-bearing shrub belong to the family Leguminosae. Research by Xinjiang has proved that all three plants extract i.e *Trigonella foenum-graecu*, *Cicer arietinum*, and *Punica granatum* has potent inhibition effects on DPP-IV, with IC₅₀ values of 0.03, 0.09, and 0.19 mg/mL, respectively.⁶⁶ Fruit peel of pomegranate plant *Punica granatum* were subjected to extraction with four solvents (distilled water, 80% methanol, 80% acetone and a mixed solvent that included methanol, ethanol, acetone and n-butanol at proportions (7:1:1:1 v/v/v/v), respectively. Methanol extract recorded the highest DPP-IV activity. (47.1 ± 1.5%).⁶⁷ Methanol extraction of pomegranate peels are recommended to be a target for investigations involved in the development of anti-T2DM and anti-cancer therapies. Components in pomegranate (punicalagin and ellagic, gallic, oleanolic, ursolic and uallic acids) were found to have antidiabetic effect. *Punica granatum* inhibit IRS-1 (Insulin receptor substrate 1), GLUT-2 (Glucose transporter 2) and GLUT-4 (Glucose transporter 4).⁶⁸

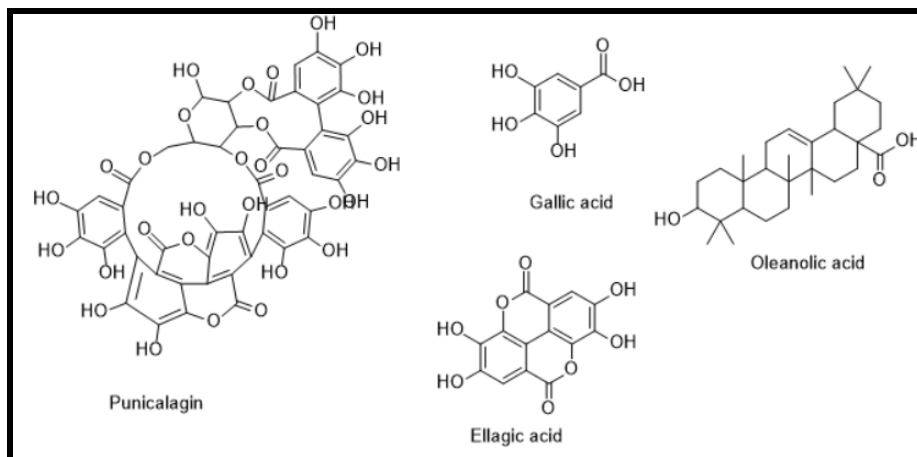


Figure-19: Active constituents of *Punica granatum* as DPP-IV inhibitors

***Calocybe indica* (Ayurvedic/Sanskrit name: Amlika; Common name: Milky white mushroom)**

Calocybe indica belongs to family Tricholomataceae, is rich in protein, flavonoids, lipid, terpenes, polyketides, fiber, carbohydrate and vitamin.⁶⁹ The ODE and LE were tested against α -amylase, α -glucosidase, and dipeptidyl peptidase-IV (DPP-IV). The oven dried extract demonstrated 50% inhibitory activity for α -amylase, α -glucosidase and DPP-IV enzyme at 62.18 μ g/ml, 47.77 μ g/ml and 91.84 μ g/ml, respectively. The lyophilized extract revealed 50% inhibitory activity for α -amylase, α -glucosidase and DPP-IV enzyme at 38.11 μ g/ml, 28.09 μ g/ml and 60.91 μ g/ml, respectively.⁷⁰

***Aronia arbutifolia* (Ayurvedic/Sanskrit name: Aronia berry; Common name: Chokeberries)**

Aronia arbutifolia (L.) Pers. *Aronia arbutifolia* (family; Rosaceae) is a genus of deciduous shrubs. It is also known as red chokeberry. Aronia berries exert lipid lowering, cardioprotective, antihypertensive, gastroprotective, anti-inflammatory, anti-oxidant and anti-diabetic activities.⁷¹ Aronia juice was fractionated by column chromatography and eluted fraction was subjected to determine DPP-IV inhibitory activity. The study also reported that cyanidin, cyaniding-3-glucoside, malvidin, luteolin, apigenin, quercetin, kaempferol, hesperetin, naringenin, eriocitrin, genistein, resveratrol, gallic acid and caffeic acid are responsible for DPP-IV inhibitory activity in Aronia juice.^{72,73}

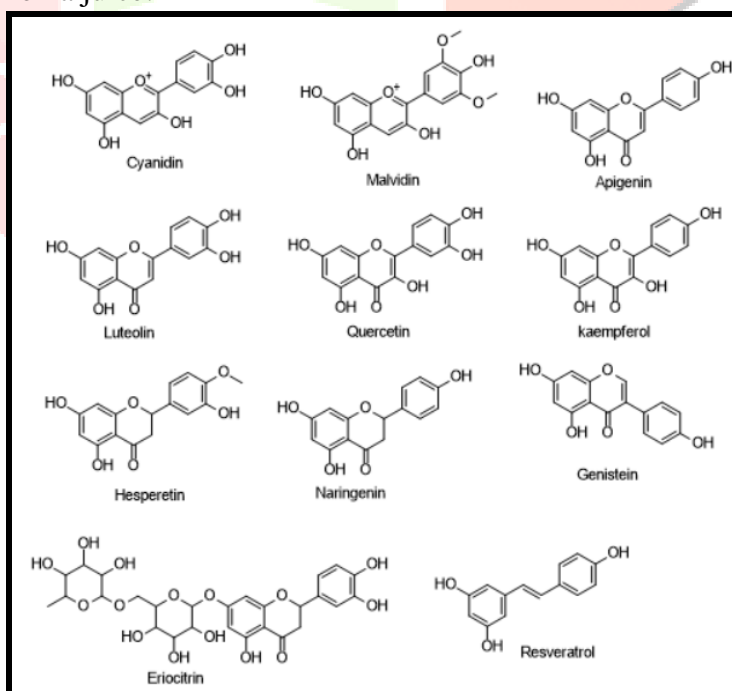


Figure-20: Active constituents of *Aronia arbutifolia* as DPP-IV inhibitors

***Urena lobata* (Ayurvedic/Sanskrit name: Sakhee Kandakeephala; Common name: Malayalam)**

Roots and leaves of *Urena lobata* have been used empirically by Nigerian people to treat Diabetes. DPP-IV inhibitory activity of *U. lobata* ethanolic extract was stronger than water extract during in-vitro testing but opposite was found with in-vivo study.⁷⁴ Yudi et al. isolate ten active substance include alkaloid, fitosterol

and flavonoid from alcoholic and water extract of *U. lobata*. DPP-IV inhibitory activity was proven by molecular docking study of leaf extract. The study shows mangiferin, stigmasterol, beta sitosterol is responsible for DPP-IV inhibitory activity.⁷⁵

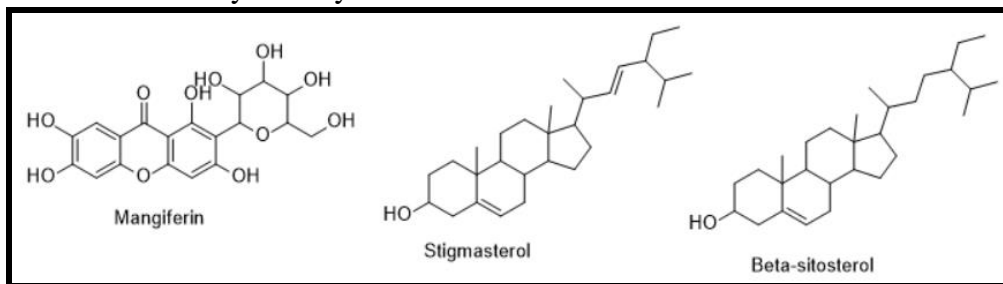


Figure-21: Active constituents of *Urena lobata* as DPP-IV inhibitors

***Pueraria tuberosa* (Ayurvedic/Sanskrit name: Vidarikand; Common name: Bilaikand, Kudzu)**

Hot water extract of roots of *P. tuberosa* showed DPP-IV inhibitory activity. The results showed a DPP-IV inhibitory activity with an IC_{50} value of 17.4 mg/mL. This value was compared with an IC_{50} value of vildagliptin (5 mg/mL) as the positive control. *In-vivo* study was carried out on normoglycemic rats by the measurement of increased plasma GLP-1 concentration via GLP-1 enzyme immunoassay kit and DPP-IV activity after a glucose load.⁷⁶ The results of the study reported inhibition of DPP-IV activity (35%), an increment of GLP-1 concentration (80%) and decrement in plasma glucose concentration in rats. Shrivastava et al. reported that puerarone and robinin are the potential phytochemicals responsible for DPP-IV inhibitory activity of roots of *P. tuberosa*.⁷⁷

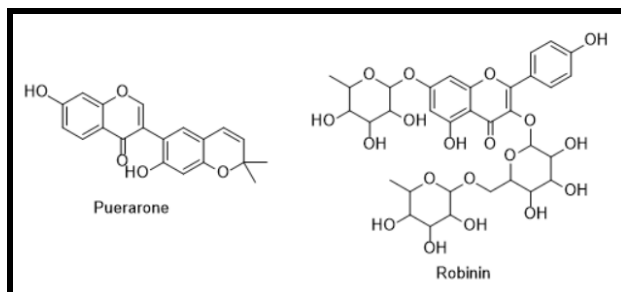


Figure-22: Active constituents of *Pueraria tuberosa* as DPP-IV inhibitors

***Antidesma madagascariense* (Ayurvedic/Sanskrit name: Antidesma bhasimbilla; Common name: Bois bigaignon batard)**

Antidesma madagascariense leaves showed antidiabetic potential via DPP-IV inhibitory activity. *In-vitro* studies of using DPP-IV inhibition assays on ethyl acetate extract of *A. madagascariense* leaves showed a DPP-IV inhibitory potential with an IC_{50} value of $79.2 \pm 2.8 \mu\text{g/mL}$.⁷⁸ Preparative scale HPLC technique was developed for isolation of active constituents responsible for DPP-IV inhibitory activity. Amentoflavone was identified as DPP-IV inhibitor with an IC_{50} value of $3.9 \pm 0.5 \mu\text{M}$. This is the first report on DPP-IV inhibition by amentoflavone.⁷⁹

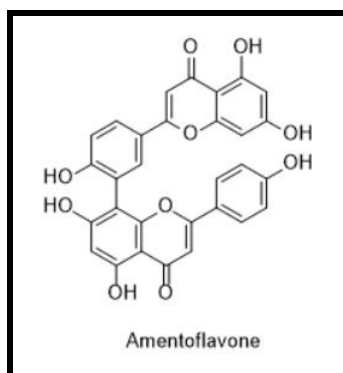


Figure-23: Active constituents of *Antidesma madagascariense* as DPP-IV inhibitors

***Avena sativa* (Ayurvedic/Sanskrit name: Avasa; Common name: Oats)**

Avena sativa known as the common oat, is a species of cereal grain grown for its seed.⁸⁰ Wang F. et al. have identified the peptides which released from oat and found out *in-vitro* inhibition activity on DPP-IV. Oats are highly responsible for inhibiting DPP-IV enzyme and showed a significant level of inhibition with IC₅₀ 0.99 mg/mL.⁸¹ Bleakley S. et al. have predicted the release of bioactive inhibitory peptides occurring from oat protein hydrolysates followed by *in silico* hydrolysis using the proteases papain and ficin. The isolated oat proteins showed DPP-IV inhibition between $3.7 \pm 3.9\%$ and $46.2 \pm 28.8\%$.⁸² Tricin is used for DPP-IV inhibitor in Oat.⁸³

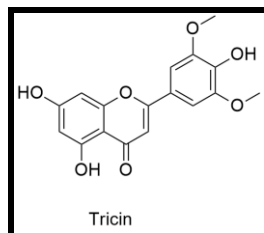


Figure-24: Active constituents of *Avena sativa* as DPP-IV inhibitors

***Eugenia Jabolana* (Ayurvedic/Sanskrit name: Jambu; Common name: Jamun)**

Eugenia jambolana Lam. or *Syzygium cumini* Skeels., a plant of the Myrtaceae family, is commonly known as jambolão in Brazil, jamun in India, and black plum in Europe. *In-vitro* assay suggested that *Eugenia Jambolana* potently inhibits DPP-IV enzyme with IC₅₀ values of 278.94 µg/mL.⁸⁴ Further, pharmacodynamic and pharmacokinetic interactions of aqueous extract of *Eugenia jambolana* seeds (400 mg/Kg) with other DPP-IV inhibitor drug sitagliptin (10mg/Kg) were studied by Vora A. et al.. These combination drug showed a better hypoglycemic action instead of individual drug.⁸⁵ Hispidulin and Petunidin is used for DPP-IV inhibitors.

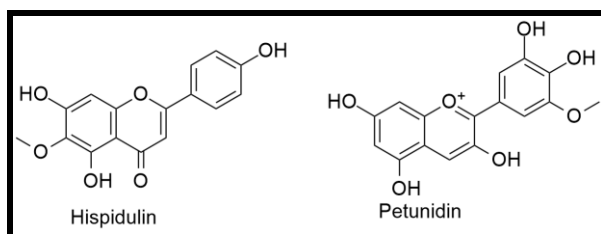


Figure-25: Active constituents of *Eugenia jabolana* as DPP-IV inhibitors

***FerulaAssa-Foetida* L. (Ayurvedic/Sanskrit name: Ramaha; Common name: Hing)**

Phytochemical analyses of *Ferula* species have confirmed the presence of sesquiterpene coumarins, sesquiterpenes, sulfides and volatile oils.⁸⁶⁻⁸⁸ Ferulic acid gives activity against DPP-IV inhibitor in *ferulaAssa-Foetida* L.⁸⁹ Yarizade A. et al. have extracted *FerulaAssa-foetida* seed using methanol, ethanol, ethanol-methanol, and water and checked for DPP-IV inhibition. All the fractions showed DPP-IV inhibition but ethanolic fraction showed the highest DPP-IV inhibition (24.5 %) ⁹⁰. Further, Nagini D. V. et al. have identified active compounds responsible for DPP-IV inhibition through GC-MS which after molecular docking studies identified Ethoxydi (tert-butyl) silane, 9,12-octadecadienoic acid, Methyl tetradecanoate and Hexadecanoic acid as most potent the standard drug saxagliptin.⁹¹

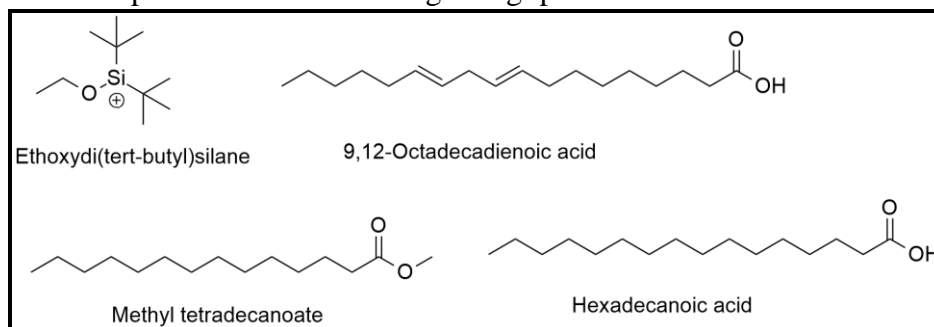


Figure-26: Active constituents of *Ferula foetida* [Asafoetida] as DPP-IV inhibitors

Fagonia cretica (Ayurvedic/Sanskrit name: Duhsparsa; Common name: Khorasanthron)

Saleem S. et al. have identified the chemical compounds responsible for DPP-IV inhibition present in *Fagonia cretica* which was previously reported natural folk medicines for the treatment of diabetes.⁹² Crude extract of the plant showed good inhibitory activity with IC₅₀ value 38.1 µg/ml. Chemical compounds which showed the inhibitory activity were isolated through bioactivity guided isolation viz. quinovic acid (IC₅₀-30.7 mM), quinovic acid-3β-O-β-D-glycopyranoside (IC₅₀-57.9 mM), quinovic acid-3β-O-β-D-glucopyranosyl-(28-1)-β-D-glucopyranosyl ester (IC₅₀- 23.5 mM), stigmasterol (IC₅₀- 4100 mM).⁹³ Further Singla R. K. et al. have carried out assessment of pharmacokinetic properties of these naturally originated potent DPP-IV inhibitors like quinovic acid, stigmasterol, quinovic acid-3-beta-D-glycopyranoside.⁹⁴

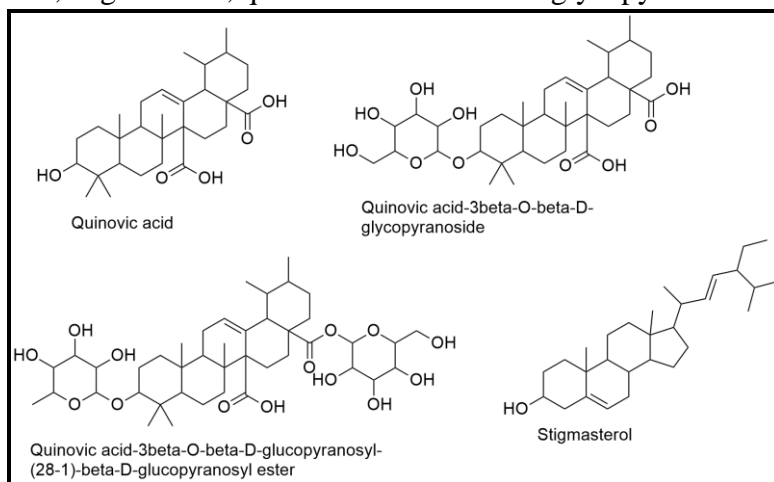


Figure-27: Active constituents of *Fagonia cretica* as DPP-IV inhibitors

Psidium guajava (Ayurvedic/Sanskrit name: Perala; Common name: Guava)

Psidium guajava L. belongs to the Myrtaceae family, have gained attention in the control of diabetes mellitus type II recently⁹⁵. Eidenberger T. et al. have investigated ethanolic extract of leaves of guava for DPP-IV inhibition activity which contain seven main flavanol-glycoside.^{96,97} The guava extract showed DPP-IV inhibition of $47.1 \pm 7.03\%$ ⁹⁶. Amongst seven main flavanol-glycoside, Peltatoside, hyperoside, isoquercitrin and guajaverin showed 5 to 10 times higher inhibitory effect than that of others. The bioactive compounds guajaverin and avicularin present in guava leaves are the potent inhibitor of glucose transporter-4 (GLUT4) and DPP-IV.⁹⁸

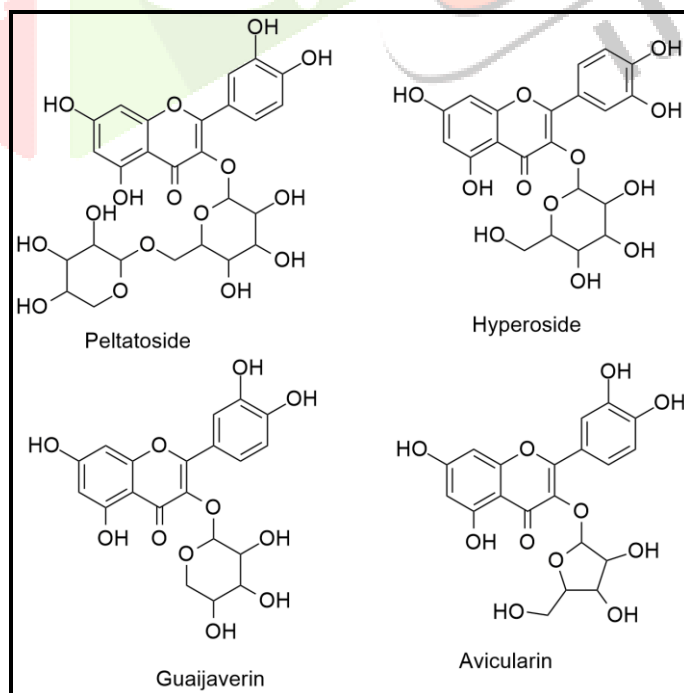


Figure-28: Active constituents of *Psidium guajava* as DPP-IV inhibitors

Morus alba (Ayurvedic/Sanskrit name: Morusalba Common name: Whitemulberry)

Morus alba, moderately fast-growing plant known as White mulberry.⁹⁹ The antidiabetic effect of the plant was identified by want H. J. et al. The leaf extract of the plant showed potent *in-vitro* α -glucosidase and DPP-IV inhibitory activities.¹⁰⁰ Agustina M. et al. investigated the effect of acid on ethanolic extract of mulberry stem bark. Apigenin, a bioactive compound of *Morus alba* stem bark extracted using ethanol with acid hydrolysis showed 23 % DPP-IV inhibition.¹⁰¹

Some DPP-IV inhibitors from natural origins were listed in **Table-1** with their active chemical constituent and testing methods. Plants with their DPP-IV inhibiting activity were listed in **Table-2** with the part of plant which can be used as DPP-IV inhibitors with other medicinal uses.

TABLE: 1 Natural DPP-IV inhibitors from different origins.

Structure sub class	Compound name	Source	Testing method	References
Alkaloids	Berberine	<i>Berberis aristata</i>	Enzymatic	48
	Withanolides	<i>Withania Somnifera</i>	Enzymatic	54
Tanin	Punicalagin, ellagic, gallic acid	<i>Punica granatum</i>	<i>In-vitro</i>	68
	Mangiferin, stigmaterol	<i>Urena lobata</i>	<i>In-vitro</i>	75
	Hispidulin, Petunidin	<i>Eugeniajam bolana</i> Lam.	<i>In vitro</i>	85
	Caffeine	<i>Camellia Senensis (Tea)</i>	<i>Ex-vivo</i>	34-35
	Arjunetin, Arjungenin, Ellagic acid and Arjunic acid	<i>Terminalia arjuna</i>	Enzymatic	52
Flavonoids	β -glucogallin	<i>Embllica officinalis</i>	Enzymatic	62
	Ellagitannins	<i>Rosa gallica</i>	<i>in-vitro</i>	63
	Ethanolic extract	<i>Bodhi leaves</i>	<i>in-vitro</i>	24
	Quercetin, carotenoids	<i>Mangifera indica</i>	<i>in-vitro</i>	19
	caesalpinianone	<i>Caesalpinia Sappan</i>	<i>in-vitro</i>	32
	α -amylase	<i>Calocybe indica</i>	<i>in-vitro</i>	70
	Puerarone and robinin	<i>Pueraria tuberosa</i>	<i>in-vivo</i>	77
	Amentoflavone	<i>Antidesma madagascariense</i>	<i>in-vitro</i>	78
	Malvidin, luteolin	<i>Aronia arbutifolia</i>	<i>in-vivo</i>	72-73
	Guaijaverin, avicularin	<i>Psidium guajava</i>	<i>in-vitro</i>	
Glycoside	Momordicin	<i>Momordica charantia.</i>	Enzymatic	6-7
	Emodin, emodic acid	<i>Cassia nigricans</i>	Enzymatic	12
	Gymnemic acid	<i>Gymnema sylvestre</i>	<i>in-vitro</i>	16
	Tricin	<i>Avena sativa</i>	Enzymatic	81
	Diosgenin	<i>Fenugreek seeds</i>	Enzymatic	57
Triterpenoids	α glucosidase	<i>Morus alba</i>	<i>in-vitro</i>	
	Eugenol, Urosolic acid	<i>Ocimum sanctum l.</i>	<i>in-vitro</i>	14
	Allicin	<i>Garlic (Allium sativum)</i>	<i>in-vitro</i>	26
	Polyphenol and flavanoids	<i>Eucalyptus globules</i>	<i>in-vitro</i>	60
Resin	Ferulic acid	<i>Ferula foetida L.</i>	<i>in-vitro</i>	89
	Guggulsterone	<i>Commiphora mukul</i>	Enzymatic	65

TABLE 2: Overview of the plants reported as a potent DPP-IV inhibition activity with their inhibition activity.

Plant name	Part of plant used	Medicinal use	DPP- IV inhibition activity (%)	References
<i>Momordica charantia</i>	fruits	Antimicrobial, antihelminthic, anticancer, antifertility, antidiabetic	53.25±0.04	9
<i>Senna migricans</i>	leaves	anticancer, antiinflammatory, antidiabetic, antioxidant	57.0± 1.91	12
<i>Ocimum sanctum L.</i>	leaves	Cough, cold, abdominal pain, antifungal, hepatoprotective, cardioprotective, antidiabetic	66.81±0.05	15
<i>Gymnema sylvestre</i>	leaves	Urinary diseases, antidiabetic, antioxidant	60.21 ± 0.35	17
<i>Mangifera indica</i>	leaves	Anti- lipid, peroxidation, cardiogenic, hypotension, antidiabetes	68.22 ± 1.14	20
<i>Fenugreek</i>	seed	Antiulcer, antifertility, antidiabetic, Immunomodulatory effect	71.29 ±0.33	57
<i>Bodhi leaves</i>	leaves	Antibacterial, gonorrhoea, skin diseases, antidiabetes	68.98 ± 1.95	25
<i>Garlic</i>	fruits	hepatoprotective, antiinflammatory, antidiabetic, antioxidant, anthelmintic	50.0±0.01	27
<i>Caeslpinia sappan</i>	leaves	Anticancer, anti diarrhoeal, antifungal	84.25±0.01	32
<i>Camellia sinensis</i>	plants	antioxidant, hypoglycemic agents	63.2 ± 0.01	39
<i>Berberine</i>	plants	Antitumor, antimicrobial, antioxidant	79.2 ± 0.18	50
<i>Terminalia arjuna</i>	bark	Cardio protective, hypotensive, hypolipidemic, wound healing activity, antidiabetes	83.39 ± 7.58	52
<i>Withania somnifera</i>	root powder	Arthritis, anxiety, trouble sleeping, antidiabetes	90.35 ± 0.85	56
<i>Fenugreek</i>	seeds	diabetes, poor appetite, inflammation, digestive problems and menopausal symptoms	71.29 ± 0.33	59
<i>Eucalyptus globules</i>	bark and leaves	diabetes, cough, cold	63.2 ± 0.1	61
<i>Emblica officinalis</i>	fruit	antioxidant, immune modulatory, antipyretic, analgesic, cytoprotective, anti ulser	85.95±7.16	62
<i>Rosa gallica</i>	flower	colds, bronchial infections, gastritis, diarrhoea, depression body decrease the blood sugar level	69.01 ± 1.3	63
<i>Commiphora mukul</i>	leaves	anti-diabetic	92.97±8.45	65
<i>Punica granatum</i>	fruit	anti-T2DM and anti-cancer	47.1 ± 1.5	67
<i>Calocybe indica</i>	fruit	anti-diabetic	62.18 ± 1.2	70
<i>Aronia arbutifolia</i>	fruit	cardioprotective, antihypertensive, gastroprotective, anti-inflammatory	28.18 ± 0.92	72
<i>Urena lobata</i>	leaves	stomach-ache, diarrhoea and dysentery	IC ₅₀ value 57.44 µg/mL	75
<i>Pueraria tuberosa</i>	root	anti-diabetic	IC ₅₀ value 17.4	77

			mg/mL	
<i>Antidesma madagascariense</i>	leaves	anti-diabetic	79.2±2.8	79
<i>Avena sativa</i>	cereals	anti-diabetic	3.7 ± 3.9	82
<i>Eugenia Jabolana</i>	fruits	anti-diabetic	IC ₅₀ value of 278.94 µg/mL	84
<i>Ferula Assa-Foetida</i> L.	Seeds (species)	Antispasmodics, anti-diabetic	Inhibition 24.5	90
<i>Fagoniacretica</i>	leaves	anti-diabetic	IC ₅₀ value of 38.1 µg/mL	93
<i>Psidium guajava</i>	fruit	anti-diabetic	47.1 ± 7.03	96
<i>Morus alba</i>	Seeds	anti-diabetic	Inhibition 23	101

Conclusion: Traditional plants are a source of bioactive compounds with diverse scaffolds, well known to treat and manage metabolic disorders like diabetes mellitus. In recent years, the enzyme DPP-IV has become an important drug target for diabetes therapy and DPP-IV inhibitors become a popular remedy for it. However, some chemically synthesized compounds are commercially available. But for the finding of the potential or more advance DPP-IV inhibitors from natural source now a day's molecular docking study is used more prominently. Although, nature is a rich source of medicinal plants that have been used to treat diabetes mellitus from ages. Therefore, the discovery of natural DPP-IV inhibitors may suggest a new chance or a new idea for developing newer medications. The present review will remarkably increase the research to find new DPP-IV inhibitors from natural sources using various modern tools. It is believed that this review will increase the attention of the research group for development of newer natural derivatives which provides better, safe and efficacious DPP-IV inhibitor.

Conflict of Interest

Authors state no conflict of interest

References:

1. World Health Organization, *Diabetes: Key Facts*, World Health Organization, Geneva, Switzerland, 2011.
2. Zimmet P., Alberti K. G., Shaw J.: Global and societal implications of the diabetes epidemic, **Nature**, 2001, 414(6865), 782-787.
3. American Diabetes Association, 2009. Standards of medical care in diabetes—2009. **Diabetes care**, 32(Suppl 1), p.S13.
4. Salvo F., Moore N., Arnaud M., Robinson P., Raschi E., De Ponti F., Bégaud B. and Pariente A.: Addition of dipeptidyl peptidase-4 inhibitors to sulphonylureas and risk of hypoglycaemia: systematic review and meta-analysis, **BMJ**, 2016, 353.
5. Demuth H. U., McIntosh C.H. and Pederson R. A., Type 2 diabetes—therapy with dipeptidyl peptidase IV inhibitors, **Biochim. Biophys. Acta. Proteins. Proteom BBA-proteom**, 2005, 1751(1), 33-44.
6. Paul A. and Raychaudhuri S. S., Medicinal uses and molecular identification of two *Momordica charantia* varieties-a review, **Electron J Biol**, 2010, 6(2), 43-51.
7. Singh J., Cumming E., Manoharan G., Kalasz H. and Adeghate E., Suppl 2: Medicinal chemistry of the anti-diabetic effects of *Momordica charantia*: active constituents and modes of actions, **Open J Med Chem journal**, 2011, 5, 70.
8. Bhat G. A., Khan H. A., Alhomida A. S., Sharma P., Singh R. and Paray B. A., GLP-I secretion in healthy and diabetic Wistar rats in response to aqueous extract of *Momordica charantia*, **BMC Complement Altern Med**, 2018, 18(1), 162.
9. Singh A. K., Jatwa R. and Joshi J., Cytoprotective and dipeptidyl peptidase IV (DPP-IV/CD26) inhibitory roles of *Ocimum sanctum* and *Momordica charantia* extract, **Health**, 2014, 20, p.22. doi: 10.1.1.989.5090.

10. Ibrahim M. A. and Md S. I., Anti-Diabetic Effects of Plants Belonging to the Genus Senna: Pharmacology, Mechanism of Action and Phytochemistry, Phytotherapy in the management of Diabetes and Hypertension, Eddouks (ed.) **Bentham Science Publishers**, 2016, 2, pp.138-153.
11. Saidu Y., Muhammad S. A., Bilbis L. S. and Sani B. M., Inhibitory activity of fractions of *Senna nigricans* toward protein tyrosine phosphatase 1B and dipeptidyl peptidase IV, **J Med Plant Res**, 2016, 10(18), 242-247.
12. Saidu Y., Muhammad S. A., Abbas A. Y., Onu A., Tsado I. M. and Muhammad L., In vitro screening for protein tyrosine phosphatase 1B and dipeptidyl peptidase IV inhibitors from selected Nigerian medicinal plants, **J Intercult Ethnopharmacol**, 2017, 6(2), p.154.
13. Inbaraj S. D. and Muniappan M., Correlation between the In-Vitro and In-Vivo Antihyperglycemic Effect of *Ocimum Sanctum*, *Trigonella Foenum Graecum* and *Curcuma Longa*, **Pharmacogn J**, 2020, 12(2).
14. Somasundaram G., Manimekalai K., Salwe K. J. and Pandiamunian J., Evaluation of the antidiabetic effect of *Ocimum sanctum* in type 2 diabetic patients, **Int J Life Sci Pharma Res**, 2012, 5, 75-81.
15. Singh A. K., Jatwa R. and Joshi J., Cytoprotective and dipeptidyl peptidase IV (DPP-IV/CD26) inhibitory roles of *Ocimum sanctum* and *Momordica charantia* extract, **Health**, 2014, 20, p.22.
16. Ahmed A. B. A., Rao A. S. and Rao M. V., In vitro callus and in vivo leaf extract of *Gymnema sylvestre* stimulate β -cells regeneration and anti-diabetic activity in Wistar rats, **Phytomedicine**, 2010, 17(13), 1033-1039.
17. Laha S. and Paul S., *Gymnema sylvestre* (Gurmar): A Potent Herb with Anti-Diabetic and Antioxidant Potential, **Pharmacogn J**, 2019, 11(2).
18. Arun L. B., Arunachalam, A. M., Arunachalam K. D., Annamalai S. K. and Kumar K. A.: In vivo anti-ulcer, anti-stress, anti-allergic, and functional properties of *Gymnemic Acid* Isolated from *Gymnema sylvestre* R Br. **BMC Complement Altern Med**, 2014, 14(1), 70.
19. Andreu G. P., Delgado R., Velho J. A., Curti C. and Vercesi A. E., Iron complexing activity of mangiferin, a naturally occurring glucosylxanthone, inhibits mitochondrial lipid peroxidation induced by Fe^{2+} -citrate, **Eur J Pharmacol**, 2005, 513(1-2), 47-55.
20. Suman R. K., Mohanty I. R., Maheshwari U., Borde M. K. and Deshmukh Y. A., Natural dipeptidyl peptidase-IV inhibitor mangiferin mitigates diabetes-and metabolic syndrome-induced changes in experimental rats, **Diabetes Metab Syndr Obes**, 2016, 9, 261.
21. Chan E. W. C., Tangah J., Inoue T., Kainuma M., Baba K., Oshiro N., Kezuka M. and Kimura N., Botany, uses, chemistry and pharmacology of *Ficus microcarpa*: a short review, **Syst Rev Pharm**, 2017, 8(1), 103.
22. Kalita P., An overview on *mangifera indica*: importance and its various pharmacological action, **PharmaTutor**, 2014, 2(12), 72-76.
23. Setyaningsih E. P., Saputri F. C. and Mun'im A., The antidiabetic effectivity of Indonesian plants extracts via DPP-IV inhibitory mechanism, **J Young Pharm**, 2019, 11(2), 161.
24. Chandrasekar S. B., Bhanumathy M., Pawar A. T. and Somasundaram T., Phytopharmacology of *Ficus religiosa*, **Pharmacogn Rev**, 2010, 4(8), p.195.
25. Riyanti S., Suganda A. G. and Sukandar E. Y., Dipeptidyl peptidase-IV inhibitory activity of some Indonesian medicinal plants, **Asian J Pharm Clin Res**, 2016, 9(2), 375-377.
26. Sheela C. G. and Augusti K. T., Antidiabetic effects of S-allyl cysteine sulphoxide isolated from garlic *Allium sativum* Linn, **Indian J Exp Biol**, 1992, 30(6), 523-526.
27. El-Demerdash F. M., Yousef M. I. and Abou El-Naga N. I., Biochemical study on the hypoglycemic effects of onion and garlic in alloxan-induced diabetic rats, **Food Chem Toxicol**, 2005, 43(1), 57-63.
28. Eidi A., Eidi M. and Esmaeili E., Antidiabetic effect of garlic (*Allium sativum* L.) in normal and streptozotocin-induced diabetic rats, **Phytomedicine**, 2006, 13(9-10), 624-629.
29. Igho O., Kang H. S., Rachel P. and Edzard E., The use of *Garcinia* extract (hydroxycitric acid) as a weight loss supplement: a systematic review and meta-analysis of randomised clinical trials, **J Obes**, 2011.
30. Sher A., Fakhar-ul-Mahmood M., Shah S. N., Bukhsh S. and Murtaza G., Effect of garlic extract on blood glucose level and lipid profile in normal and alloxan diabetic rabbits, **Adv clin exp med**, 2012, 21(6), 705.

31. Kalhotra P., Chittepu V. C. S. R., Osorio-Revilla G., and Gallardo-Velazquez T.: Phytochemicals in Garlic Extract Inhibit Therapeutic Enzyme DPP-4 and Induce Skeletal Muscle Cell Proliferation: A Possible Mechanism of Action to Benefit the Treatment of Diabetes Mellitus, *Biomolecules*, 2020, 10(2)305.
32. Setiawan H., Angela I. L., Rohmah N., Wijaya O., Mun'im A.: Application of Natural Deep Eutectic Solvents (NADES) for Sappan Wood (*Caesalpinia sappan* L.) extraction to test for inhibition of DPP-IV activity, *J Res Pharm*, 2020, 24(3) 380-388.
33. Nirmal N. P., Rajput M. S., Prasad R. G., & Ahmad M.:Brazilin from *Caesalpinia sappan* heartwood and its pharmacological activities: A review,Asian Pac. *J. Trop. Med.*, 2015, 8(6)421-430.
34. Wang Y., Yang Z. and Wei X.: Sugar compositions, α -glucosidase inhibitory and amylase inhibitory activities of polysaccharides from leaves and flowers of *Camellia sinensis* obtained by different extraction methods, *Int. J. Biol. Macromol*, 2010,47(4).534-539.
35. Wang Y., Shao S., Xu, P., Chen H., Lin-Shiau S. Y., Deng Y. T. and Lin J. K.: Fermentation process enhanced production and bioactivities of oolong tea polysaccharides, *Food Res. Int.*, 2012, 46(1), 158-166.
36. Anderson R. A. and Polansky M. M.: Tea enhances insulin activity, *J. Agric. Food Chem*, 2002, 50(24) 7182–7186.
37. Dembinska-Kiec A., Mykkänen O., Kiec-Wilk B., and Mykkänen, H., Antioxidant phytochemicals against type 2 diabetes, *Br. J. Nutr.*, 2008, 99(E-S1), 109-117.
38. Ma J., Li Z., Xing S., Ho W. T., Fu X. and Zhao Z. J.: Tea contains potent inhibitors of tyrosine phosphatase PTP1B, *Biochem. Biophys. Res. Commun*, 2011, 407(1)98-102.
39. Wolfram S., Raederstorff D., Preller M., Wang Y., Teixeira S. R., Riegger C. and Weber P.: Epigallocatechin gallate supplementation alleviates diabetes in rodents, *J Nutr*, 2006 , 136(10), 2512-8.
40. Ekayanti M., Sauriasari R. and Elya B.: Dipeptidyl peptidase IV inhibitory activity of fraction from white tea ethanolic extract (*Camellia sinensis* (L.) Kuntze) ex vivo, *J Pharmacogn.*, 2018, 10(1)190-193.
41. Lu Y., Lu P., Wang Y., Fang X., Wu J. and Wang X.: A Novel Dipeptidyl Peptidase IV Inhibitory Tea Peptide Improves Pancreatic β -Cell Function and Reduces α -Cell Proliferation in Streptozotocin-Induced Diabetic Mice, *Int. J. Mol. Sci.*, 2019, 20(2), 322.
42. Ortsäter H., Grankvist N., Wolfram S., Kuehn N., Sjöholm Å.: Diet supplementation with green tea extract epigallocatechin gallate prevents progression to glucose intolerance in db/db mice, *Nutr. Metab.*, 2012, 9(1), 1-10.
43. Meng F., Abedini A., Plesner A., Verchere C.B. and Raleigh D.P.: The flavanol (–)-epigallocatechin 3-gallate inhibits amyloid formation by islet amyloid polypeptide, disaggregates amyloid fibrils, and protects cultured cells against IAPP-induced toxicity, *Biochemistry*, 2010, 49(37), 8127-8133.
44. Meng F., Abedini A., Plesner A., Verchere C.B. and Raleigh D.P.: The flavanol (–)-epigallocatechin 3-gallate inhibits amyloid formation by islet amyloid polypeptide, disaggregates amyloid fibrils, and protects cultured cells against IAPP-induced toxicity, *Biochemistry*, 2010, 49(37), 8127-8133.
45. Hwang J.T., Park I.J., Shin J.I., Lee Y.K., Lee S.K., Baik H.W., Ha J. and Park O.J.: Genistein, EGCG, and capsaicin inhibit adipocyte differentiation process via activating AMP-activated protein kinase, *Biochem. Biophys. Res. Commun.*, 2005, 338(2), 694-699.
46. Hou H., Wang Y., Li C., Wang J. and Cao Y.: Dipeptidyl Peptidase-4 Is a Target Protein of Epigallocatechin-3-Gallate, *Biomed Res. Int.*, 2020.
47. Kirthikar K. R. and Basu B. D. I., Indian Medicinal Plants, **International Book Distributors**, Dehradun, 2005, 1064-1073.
48. Shahid M., Rahim T., Shahzad A., Latif T.A., Fatma T., Rashid M., Raza A. and Mustafa S. Ethnobotanical studies on *Berberis aristata* DC. Root extracts, *Afr. J. Biotechnol*, 2009, 8(4), 45-58.
49. Mittal M., Juyal V. and Singh A.: Phytochemical, antidiabetic, and cytoprotective properties of *Berberis aristata* DC. **Root extracts. Pharm. Crop**, 2012, 3(1), 64-68.
50. Chakrabarti R., Bhavtaran S., Narendra P., Varghese N., Vanchhawng L., Mohamed Sham Shihabudeen, H. and Thirumurgan, K., Dipeptidyl peptidase-IV inhibitory activity of *Berberis aristata*, *J Nat Prod*, 2011, 4, 158-163.
51. Mandal A., Das K. and Nandi D. K.: *In vitro* bioactivity study of bark extract of *Terminalia arjuna* on probiotics, commercially available probiotic formulation. *Int J Phytopharmacol*, 2010, 1(2), 109–113.

52. Borde M. K., Mohanty I. R., Suman R. K. and Deshmukh Y. A.: Dipeptidyl peptidase-IV inhibitory activities of medicinal plants: Terminalia arjuna, Commiphora mukul, Gymnema sylvestre, Morinda citrifolia, Emblica officinalis, **Asian J Pharm Clin Res**, 2016, 9(3), 180-182.
53. Mohanty I.R., Borde M, Selvaa Kumar C, Maheshwari U,: Dipeptidyl peptidase IV Inhibitory activity of Terminalia arjuna attributes to its cardioprotective effects in experimental diabetes: In silico, in vitro and in vivo analyses , **phytomedicine**, 2019, 57, 158-165.
54. Andallu B. and Radhika B., Hypoglycemic, diuretic and hypocholesterolemic effect of winter cherry (Withania somnifera, Dunal) root, **Indian J Exp Biol**, 2000, 38, 607-613.
55. Singh G., Sharma P.K., Dudhe R. and Singh S.: Biological activities of Withania somnifera, **Ann Biol Res**, 2010, 1(3), 56-63.
56. Kempegowda P. K., Zameer F. and Murari S. K.: Delineating antidiabetic proficiency of catechin from Withania somnifera and its Inhibitory action on dipeptidyl peptidase-4 (DPP-4), **Biomed Res**, 2018, 29(16), 3192-3200.
57. Khosla P., Gupta D.D. and Nagpal R.K.: Effect of Trigonella foenum graecum (Fenugreek) on blood glucose in normal and diabetic rats. **Indian J. Physiol. Pharmacol**, 1995, 39, 173-173.
58. Bordia A., Verma S. K. and Srivastava K. C.: Effect of ginger (Zingiber officinale Rosc.) and fenugreek (Trigonella foenumgraecum L.) on blood lipids, blood sugar and platelet aggregation in patients with coronary artery disease. **Prostaglandins Leukot. Essent. Fatty Acids**, 1997, 56(5), 379-384.
59. Riyanti S., Suganda A. G. and Sukandar E. Y.: Dipeptidyl peptidase-IV inhibitory activity of some Indonesian medicinal plants. **Asian J Pharm Clin Res**, 2016, 9(2), 375-377.
60. Dey B., Mitra A., Katakam P. and Singla R. K.: Exploration of natural enzyme inhibitors with hypoglycemic potentials amongst Eucalyptus Spp. by in vitro assays. **World J. Diabetes**, 2014, 5(2), 209-218.
61. Kato E. K. and Kawabata J.: Macrocarpal C isolated from Eucalyptus globulus inhibits dipeptidyl peptidase 4 in an aggregated form, **J of enz inhibition and med chem**, 2018, 33(1), 106-109.
62. Majeed M. S., Mundkur L., Nagabhushanam K., Arumugam S., Beede K. and Ali F.: Standardized Emblica officinalis fruit extract inhibited the activities of α -amylase, α -glucosidase, and dipeptidyl peptidase-4 and displayed antioxidant potential, **J of the Sci of Food and Agri**, 2020, 100(2), 509-516.
63. Kato E. S., Uenishi Y. A., Inagaki Y. B., Kurokawa M. A. and Kawabata J.: Isolation of rugosin A, B and related compounds as dipeptidyl peptidase-IV inhibitors from rose bud extract powder, **J Biosci, biotechno, and biochem**, 2016, 80(11), 2087-2092.
64. Sharma D. K. and Kumar S. D.: DPP-IV Inhibitors from natural sources: An alternative approach for treatment and management of diabetes, **Ind J of Natural Products and Res**, 2019, 10(4), 227-237.
65. Sharma B. S., Salunke R. S., Majumder C. P. and Roy P. G.: Effects of guggulsterone isolated from Commiphora mukul in high fat diet induced diabetic rats, **Food and chemical toxicology**, 2009, 47(10), 2631-2639.
66. Sharma D. K. and Kumar S. D.: DPP-IV Inhibitors from natural sources: An alternative approach for treatment and management of diabetes, **Ind J of Natural Products and Res**, 2019, 10(4), 227-237.
67. Malik L. A., Ad'hiah A. H. and Aziz G. M.: Phytochemical content and the potential of Punica granatum peel extracts as radical scavengers and dipeptidyl peptidase-4 inhibitors, **J of Biotech Res Cen**, 2019, 13(1), 5-11.
68. Gharib E. S. and Kouhsari, S.M.: Study of the antidiabetic activity of Punica granatum L. fruits aqueous extract on the Alloxan-diabetic Wistar rats, **Iranian J of Pharma Res (IJPR)**, 2019, 18(1), 358-372.
69. Amit R. and Pushpa P.: Assessment of antihyperglycemic potential of lyophilized and oven-dried extract of Calocybe indica in experimentally streptozotocin-nicotinamide induced diabetic rats, **Int J of Med Res & Health Sci**, 2016, 5(4), 82-88.
70. Faden A. A.: Evaluation of antibacterial activities of aqueous and methanolic extracts of areca catechu against some opportunistic oral bacteria, **Biosci Biotech Res Asia**, 2018, 15(3), 655-659.
71. Banjari I., Misir A. S., Jokić S., Molnar M., De Zoysa H. K. S. and Waisundara V. Y.: Antidiabetic effects of Aronia melanocarpa and its other therapeutic properties, **Frontiers in nutrition**, 2017, 4, 53-62.

72. Naruszewicz M., Łaniewska, I., Millo B. and Dłużniewski M.: Combination therapy of statin with flavonoids rich extract from chokeberry fruits enhanced reduction in cardiovascular risk markers in patients after myocardial infraction (MI), **Atherosclerosis**, 2007, 194(2), e179-e184.
73. Ohgami K., Ilieva I., Shiratori K., Koyama Y., Jin X. H., Yoshida K., Kase S., Kitaichi N., Suzuki Y., Tanaka T. and Ohno S.: Anti-inflammatory effects of aronia extract on rat endotoxin-induced uveitis, **Investigative Ophthalmology & Visual Sci**, 2005, 46(1), 275-281.
74. Nomo Y., Soeatmadji D., Sumitro, S. B. and Widodo M. A.: Inhibitory activity of Urena lobata leaf extract on dipeptidyl peptidase-4 (DPP-IV): Is it different in vitro and in vivo? **Medicinal Plants-Int J of Phytomedicines and Related Industries**, 2018, 10(2), 99-105.
75. Purnomo Y., Soeatmadji D. W., Sumitro S. B. and Widod, M. A.: Anti-diabetic potential of Urena lobata leaf extract through inhibition of dipeptidyl peptidase IV activity, **Asian Pacific J of Tropical Biomedicine**, 2015, 5(8), 645-649.
76. Srivastava S., Koley T. K., Singh S. K. and Tripathi Y. B.: The tuber extract of pueraria tuberosa Linn. Competitively inhibits DPP-IV activity in normoglycemic rats, **Notes**, 2015, 20(90), 4-9.
77. Srivastava S., Shree P. and Tripathi Y. B.: Active phytochemicals of Pueraria tuberosa for DPP-IV inhibition: in silico and experimental approach, **J of Diabetes & Metabolic Disorders**, 2017, 16(1), 46-55.
78. Wasana K. G., Attanayake A. P., Jayatilaka K. A., and Weerarathna T. P.: Natural drug leads as novel DPP-IV inhibitors targeting the management of type 2 diabetes mellitus. **J of Complementary Medicine Res**, 2020, 11(1), 43-53.
79. Beidokhti M. N., Lobbens E. S., Rasoavaivo P., Staerk D. and Jäger A. K.: Investigation of medicinal plants from Madagascar against DPP-IV linked to type 2 diabetes, **South African J of Botany**, 2018, 115, 113-119.
80. Sharma D., Kumar S., Kumar S. and Kumar D., DPP-IV Inhibitors from natural sources: An alternative approach for treatment and management of diabetes, **Indian J of Natural Products and Res**, 2019, 10(4), 227-237.
81. Wang F, Yu G, Zhang Y, Zhang B, and Fan J, Dipeptidyl peptidase IV inhibitory peptides derived from oat (*Avena sativa* L.), buckwheat (*Fagopyrum esculentum*), and highland barley (*Hordeum vulgare trifurcatum* (L.) Trofim) proteins, **J Agric Food Chem**, 2015, 63(43), 9543–9549.
82. Bleakley S., Hayes M., Shea N. O., Gallagher E. And Lafarga T., Predicted Release and Analysis of Novel ACE-I, Renin, and DPP-IV Inhibitory Peptides from Common Oat (*Avena sativa*) Protein Hydrolysates Using in Silico Analysis, **Foods**, 2017, 6, 108-112.
83. Czerwiński J., Bartnikowska E., Leontowicz H., Lange E., Leontowicz M., Katrich E., Trakhtenberg S., Gorinstein S., Oat (*Avena sativa* L.) and amaranth (*Amaranthus hypochondriacus*) meals positively affect plasma lipid profile in rats fed cholesterol-containing diets, **J Nutr Biochem**, 2004, 15(10), 622-629.
84. Ayyanar M., Subash-Babu P., Ignacimuthu S., *Syzygiumcumini* (L.) Skeels, a novel therapeutic agent for diabetes: Folk medicinal and pharmacological evidences, **Complementary Therapies in Medicine**, 2013, 21, 232-243.
85. Kosaraju J., Dubala A., Chinni S., Khatwal R. B., Kumar M. N. S., et al., A molecular connection of *Pterocarpus marsupium*, *Eugenia jambolana* and *Gymnemasylvestre* with dipeptidyl peptidase-4 in the treatment of diabetes, **Pharm Biol**, 2014, 52(2), 268-271.
86. Vora, A., Varghese, A., Kachwala, Y., Bhaskar, M., Laddha, A., Jamal, A. and Yadav, P., 2019. *Eugenia jambolana* extract reduces the systemic exposure of Sitagliptin and improves conditions associated with diabetes: a pharmacokinetic and a pharmacodynamic herb-drug interaction study. **J of Trad and Comp Medi**, 9(4), 364-371.
87. Appendino G., Tagliapietra S., Nona G. M., Jakupovic J., Sesquiterpene coumarin ethers from *Asafetida*, **Phytochemistry**, 1994, 35(1), 183-186.
88. Fattahian K., Shokoohinia Y., Ghannadi A., Behbahani M., Shahnoush A., Anti-viral evaluation of sesquiterpene coumarins from *Ferulaassa-foetida* against HSV-1, **Iranian J Pharm Res**, 2013, 5(7), 78-87.
89. Al-Hazimi H. M. G., Terpenoids and a coumarin from *Ferulasinaica*. **Phytochemistry**. 1986, 25(10), 2417-2419.

90. Bahramia G., Soltanib R., Sajjadie S. E., Kananid M. R., Naderie R., Ghiasvandf N., Shokoohiniag Y. H., Essential Oil Composition of *Ferula Assa-Foetida* L. Fruits from Western Iran, **J Reports in Pharm Sci**, 2013, 2(2), 90-97.
91. Yarizade, A., Kumleh, H. H., and Niazi, A. L. I., In vitro antidiabetic effects of *ferula foetida* extracts through dipeptidyl peptidase IV and α -glucosidase inhibitory activity. **In Vitro**, 2017, 10(5), 104-116.
92. Nagini D. V., Krishna M. S. R. and Karthikeyan S., Identification of Novel Dipeptidyl Peptidase-IV Inhibitors from *Ferula asafoetida* through GC-MS and Molecular Docking Studies. **Res. J. Pharm. Tech**, 2020, 13(11), 5072-5076.
93. Marles R. J. and Farnsworth N. R., Antidiabetic plants and their active constituents. **Phytomedicine**, 1995, 2(2), 137-189.
94. Saleem S., Jafri L., ul Haq I., Chang L. C., Calderwood D., Green B. D. and Mirza B., Plants *Fagonia cretica* L. and *Hedera nepalensis* K. Koch contain natural compounds with potent dipeptidyl peptidase-4 (DPP-4) inhibitory activity. **J ethnopharma**, 2014, 156, 26-32.
95. Singla R. K. and Shen B., In Silico ADMET evaluation of natural DPP-IV inhibitors for rational drug design against diabetes. **Current Drug Metabolism**, 2020, 21(10), 768-777.
96. Matsuda H, Morikawa T, Yoshikawa M. Antidiabetogenic constituents from several natural medicines. **Pure Appl Chem** 2002, 74, 1301-8.
97. Eidenberger T., Selg M. and Krennhuber K., Inhibition of dipeptidyl peptidase activity by flavonol glycosides of guava (*Psidium guajava* L.): A key to the beneficial effects of guava in type II diabetes mellitus. **Fitoterapia**, 2013, 89,74-79.
98. Ojewole J. A. O., Hypoglycemic and hypotensive effects of *Psidium guajava* Linn. (Myrtaceae) leaf aqueous extract. **Methods and findings in exp and clin pharmacology**, 2005, 27(10), 689-696.
99. Kumar M., Tomar M., Amarowicz R., Saurabh V., Nair M. S., Maheshwari C., Sasi M., Prajapati U., Hasan M., Singh S. and Changan S., Guava (*Psidium guajava* L.) leaves: nutritional composition, phytochemical profile, and health-promoting bioactivities. **Foods**, 2021, 10(4), 752.
100. Wang H. J. and Chiang B. H., Anti-diabetic effect of a traditional Chinese medicine formula. **Food & Function**, 2012, 3(11), 1161-1169.
101. Agusfina M., Saputri F. C., Sakti A. S. and Mun'im A., Difference of Acidic Adding Effect in Ethanol Extraction of White Mulberry Stem Bark (*Morus alba*) and DPP-4 Inhibiting Activity Screening for Identifying its Antidiabetic Potential. **Pharmacognosy Journal**, 2019, 11(4), 54-68.

