



# PREPARE AND CHARACTERIZE SILK FIBRION NANOPARTICLES LOADED WITH MESALAMINE

<sup>1</sup>Sunil Bharti, <sup>2</sup>Kaushalya Bains, <sup>3</sup>Raveena Devi, <sup>4</sup>Dr. Sakshi

<sup>1</sup>Student, <sup>2</sup>Guide, <sup>3</sup> Teacher, <sup>4</sup>HOD

<sup>1</sup>Himalyan Institute Of Pharmacy

<sup>1</sup>Himalyan Group Of Professional Institute, Kala-Amb, H.P. India

## ABSTRACT

A major silk protein called fibroin consumes the perfect characteristics to be used such as a biomaterial for medication supply. Recently, here has been a proportion of research done on the creation of fibroin nanoparticle (FNPs) aimed at various medicinal purposes. FNP can encapsulate a variation of therapeutic substances, excluding tiny and large molecule, protein, enzyme, vaccine, and hereditary material because of their adaptability and chemical modifiabilities. FNPs can also be supplied non-parenteral as well as parenteral. The fundamental facts about the origin and properties of silk and fibroin are briefly summarized in this review, which is followed by the most recent information on the techniques used to prepare and characterize FNPs. Additionally, their uses in medicine as a method of drug delivery are thoroughly investigated using a variety of administration methods, including parenteral, oral, trans-dermal, ophthalmic, orthopedic, and respirational. To end with, the issues, potential fixes, and prospects for these systems' future are examined.

**Keywords :- Nanoparticles, Silk Fibrion, Mesalamine, Administration**

## 1.Introduction

The effectiveness of a drug is determined by both its healing efficacy and, extra crucially, her adverse effect in the current era of modified medicine. These adverse effects, which can range in severity from moderate to severe, are frequently what prevents people from using drugs more frequently. The enormous bodily distribution of pharmaceutical substances is the primary cause of the majority, if not all, adverse consequences. Only a small portion of the provided dose for conservative dosage form (such as tablet, capsule, and intra-venous injection) reach the target place, but the rest of the drug molecule disperse during the complete body depending on this one physicochemical qualities. To exercise its therapeutic impact, for example, fewer than 0.5 percent of a paclitaxel intra-venous vaccination primary dose is available in the vicinity indoors the lung cancer. Therefore, it stands essential to create a drug distribution structure that delivers the medication indirectly to the target areas, enhancing its activity though minimizing any negative side effect.

Artificial then ordinary polymer based NPs can compress the drug object addicted to their essential (Nano capsules) or spread the drug calmly throughout their medium (Nano spheres) by using polymers as a carrier. Numerous synthetic polymers have been employed, including polyesters, polyanhydrides, and polyphosphazenes. First a few number of candidates have received FDA approval due to their bio-incompatibility, such by way of poly (lactic-co-glycolic acids). However, the lactic acids produced by the

decomposition of PLGA NPs lowers the pH in the area, which denatures the pharmaceuticals that are entrapped, particularly acid-labile proteins, and lessens their therapeutic effects. As a result, some natural polymers with high biocompatibility and biodegradability, excluding chitosan, alginates, gelatin, and fibroin; Aram wit, have been used as substitutes. Among these, fibroin is a polymer that has received FDA approval and has a long past of use in medicinal settings for things like suture, tissue renaissance, coatings for device, and drug delivery system. Fibroin, which is primarily produced from the cocoons of farmed silkworms (Bombaymore), has attracted growing interest for its superior mechanical qualities, high biocompatibility, biodegradability, affordability, and adaptability in preparation. These qualities make fibroin nanoparticles (FNPs) formulation desirable. Furthermore, because of its amphoteric qualities, fibroin can be cross-linked either by himself or with extracompletely charged polymers such as poly (ethylenimine). Payable to its superior power-driven qualities, great bio-compatibility, bio-degradability, affordability, in addition adaptability in groundwork, fibroin, which is primarily collected from the cocoon of domesticated silkwormBombaymore, has attracted growing interest. For the formulation of fibroin nanoparticles (FNPs), these qualities are ideal. As a result of this one amphoteric qualities, fibroin can also be strengthen and changed by cross-linking either by him or with other absolutely charged polymer such poly (ethyl-enimine).

## 2. FIBROIN EXTRACTION AND SILK SOURCE

### 2.1 Silk Source

That one is important to note that multiple other insect orders, as well as extra than 30,000 spider specie and over 113,000 species of Lepidoptera insects, are capable of producing thousands of different types of silk. Several of them still go unidentified. Among these, mul-berry silk, which is primarily generated by the silkworm Bombayadded, accounts for more than 90% of the available silk.

Throughout the more than 5000 year present of the textiles business, silk has stood extensively employed, particularly in Asia-Pacific nations. A Bombaymore silk cocoon that was discovered, Shanxi, ancient Best china, and carbon date to between 4000 to 3000 BC, is the oldest example of silks ever discovered. Similar to this, the first silk fabric was used to wrap a child body in Henan, the source of Chinese evolution, around 3630 BC. For thousands of years, only China produced silk since the process stayed a closely-guarded top top-undisclosed within the Chinese empires. Silk was described by traders as a substance "made from the fleece of lambs sprayed with water and exposed to sunlight." Later, other Asian nations including Korea, Japan, and India came to know the secret. The Han dynasty in China (206 BC-220 AD) built the Silk Highway, a commercial complex linking the East (Hangman, now China) and the West. It was during this time that the Western cultures first recognized silk. Silk continued to be China's main export despite the exchange of many other goods and ideas, including paper, gunpowder, and religions like Buddhism.

### 2.2 Fibrion Extraction

Normally, it is probable to extract fibroin since silkworm cocoons. A silksfiber is made up of a core of fibroin (about 75% weight to weight) and an outer coating of servicing (around 25% weight to weight), a protein that resembles glue. Servicing is made up of number of water soluble spherical glycoproteins that can elicit immune reactions and that can be eliminated using the thermochemical procedure known as degumming. By dismembering mature fifth instar silkworm larvae, fibroin can be recovered directly from the worm subsequent gland without the necessity for degumming, which is unethical. Fibroin is denoted to as silk I in this instance and is liquid and water soluble. The degummed silk fiber, on the other arrow, contains insoluble fibroin silk II and needs additional processing to be converted back to silk . The document presents a typical process for making reinforced fibroin from silkworm cocoon. Although the techniques used in various studies may change significantly, they consistently use Sodium carbonateas a degumming cause and a chemotropic salt clarification as a silk II to silk I transform ant. For instance, little bits of cocoon are dissolved in a Sodium carbonatesolution to degum them, and then they are boiled for 30 to 60 minutes at 100 °C. The silk fibroin will be cleaned three times with ultra-pure water before being air dried and kept at room infection for storage because it is insoluble in Na<sub>2</sub>CO<sub>3</sub>. The fibroin from the preceding process is further dissolved in Libra or CaCl<sub>2</sub> fibroin saltsexplanationrelation of 1:4 w/w), and then heated to 60 to 90 °C to create regenerated fibroin.

## 2.3 Respiratory Administration

Both local and systemic treatments may be able to deliver drugs to the lung. In comparison to standard dosage forms, one container locally treat lung and respiratory disorders (such as lung tumor and tuberculosis TB) through a lower dosage and fewer adverse effects. Due to the vast surface area of the lungs, a medicine can be administered orally and be quickly and effectively absorbed into the body without being broken down by first-pass metabolism. Their aerodynamic particle size turns out to be a crucial component in the efficient delivery of the FNP to the lung. The geometric diameter (defined as the diameter dignified by DLS or SEM or TEM technique) and particle density are both factors that affect the aerodynamic diameter for spherical particles. Particles with an aerodynamic diameter of (1) greater than 10 micrometers accumulate in the oropharyngeal region, primarily the larynx 2 between 5 and 10 micrometers are mostly lodged in the large airways between one and five micrometers can be deposited in the unimportant airways and alveoli and less than 500 nanometers may be diffused back into the atmosphere during exhalation. FNPs are an intriguing potential for this use because numerous inhalation treatments intended to close by treat the lung illnesses are in the clinical increase stage. This field of study is still relatively unexplored and requires significant work in the near future.

Employing the spray-freeze-drying process, cisplatin-loaded FNPs were created for the first time as a pulmonary drug transfer organization for the cure of lung cancer. An *in vitro* aerosolization impact or was employed to heating pad the FNP precipitate into the waterless powder inhaler apparatus while using manifold as an excipient. According to *in vitro* lung deposition, all particles exhibit high aerosolization capability that is on par with that of commercial dry powder inhalers. Furthermore, the A549 animal lung epithelial cell line stayed very sensitive to the cisplatin-loaded FNPs, while the blank FNPs were bio-compatible.

## 2.4 Transdermal Administrations

The layer corneum, the skin outermost layer, obliges as a defence against external elements like germs, viruses, and chemical agents. The skin stands the major tissue in the body. The medicinal chemicals must therefore penetrate this barrier in order to spread deeper layer (such as the derm) and ultimately the systemic movement in order to be administered transdermal. In general, NPs with a callous size of less than 4 nm can infiltrate and permeate integral skin; those amongst 4 and 20 nm may do the same for damaged skin; those amongst 21 to 45 nm can only do so for damaged skin and those greater than 45 nm are unable to do either and instead disperse in the stratum corneum. To this purpose, the amphiphilic characteristics of both crystalline hydrophobic counties and shapeless hydrophilic dominions, as well as its capacity to favorably adjust the element extent, make fibroin a promising carrier. Research in this field is scarce despite the potential. Therefore, it might be a worthwhile strategy for expanding the use of FNP trans-dermal in the near yet to come.

The desolation approach was effectively used to create globular FNPs with a kind size of 42.3 nm and a tight size delivery with poly-dispersity indices of fewer than 0.3. For a week, the elements remained unchanging in their fluid dispersion form. Then, fluorescent NHS-rhodamine was coupled to FNPs for particle tracking. Mice were used in an *in vivo* test to assess skin permeability. Fluorescent signals were discovered six hours after injection in the stratum corneum, hair follicle, the epidermis, and the dermis layer, demonstrating that minor-sized FNPs can pass through the stratum corneum via the paracellular path and advance deeply into the skin.

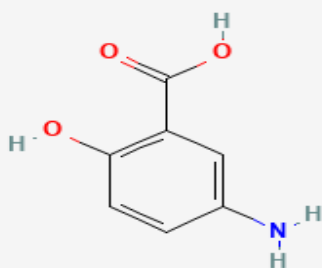
In a different study found that fibroins, in the method of a hydrogel, be able to improve the trans-dermal transport of curcumin-loaded polymeric nanoparticles in a mouse model of psoriasis. As a highly lipophilic substance, curcumin is readily entrapped in the stratum corneum. As a result, the authors enclosed curcumin in RRR-tocopheryl succinate-grafted-polyline, a self-assembling amphiphilic polymer. The round 24.4 nm particles were before further combined into fibroin hydro-gel, where they had an entrapment effectiveness of 78.45% and a drug loading capacity of 3.49%. Occlusive effect resulted from the creation of a tinny fibroin gel coat, which kept the skin's external moist and lengthened the time the formulation was in contact with the skin. As a result, the NPs were translocated crosswise the profounder skin layers.

## 3. Mesalamine

Meclizine, sometimes referred to as mesalamine before 5-aminosalicylic acid (5-ASA), is a drug used to treat ulcerative colitis and Crohn's disease as well as other inflammatory bowel disorders. It is typically used for diseases that are mild to fairly severe. It can be ingested or used externally. A prescription drug called melamine acts as an anti-inflammatory by reducing swelling or inflammation. Inflammatory bowel disease

ulcerative colitis can be treated with melamine (IBD). It is a member of the 5-aminosalicylic acid drug class (5-ASA). Meclizine is a different name for melamine. Apprise, Asarco, Delzicol, Lialda, and Pentose are a few of the oral melamine brand names. Canasta, Rows, and Pentose are a few of the melamine brand names that can be administered rectally.

When a person learns that she is pregnant, they may consider quitting their medicine altogether or modifying how they take it. Before changing how you take this medication, it is crucial to consult with your healthcare professionals. Your healthcare professionals can discuss with you the advantages of treating your disease and the dangers of leaving a sickness untreated while pregnant. It's vital to weigh the advantages of managing your IBD while pregnant. IBD left untreated raises the possibility of difficulties for both the expectant mother and the fetus. In order to learn more about IBD.



#### 4. Challenges, conclusions, and outlooks

Fibroin immobile has several drawbacks that need to be overcome despite having many benefits as a drug transfer technology used in several administration routes. First, a thorough and appropriate servicing removal process from silk fibres is required since servicing may have immunogenic effects. Another, the gradual poverty of silk II's crystalline antiparallel sheet dominions could be problematic in some applications that need for a quick and complete removal of the Nano particulate carrier. Silk II content can be determined and considered using systematic techniques like FTIR, XRD, DSC, then NMR. Because each preparation method produces a different amount of crystalline material, watchful consideration and scheming may be helpful in selecting the best technique. Thirdly, because fibroin is a protein, immune system components like macrophages and giant cells may attack it with proteolysis activity. As a result, off-target drug release occurs as a result of granuloma formation and encapsulation within these immune system components. The issues might be resolved by coating or integrating fibroin before FNPthru PEG or other hydro-philic polymers. Fourth, just like other normal products, fibroin can be mined from a variety of sources. As a result, each batch's properties vary slightly due to variations in the post translational process between different species and people. Therefore, a consistent abstraction technique and the sample characteristics (i.e. MW) are required. Practise of heritably recombinant fibroin might be able to solve this problem. Finally, despite showing great promise in enhancing the stability, extending the release profiles, and safeguarding the encapsulated pharmaceuticals, FNPs are not the sole means of delivering targeted drugs. As a result of the unspecific targeting, limited treatment effectiveness and systemic toxicity may result. Fibroin surface variation with a particular ligand—such as folic acid for tumour targeting—by both covalent and non-covalent bonding demonstrates its efficacy in this regard. These restrictions and difficulties could, however, be solved in one way or another. As a result, fibroin, particularly FNPs, has a strong propensity to be the preferred delivery route for a variety of beneficial agents such as small molecule medications, protein therapeutics, genes, and vaccines. Additionally, a variety of FNP administration methods, including parenteral, oral, trans-dermal, ophthalmic, local bone grafting, and respiratory, have been studied. Further research should concentrate on the less explored yet viable pathways, notably ocular and respiratory, due to the FNP's favourable features. The majority of research on FNPs is based on in vitro besides in-vivo test, thus further medical trials need be carried out to possibly bring the use of FNP to the arcade.

## 5. Acknowledgements

I situate energy on this plan. Though, short of the caring sustenance and support of numerous folks and organisations, it would not take stood practicable. I poverty to express my sincere thankfulness to each and each one of them. Helpful assistance and encouragement that aid in the accomplishment of this job. I want to suggestion my sincere gratefulness and thanks to persons in the occupation who took the time to fee me such near helpfulness.

## 6. References :-

- Altman GH, Diaz F, Jakuba C, et al. (2003). Silk-based biomaterials. *Biomaterials* 24:401–16. [[PubMed](#)] [[Google Scholar](#)]
- Aramwit P. (2012). Silk materials for drug delivery devices. In: Pornanong A, ed. *Silk Properties, Production and Uses*. Nova Publishers, 219–246. [[Google Scholar](#)]
- Baimark Y, Srihanam P, Srisuwan Y, Phinyocheep P. (2010). Preparation of porous silk fibroin microparticles by a water-in-oil emulsification-diffusion method. *J Appl Polym Sci* 118:1127–33. [[Google Scholar](#)]
- Banerjee A, Qi J, Gogoi R, et al. (2016). Role of nanoparticle size, shape and surface chemistry in oral drug delivery. *J Control Release* 238:176–85. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
- Bekersky I, Fielding RM, Dressler DE, et al. (2002). Plasma protein binding of amphotericin B and pharmacokinetics of bound versus unbound amphotericin B after administration of intravenous liposomal amphotericin B (AmBisome) and amphotericin B deoxycholate. *Antimicrob Agents Chemother* 46:834–40. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
- Belanger WA. (2011). *The silk road in world history*. By Xinru Liu. Oxford: Oxford University Press; 2010. x, 168 pp. \$74.00 (cloth); \$19.95 (paper). *J Asian Stud* 70:1156–7. [[Google Scholar](#)]
- Belton DJ, Plowright R, Kaplan DL, Perry CC. (2018). A robust spectroscopic method for the determination of protein conformational composition – application to the annealing of silk. *Acta Biomater* 73:355–64. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
- Calvert P. (1998). Silk and sequence. *Nature* 393:309–11. [[Google Scholar](#)]
- Cao T-T, Zhou Z-Z, Zhang Y-Q. (2014). Processing of  $\beta$ -glucosidase–silk fibroin nanoparticle bioconjugates and their characteristics. *Appl Biochem Biotechnol* 173:544–51. [[PubMed](#)] [[Google Scholar](#)]
- Cao Y, Wang B. (2009). Biodegradation of silk biomaterials. *Int J Mol Sci* 10:1514–24. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
- Cao Z, Chen X, Yao J, et al. (2007). The preparation of regenerated silk fibroin microspheres. *Soft Matter* 3:910–5. [[PubMed](#)] [[Google Scholar](#)]
- Chen BQ, Kankala RK, He GY, et al. (2018). Supercritical fluid-assisted fabrication of indocyanine green-encapsulated silk fibroin nanoparticles for dual-triggered cancer therapy. *ACS Biomater Sci Eng* 4:3487–97. [[PubMed](#)] [[Google Scholar](#)]
- Cheng G, Wang X, Tao S, et al. (2015). Differences in regenerated silk fibroin prepared with different solvent systems: from structures to conformational changes. *J Appl Polym Sci* 132: 41959. [[Google Scholar](#)]
- Chomchalao P, Nimtrakul P, Pham DT, Tiyaboonchai W. (2020). Development of amphotericin B-loaded fibroin nanoparticles: a novel approach for topical ocular application. *J Mater Sci* 55:5268–79. [[Google Scholar](#)]
- Chomchalao P, Pongcharoen S, Sutheerawattananonda M, Tiyaboonchai W. (2013). Fibroin and fibroin blended three-dimensional scaffolds for rat chondrocyte culture. *Biomed Eng Online* 12:28. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
- Couvreur P, Barratt G, Fattal E, et al. (2002). Nanocapsule technology: a review. *Crit Rev Ther Drug Carrier Syst* 19:99–134. [[PubMed](#)] [[Google Scholar](#)]
- Date AA, Hanes J, Ensign LM. (2016). Nanoparticles for oral delivery: design, evaluation and state-of-the-art. *J Control Release* 240:504–26. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
- deMoraes MA, Nogueira GM, Weska RF, Beppu MM. (2010). Preparation and characterization of insoluble silk fibroin/chitosan blend films. *Polymer* 2:719–27. [[Google Scholar](#)]

- Ding B, Wahid MA, Wang Z, et al. (2017). Triptolide and celastrol loaded silk fibroin nanoparticles show synergistic effect against human pancreatic cancer cells. *Nanoscale* 9:11739–53. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
- Dong Y, Dong P, Huang D, et al. (2015). Fabrication and characterization of silk fibroin-coated liposomes for ocular drug delivery. *Eur J Pharm Biopharm* 91:82–90. [[PubMed](#)] [[Google Scholar](#)]
- Estey T, Kang J, Schwendeman SP, Carpenter JF. (2006). BSA degradation under acidic conditions: a model for protein instability during release from PLGA delivery systems. *J Pharm Sci* 95:1626–39. [[PubMed](#)] [[Google Scholar](#)]
- Etheridge ML, Campbell SA, Erdman AG, et al. (2013). The big picture on nanomedicine: the state of investigational and approved nanomedicine products. *NanomedNanotechnolBiol Med* 9:1–14. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
- Farokhi M, Mottaghitalab F, Ai J, Shokrgozar MA. (2013). Sustained release of platelet-derived growth factor and vascular endothelial growth factor from silk/calcium phosphate/PLGA based nanocomposite scaffold. *Int J Pharm* 454:216–25. [[PubMed](#)] [[Google Scholar](#)]
- Farokhi M, Mottaghitalab F, Hadjati J, et al. (2014). Structural and functional changes of silk fibroin scaffold due to hydrolytic degradation. *J Appl Polym Sci* 131:1–8. [[Google Scholar](#)]
- Filon FL, Mauro M, Adami G, et al. (2015). Nanoparticles skin absorption: new aspects for a safety profile evaluation. *Regul Toxicol Pharmacol* 72:310–22. [[PubMed](#)] [[Google Scholar](#)]
- Foo CWP, Bini E, Hensman J, et al. (2006). Role of pH and charge on silk protein assembly in insects and spiders. *Appl Phys A* 82:223–33. [[Google Scholar](#)]
- Gianak O, Kyzas GZ, Samanidou VF, Deliyanni EA. (2019). A review for the synthesis of silk fibroin nanoparticles with different techniques and their ability to be used for drug delivery. *Curr Anal Chem* 15:339–48. [[Google Scholar](#)]
- Giteau A, Venier-Julienne MC, Aubert-Pouëssel A, Benoit JP. (2008). How to achieve sustained and complete protein release from PLGA-based microparticles? *Int J Pharm* 350:14–26. [[PubMed](#)] [[Google Scholar](#)]
- Gong Y, Li L, Gong D, et al. (2016). Biomolecular evidence of silk from 8,500 years ago. *PLoS One* 11:e0168042. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
- Gou S, Huang Y, Wan Y, et al. (2019). Multi-bioresponsive silk fibroin-based nanoparticles with on-demand cytoplasmic drug release capacity for CD44-targeted alleviation of ulcerative colitis. *Biomaterials* 212:39–54. [[PubMed](#)] [[Google Scholar](#)]
- Gulsen D, Chauhan A. (2005). Dispersion of microemulsion drops in HEMA hydrogel: a potential ophthalmic drug delivery vehicle. *Int J Pharm* 292:95–117. [[PubMed](#)] [[Google Scholar](#)]
- Gupta V, Aseh A, Ríos CN, et al. (2009). Fabrication and characterization of silk fibroin-derived curcumin nanoparticles for cancer therapy. *Int J Nanomedicine* 4:115–22. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
- Hardy JG, Römer LM, Scheibel TR. (2008). Polymeric materials based on silk proteins. *Polymer (Guildf)* 49:4309–27. [[Google Scholar](#)]
- HassaniBesheli N, Damoogh S, Zafar B, et al. (2018). Preparation of a codelivery system based on vancomycin/silk scaffold containing silk nanoparticle loaded VEGF. *ACS Biomater Sci Eng* 4:2836–46. [[PubMed](#)] [[Google Scholar](#)]
- HassaniBesheli N, Mottaghitalab F, Eslami M, et al. (2017). Sustainable release of vancomycin from silk fibroin nanoparticles for treating severe bone infection in rat tibia osteomyelitis model. *ACS Appl Mater Interfaces* 9:5128–38. [[PubMed](#)] [[Google Scholar](#)]
- Holland C, Numata K, Rnjak-Kovacina J, Seib FP. (2019). The biomedical use of silk: past, present, future. *Adv Healthcare Mater* 8:1800465. [[PubMed](#)] [[Google Scholar](#)]
- Hoshyar N, Gray S, Han H, Bao G. (2016). The effect of nanoparticle size on in vivo pharmacokinetics and cellular interaction. *Nanomedicine (Lond)* 11:673–92. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
- Huang D, Wang L, Dong Y, et al. (2014). A novel technology using transscleral ultrasound to deliver protein loaded nanoparticles. *Eur J Pharm Biopharm* 88:104–15. [[PubMed](#)] [[Google Scholar](#)]
- Huang Y, Lu Q, Li M, et al. (2011). Silk fibroin microsphere drug carriers prepared under electric fields. *Chin Sci Bull* 56:1013–8. [[Google Scholar](#)]
- Jeencham R, Sutheerawattananonda M, Tiyaboonchai W. (2019). Preparation and characterization of chitosan/regenerated silk fibroin (CS/RSF) films as a biomaterial for contact lenses-based ophthalmic drug delivery system. *Int J Appl Pharm* 11:275–84. [[Google Scholar](#)]

- Jia L, Guo L, Zhu J, Ma Y. (2014). Stability and cytocompatibility of silk fibroin-capped gold nanoparticles. *Mater SciEng C* 43:231–6. [[PubMed](#)] [[Google Scholar](#)]
- Jiang S, Franco YL, Zhou Y, Chen J. (2018). Nanotechnology in retinal drug delivery. *Int J Ophthalmol* 11:1038–44. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
- Jin H-J, Kaplan DL. (2003). Mechanism of silk processing in insects and spiders. *Nature* 424:1057–61. [[PubMed](#)] [[Google Scholar](#)]
- Joseph RR, Venkatraman SS. (2017). Drug delivery to the eye: what benefits do nanocarriers offer? *Nanomedicine* 12:683–702. [[PubMed](#)] [[Google Scholar](#)]
- Kanokpanont S, Damrongsakkul S, Ratanavaraporn J, Aramwit P. (2013). Physico-chemical properties and efficacy of silk fibroin fabric coated with different waxes as wound dressing. *Int J BiolMacromol* 55:88–97. [[PubMed](#)] [[Google Scholar](#)]
- Kaplan D, Adams WW, Farmer B, Viney C. (1993). Silk: biology, structure, properties, and genetics. In: Kaplan D, Wade Adams W, Farmer B, Viney C, eds. *Silk polymers*. Washington, DC: American Chemical Society, 1–2. [[Google Scholar](#)]
- Kazemimostaghim M, Rajkhowa R, Tsuzuki T, Wang X. (2013). Production of submicron silk particles by milling. *Powder Technol* 241:230–5. [[Google Scholar](#)]
- Keten S, Xu Z, Ihle B, Buehler MJ. (2010). Nanoconfinement controls stiffness, strength and mechanical toughness of  $\beta$ -sheet crystals in silk. *Nat Mater* 9:359–67. [[PubMed](#)] [[Google Scholar](#)]
- Ki CS, Park YH, Jin H-J. (2009). Silk protein as a fascinating biomedical polymer: structural fundamentals and applications. *Macromol Res* 17:935–42. [[Google Scholar](#)]
- Kim SY, Naskar D, Kundu SC, et al. (2015). Formulation of biologically-inspired silk-based drug carriers for pulmonary delivery targeted for lung cancer. *Sci Rep* 5:11878. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
- Kim W-J, Islam R, Kim B-S, et al. (2017). Direct delivery of recombinant pin1 protein rescued osteoblast differentiation of pin1-deficient cells. *J Cell Physiol* 232:2798–805. [[PubMed](#)] [[Google Scholar](#)]
- Kojthung A, Meesilpa P, Sudatis B, et al. (2008). Effects of gamma radiation on biodegradation of *Bombyxmori* silk fibroin. *IntBiodeteriorBiodegrad* 62:487–90. [[Google Scholar](#)]
- Kundu J, Chung Y-I, Kim YH, et al. (2010). Silk fibroin nanoparticles for cellular uptake and control release. *Int J Pharm* 388:242–50. [[PubMed](#)] [[Google Scholar](#)]
- Kurioka A, Masayoshi Y, Hirano H. (1999). Primary structure and possible functions of a trypsin inhibitor of *Bombyxmori*. *Eur J Biochem* 259:120–6. [[PubMed](#)] [[Google Scholar](#)]
- Labiris NR, Dolovich MB. (2003). Pulmonary drug delivery. Part I: physiological factors affecting therapeutic effectiveness of aerosolized medications. *Br J ClinPharmacol* 56:588–99. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
- Lammel AS, Hu X, Park S-H, et al. (2010). Controlling silk fibroin particle features for drug delivery. *Biomaterials* 31:4583–91. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
- Lang JC. (1995). Ocular drug delivery conventional ocular formulations. *Adv Drug Deliv Rev* 16:39–43. [[Google Scholar](#)]
- Li C, Vepari C, Jin H-J, et al. (2006). Electrospun silk-BMP-2 scaffolds for bone tissue engineering. *Biomaterials* 27:3115–24. [[PubMed](#)] [[Google Scholar](#)]
- Li H, Tian J, Wu A, et al. (2016). Self-assembled silk fibroin nanoparticles loaded with binary drugs in the treatment of breast carcinoma. *Int J Nanomedicine* 11:4373–80. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
- Liechty WB, Kryscio DR, Slaughter BV, Peppas NA. (2010). Polymers for drug delivery systems. *Annu Rev ChemBiomolEng* 1:149–73. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
- Liu Y, Lv Z, Zhang C, et al. (2014). Preparation and immunogenicity of silk fibroin/chitosan microspheres for DNA vaccine delivery against infectious bursal disease virus. *ShengwuGongchengXuebao/Chin J Biotechnol* 30:393–403. [[PubMed](#)] [[Google Scholar](#)]
- Lozano-Pérez AA, Rivero HC, del Carmen Pérez Hernández M, et al. (2017). Silk fibroin nanoparticles: efficient vehicles for the natural antioxidant quercetin. *Int J Pharm* 518:11–9. [[PubMed](#)] [[Google Scholar](#)]
- Lozano-Pérez AA, Rodriguez-Nogales A, Ortiz-Cullera V, et al. (2014). Silk fibroin nanoparticles constitute a vector for controlled release of resveratrol in an experimental model of inflammatory bowel disease in rats. *Int J Nanomedicine* 9:4507–20. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]

- Lucas F, Barber M, Wolstenholme WA, et al. (1969). Mass-spectrometric determination of the amino acid sequences in peptides isolated from the protein silk fibroin of *Bombyxmori*. *Biochem J* 114:695–702. [PMC free article] [PubMed] [Google Scholar]
- Ma C, Lv L, Liu Y, et al. (2014). *Antheraepernyi* silk fibroin for targeted gene delivery of VEGF165-Ang-1 with PEI. *Biomed Mater* 9:35015. [PubMed] [Google Scholar]
- Mandal BB, Kundu SC. (2008). A novel method for dissolution and stabilization of non-mulberry silk gland protein fibroin using anionic surfactant sodium dodecyl sulfate. *BiotechnolBioeng* 99:1482–9. [PubMed] [Google Scholar]
- Mao K-L, Fan Z-L, Yuan J-D, et al. (2017). Skin-penetrating polymeric nanoparticles incorporated in silk fibroin hydrogel for topical delivery of curcumin to improve its therapeutic effect on psoriasis mouse model. *Colloids Surf B Biointerfaces* 160:704–14. [PubMed] [Google Scholar]
- Mao S, Guo C, Shi Y, Li LC. (2012). Recent advances in polymeric microspheres for parenteral drug delivery – part 1. *Expert Opin Drug Deliv* 9:1161–76. [PubMed] [Google Scholar]
- Mathur AB, Gupta V. (2010). Silk fibroin-derived nanoparticles for biomedical applications. *Nanomedicine* 5:807–20. [PubMed] [Google Scholar]
- Matsumoto A, Chen J, Collette AL, et al. (2006). Mechanisms of silk fibroin sol–gel transitions. *J PhysChem B* 110:21630–8. [PubMed] [Google Scholar]
- Matteis VD. (2017). Exposure to inorganic nanoparticles: routes of entry, immune response, biodistribution and in vitro/in vivo toxicity evaluation. *Toxics* 5:29. [PMC free article] [PubMed] [Google Scholar]
- Mehnert W, Mäder K. (2001). Solid lipid nanoparticles: production, characterization and applications. *Adv Drug Deliv Rev* 47:165–96. [PubMed] [Google Scholar]
- Melke J, Midha S, Ghosh S, et al. (2016). Silk fibroin as biomaterial for bone tissue engineering. *ActaBiomater* 31:1–16. [PubMed] [Google Scholar]
- Min B-M, Jeong L, Lee KY, Park WH. (2006). Regenerated silk fibroin nanofibers: water vapor-induced structural changes and their effects on the behavior of normal human cells. *MacromolBiosci* 6:285–92. [PubMed] [Google Scholar]
- Minoura N, Aiba SI, Higuchi M, et al. (1995). Attachment and growth of fibroblast cells on silk fibroin. *BiochemBiophys Res Commun* 208:511–6. [PubMed] [Google Scholar]
- Mohammed MA, Syeda JTM, Wasan KM, Wasan EK. (2017). An overview of chitosan nanoparticles and its application in non-parenteral drug delivery. *Pharmaceutics* 9:53. [PMC free article] [PubMed] [Google Scholar]
- Motta A, Fambri L, Migliaresi C. (2002). Regenerated silk fibroin films: thermal and dynamic mechanical analysis. *MacromolChemPhys* 203:1658–65. [Google Scholar]
- Mottaghitlab F, Farokhi M, Shokrgozar MA, et al. (2015). Silk fibroin nanoparticle as a novel drug delivery system. *J Control Release* 206:161–76. [PubMed] [Google Scholar]
- Myung SJ, Kim H-S, Kim Y, et al. (2008). Fluorescent silk fibroin nanoparticles prepared using a reverse microemulsion. *Macromol Res* 16:604–8. [Google Scholar]
- Nair LS, Laurencin CT. (2006). Polymers as biomaterials for tissue engineering and controlled drug delivery. In: Lee K, Kaplan D, eds. *Tissue Engineering I. Advances in Biochemical Engineering/Biotechnology*. Berlin, Heidelberg: Springer, 47–90. [PubMed] [Google Scholar]
- Nam J, Park YH. (2001). Morphology of regenerated silk fibroin: effects of freezing temperature, alcohol addition, and molecular weight. *J ApplPolymSci* 81:3008–21. [Google Scholar]
- Naskar S, Kuotsu K, Sharma S. (2019). Chitosan-based nanoparticles as drug delivery systems: a review on two decades of research. *J Drug Target* 27:379–93. [PubMed] [Google Scholar]
- Numata K, Cebe P, Kaplan DL. (2010). Mechanism of enzymatic degradation of beta-sheet crystals. *Biomaterials* 31:2926–33. [PMC free article] [PubMed] [Google Scholar]
- Numata K, Kaplan DL. (2010). Silk-based delivery systems of bioactive molecules. *Adv Drug Deliv Rev* 62:1497–508. [PMC free article] [PubMed] [Google Scholar]
- Padol AR, Jayakumar K, Shridhar NB, et al. (2011). Safety evaluation of silk protein film (a novel wound healing agent) in terms of acute dermal toxicity, acute dermal irritation and skin sensitization. *ToxicolInt* 18:17–21. [PMC free article] [PubMed] [Google Scholar]
- Pascoli M, de Lima R, Fraceto LF. (2018a). Zein nanoparticles and strategies to improve colloidal stability: a mini-review. *Front Chem* 6:6. [PMC free article] [PubMed] [Google Scholar]



- Pham DT, Saelim N, Tiyaboonchai W. (2018b). Crosslinked fibroin nanoparticles using EDC or PEI for drug delivery: physicochemical properties, crystallinity and structure. *J Mater Sci* 53:14087–103. [[Google Scholar](#)]
- Pham DT, Saelim N, Tiyaboonchai W. (2018b). Design of experiments model for the optimization of silk fibroin based nanoparticles. *Int J Appl Pharm* 10:195–201. [[Google Scholar](#)]
- Pham DT, Saelim N, Tiyaboonchai W. (2019a). Alpha mangostin loaded crosslinked silk fibroin-based nanoparticles for cancer chemotherapy. *Colloids Surf B Biointerfaces* 181:705–13. [[PubMed](#)] [[Google Scholar](#)]
- Pham DT, Saelim N, Tiyaboonchai W. (2019b). Paclitaxel loaded EDC-crosslinked fibroin nanoparticles: a potential approach for colon cancer treatment. *Drug DelivTransl Res*. [[PubMed](#)] [[Google Scholar](#)]
- Pham DT, Tetyczka C, Hartl S, et al. (2019c). Comprehensive investigations of fibroin and poly(ethylenimine) functionalized fibroin nanoparticles for ulcerative colitis treatment. *J Drug DelivSci Technol*. [[Google Scholar](#)]
- Qi Y, Wang H, Wei K, et al. (2017). A review of structure construction of silk fibroin biomaterials from single structures to multi-level structures. *Int J MolSci* 18:237. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
- Qu J, Liu Y, Yu Y, et al. (2014). Silk fibroin nanoparticles prepared by electrospray as controlled release carriers of cisplatin. *Mater SciEng C Mater BiolAppl* 44:166–74. [[PubMed](#)] [[Google Scholar](#)]
- Rajkhowa R, Wang L, Kanwar J, Wang X. (2009). Fabrication of ultrafine powder from eri silk through attritor and jet milling. *Powder Technol* 191:155–63. [[Google Scholar](#)]
- Rajkhowa R, Wang L, Wang X. (2008). Ultra-fine silk powder preparation through rotary and ball milling. *Powder Technol* 185:87–95. [[Google Scholar](#)]
- Rockwood DN, Preda RC, Yücel T, et al. (2011). Materials fabrication from *Bombyxmori* silk fibroin. *Nat Protoc* 6:1612–31. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
- Roy K, Patel YS, Kanwar RK, et al. (2016). Biodegradable Eri silk nanoparticles as a delivery vehicle for bovine lactoferrin against MDA-MB-231 and MCF-7 breast cancer cells. *Int J Nanomedicine* 11:25–44. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)]
- Sahoo N, Sahoo RK, Biswas N, et al. (2015). Recent advancement of gelatin nanoparticles in drug and vaccine delivery. *Int J BiolMacromol* 81:317–31. [[PubMed](#)] [[Google Scholar](#)]
- Sakabe H, Ito H, Miyamoto T, et al. (1989). In vivo blood compatibility of regenerated silk fibroin. *Sen'iGakkaishi* 45:487–90. [[Google Scholar](#)]
- Santin M, Motta A, Freddi G, Cannas M. (1999). In vitro evaluation of the inflammatory potential of the silk fibroin. *J Biomed Mater Res* 46:382–9. [[PubMed](#)] [[Google Scholar](#)]
- Shahbazi B, Taghipour M, Rahmani H, et al. (2015). Preparation and characterization of silk fibroin/oligochitosan nanoparticles for siRNA delivery. *Colloids Surf B Biointerfaces* 136:867–77. [[PubMed](#)] [[Google Scholar](#)]
- Shi P, Abbah SA, Saran K, et al. (2013). Silk fibroin-based complex particles with bioactive encrustation for bone morphogenetic protein 2 delivery. *Biomacromolecules* 14:4465–74. [[PubMed](#)] [[Google Scholar](#)]