IJCRT.ORG

ISSN: 2320-2882



INTERNATIONAL JOURNAL OF CREATIVE RESEARCH THOUGHTS (IJCRT)

An International Open Access, Peer-reviewed, Refereed Journal

"Overview of Vaccine Adjuvants"

Prajakta Awatade*, Manisha Sukre, Amit Kasabe , Vikram Veer Shankarrao Ursal College of Pharmaceutical Sciences and Research Centre, Kharadi, Pune-14

Abstract: This review describes vaccines adjuvants and their role in vaccine development. Their mechanism of action, safety aspects, and carriers in vaccination. Synthetic glycopeptides, Glycolipids are important vaccine adjuvants. Nowdays there is tremendous interest in the use of lipidated and carbohydrates based vaccine adjuvants. Therefore particularly special attention is given on vaccine adjuvants containing lipids or and monosaccharides or made from both.

Keywords-Adjuvants, Vaccine, Immunoadjuvant

1.0 Introduction

Infectious diseases continue to pose challaenges for human and animal communities and the associated inflammation phenomena being one of the earliest medical symtoms identified. Immunisation being foremost healthcare intervention against infectious diseases, has been around for more than 300 years. A vaccination includes pathogens(microbial agent that can cause disease) that are modified in such a manner that alteration renders the pathogen inactive with the ability to inflict disease while retaining adequate similarity with the unmodified pathogen to stimulate the vaccinated host to become immune to disease by the host's immune system actions. Vaccination has proved to be a very cost-effective way of controlling infectious diseases caused by microbial infections and has been recognized in its current form since Jenner introduced vaccinia (cowpox virus) as the first safe vaccine in his groundbreaking work in the late 18th century. [1] Numerous other promising vaccines have been formulated, based on pathogens or the pathogen derived toxins that have been subdued or destroyed. Despite this, effective vaccines against HIV1, tuberculosis, malaria and various infectious respiratory diseases are inadequate. For example, the eradication of smallpox (officially declared extinct in 1980) and the sudden and striking fall of formerly widespread dangerous diseases such as polio have been great achievements in vaccination. Nevertheless, there are still large number of various infectious diseases around world (malaria, tuberculosis, and bacterial and viral diseases). These infections are commonly triggered by complex pathogens where there is a need for a more rational approach to design of vaccines. Many of these "complex" pathogens target the immune system itself or may alter quickly to prevent the immune system. [2]

There is an emerging trend to target these infectious diseases with new-generation vaccines. With an improved understanding of the immune system at molecular level, we are able to use well-defined chemical processes to generate such new vaccines with optimal properties. In the formulation of successful vaccines, the essential factor is the integration of effective immune potentiators (also known as Adjuvants) with the antigen. Traditional vaccines also contain several immune stimulatory components that can benefit or serve as ajuvants for additional T-cell protection, such as bacterial DNA or LPS in whole-cell vaccines. As modern safe vaccine use purified subunit antigens or recombinant proteins, the vital immunestimulatory ingradients have been lost, which needs to recompensated through addition of an appropriate adjuvant, which in turn lead to increased the search for molecularly identified adjuvants i.e.chemicals, which are non-immunogenic but are capable of stimulating humoral and cellular immunity and ,most of all beneficial for both animals and humans.there is an extreme and increasing interest in the roduction of vaccine adjuvants nowadays since many of the current subunit vaccines candidates lack adequate immunogenicity. There is also a great need for a healthy and powerful adjuvants that can activate the cellular and humoral immune response. [3]

2.0 Adjuvants

"The Adjuvant" came from the Latin "adjuvare," which means "to assist, strengthen, or improve" (Vogel, 1998). Ramon first explained immunological adjuvant in 1924 as "substances used in combination with an exact antigen that developed a stronger immunological response than the antigen itself. Adjuvants are chemical substances that induce a stronger and more durable response to the vaccine when used in conjunction with vaccine antigens, relative to that produced by the vaccine itself. Most of the vaccines generally used are administered along with adjuvants, especially those comprising of inactivated (killed) microorganisms and those comprising purified molecules. The first adjuvants ever identified in the scientific literature, i.e. aluminum salts, remain normal adjuvants accepted for use in humans, considering the significant attempts made in recent decades to create new vaccine adjuvants.^[4] An ideal subunit vaccine contains a pathogen derived antigen which has been shown to elicit a defensive immune response. Thus, historically, the first subunit vaccines produced are still bacterial toxins in their inactivated states are historically, first subunit vaccines produced.^[5] The first attempts to strengthen the immunogenicity of these toxoids in animals were established by Ramon, who found that the highest antibody titers were obtained in animals in which localized inflammatory reactions were caused by the concomitant injection of other substances, such as sterilized tapioca, aluminum salts, lanolin, tannin, kaolin, etc. [6] Glenny et al. in 1926 proposed the use of aluminum hydroxide as an adjuvant.

2.1 Role of adjuvants in the immune responses

Everthrough the exact mechanism of adjuvanticity is not completely understood till date, immune bosting propensity of adjuvants is attributed to the following:

- Up-regulation of co-stimulatory signals, expression of major histocompatibility complexes (MHC), and improved cytokine release in the activation state of APCs
- Improved antigen presentation by APCs to enhance the magnitude, swiftness and durability of the immune response.
- Enhanced antibody binding affinity to the antigen.
- Elicit cell-mediated immunity and non-apecific lymphocyte proliferation.

2.2 Classification of adjuvants

In the classification of the adjuvant, various scientists considered different parameters. Classification of adjuvant by Cox and Coulter as particulate and non-particulate in two classes. The adjuvants were classified by Vogel in 1998 according to their origin, mode or mechanism of action, and physicochemical characteristics. It is possible to distinguish adjuvants according to their molecular character, origin, or physical-chemical properties (Cox and Coulter 1997, Kwak and Longo 1996), but similar compounds also have divergent immunomodulatory properties.

Broad classification of adjuvants into main groups: -

- Immune potentiators or Immunostimulants; improve responses to antigens, they function directly on the immune system, e.g. TLR ligands, cytokines, bacterial exotoxins, and saponins that induce immune responses.
- Delivery systems or Vehicles; enhance antigen specific immune response, by effective presentation of antigens to the immune system, including controlled release via depot/delivery mechanisms. e.g. Mineral salts, emulsions, liposomes, virosomes, nanoparticles.

Immunopotentiators

Immunopotentiators or immunoadjuvants exerts effects through concerted pathways. The structural analysis of certain adjuvants and the discovery of immune system cell receptors associated with their actions elicit a greater understanding at the molecular level of their modes of action. The mammalian defense mechanism can be broadly classified as "an innate and adaptive immune response". The adaptive immunity, is specific immunity mainly responsible for decreasing or totally restricting pathogen growth. The innate immunity which is non-specific immunity, activates adaptive immunity. [7] The innate immunity is the initial layer of protection, and it uses pattern recognition receptors called PRRs, to identify infections. These PRRs are generally found inside or on the surface of APC, eg. dendritic cells (DCs). These receptors identify pathogen-associated molecular patterns (PAMPs). PAMPs include nucleic acid microbial lipopolysaccharide (LPS), flagella proteins, lipoteichoic acids and glycoprotein (PG) (Figure 2). The TLRs (toll-like receptors), CLRs (lectin type C receptors), NLRs (nucleotide oligomeric domain-like receptors) and RLRs (RIG-like receptors) are the main representatives of PRRs. [8]

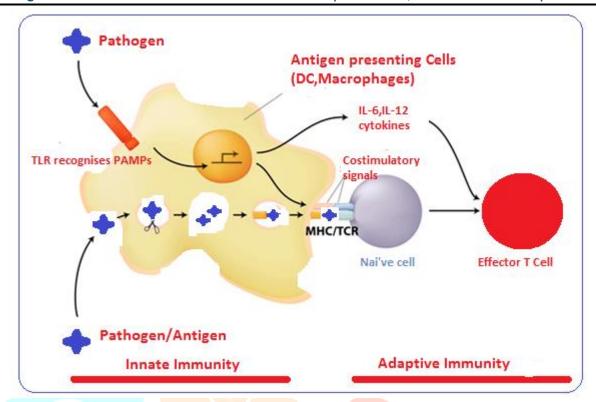


Figure 1: TLR signal involving in immune response propagation

A sequence of downstream signaling events trigger key transcription factors associated with the proliferation of immune responses following pathogen binding at the cell surface TLR. Specifically, NF-kB starts transcribing a variety of interleukins (responsible for growth/differentiation of lymphocytes) and TNF- α (responsible for activating macrophage cells). Additional gene products provide the necessary co-stimulatory signals for naïve Tcell recruitment and subsequent activation of lymphocytes. At the cellular level, these activities represent the bridge between innate and adaptive immunity.

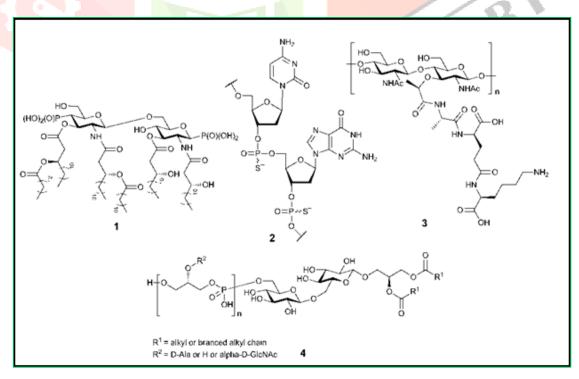


Figure 2: Examples of pathogen-associated molecular patterns (PAMP): LPS lipidA (1), nucleic acids (CpG DNA, 2), peptidoglycan (3), and lipoteichoic acid (4).

Figure 3: The general repeating unit of peptidoglycan (PG) of Gram +Ve (5), Gram -Ve bacteria (6).

TLRs, which could be present just on surface of the cell or even in the endosomes of certain immune cells, seem to be the most widely researched PRRs. TLR 1 to 10 found in humans and TLR 1 to 9, 11 to 12 found in mice. Special attention is being paid to TLR2 and NOD2 in this thesis. With either TLR1 or TLR6, TLR2 receptors form heterodimers and each combination recognizes a variety of ligands. NLRs are found within the cytosol and therefore are not linked to a membrane.

Freund was the first scientist to understand that the immune system can be activated by samples derived from pathogenic bacteria. He discovered that heat mediated systemic immune activation in mammals by killing mycobacterium cells suspended in mineral oil. This discovery resulted in Freund's adjuvant, which is used as an immune stimulator. Freunds adjuvant timulation is not only confined to mycobacterium, but several other bacteria have shown similar effects. It has been proved that the PRRs recognize the chemicals produced from the cell-wall of both Gram+Ve 5 and Gram-Ve 6 bacteria. The TLR2 receptor recognizes lipopeptides and lipoproteins. Peptidoglycan (PG) fragments are recognised either by NOD1 or NOD2 receptors which is an indispensible component of the cell walls of both bacteria types. The fact that bacterial peptidoglycan compounds can activate the innate immunity has stimulated the synthesis of well-defined peptidoglycan fragments to establish the Structural elements essential for receptor activation. [9]

Figure 4: NOD1 ligand iE-DAP (7), NOD2 ligand MDP (8) and TLR2 ligand Pam₃CSK₄ (9)

D-glutamyl-(2S, 6R)-diaminopimelic acid (iE-DAP, 7)^[10] and muramyl dipeptide (MDP, 8)^[11] were established as minimum cores for said NOD1 and NOD2 receptors, respectively (Figure 4). ^[8, 12] Synthetic lipopeptide Pam₃CSK₄ (9) was TLR2 agonist. ^[13] Generally, PRR ligands having immunologic activity are powerful tools for elucidating the molecular pathways of the immune response. ^[8] While sufficient information is obtained from compounds isolated from different bioactive compounds, it is necessary to have synthetic compounds available. Synthetic chemicals are much easier to purify relatively to biological isolated substances to preserve consistency, thereby lacking possible biologically active contamination. Synthetic chemicals easily empower the insertion of appropriate handles, like fluorescent tag that can be designed and synthesized. ^[13] Structure-relevant PRR ligands either with agonistic or antagonistic activities are particularly important in biological studies. Ligands with agonistic properties can be used for the formulation of effecaceous and novel vaccines against various diseases, while antagonists are excellent testing targets for autoimmune disorders. Immunological illnesses like Crohn's disease, Blau condition and inflammatory bowel diseases also include PRRs.

3.0 Pattern Recognition Receptors

While the discovery of PRR ligands with a particular immunological activity has been given a great deal of importance, the number of well-defined identified ligands is limited. Few examples of synthesised ligands are TLR2 (Pam₃CysSK₄), TLR4 (lipidA), TLR9 (CpG oligonucleotides and TLR7 (imiquimod analogues)) which have made significant progress. Additionally, studies on the preparation and screening of peptidoglycan fragments as TLR and NOD receptor agonists have been performed. A summary of the structure-activity relationship (SAR) and assessment of TLR2, NOD1, and NOD2 ligands is given in the following pages.

3.1 Toll-like receptors

Christiane Nüsslein-Volhard was the first one to identify the Toll-gene in Drosophila melanogaster. The genes are responsible for embryonic dorsal-ventral polarity in fruit flies, which means that embryos with such a specific mutation in this gene have their embryonic pattern inverted or lateralized. Nüsslein-Volhard screamed "Toll!" (German for wonderful) when she initially identified embryonic fly mutants having inversed dorsoventral-polarity, and afterwards termed the altered gene the Toll-gene. Later research revealed that the

same gene is involved in the immunological response of insects to fungus.^[16] This Toll-like receptor family is made up of multiple members that have been found in various species, including vertebrates (TLRs). In mammals, 10 TLR subtypes (TLR1-TLR10) have been found, each of which plays important role in the activation of an innate immunity to fight microbial infections.^[17] TLRs play a vital role in the growth of autoimmune and systemic inflammatory diseases ^[18], and cancer.^[19] As a result, TLR2 antagonists are advised for the treatment of such type of disorders.^[20]

3.2 NOD (Nucleotide-binding Oligomerization Domain) like receptors

NOD-LRR cytoplasmic proteins with three domain types: 1) a C-terminal LRR domain for ligand binding, 2) a nucleotide-binding oligomerization domain (NOD domain), and 3) an N-terminal effector's binding domain (EBD domain) for signalling initiation (Inohar et al. 2005 and Martinon, Tschopp 2005). Mammalian genomes include about 20 such proteins, which are divided into two main subfamilies named NODs and NALPs. According to modern evidence, a few of these proteins recognise endogenous or bacterial molecules or stress reactions and form oligomers which activate inflammatory caspases (eg.caspase 1), allowing important inflammatory cytokines like IL-1 to be cleaved and triggered, and/or stimulate the NF-kB signalling pathway, inducing the production of inflammatory molecules. This nod like receptor family is also known as the CATERPILLER (or CLR) or NOD-LRR family. NOD1 and NOD2 are currently known receptors in the domain. NOD1 recognises the peptidoglycan molecule termed meso-DAP found mainly in Gramnegative bacterias, reported Chamaillard in 2003. According to Girardin et al, NOD2 recognises intracellular muramyl dipeptide (MDP) and a peptidoglycan (PG) component found in both types of bacteria.

3.3 NALPS

In humans, there will be 14 members of this group (NALP1- NALP14). NALP proteins may recognize a wide range of ligands from various pathogens (Tschopp, J. et al. 2003). NALP3 activators can be found in muramyl dipeptide, bacteria DNA, toxins, double-stranded RNA and uric acid (Kanneganti et al in 2006; Mariathasan et al in 2006; Martinon et al. 2006). Although it has been demonstrated that these particular compounds activate NALP3, it is unclear whether this is due to actual binding or cellular stresses produced by such ligands. In response to Salmonella and Legionella, several NLRs such as IPAF or NAIP5 activate caspase-1 (Molofsky et al. 2006; Ren et al. 2006).

3.4 Mannose receptors

The mannose receptors (Stahl, Ezekowitz 1998) still a PRR located at the surface of macrophages and dendritic cells. This belongs to a protein family of multilectin receptor and acts as bridge between innate and adaptive iimmunuty. These are type I C-type lectin receptors having lengthy extracellular part which includes an N-terminal cysteine-rich domain, a fibronectin type II (FNII) domain, and a special set of eight C-type lectin-like domains which is referred as the carbohydrate—recognition domains (CRDs), which helps in the idendification of various carbohydrate motifs such as mannose, fucose and glycoconjugates (Taylor and Drickamer 1993). The mannose receptor binds to carbohydrates found on pathogen surfaces and this activation

induces pathogen endocytosis and phagocytosis. Common ligands for mannose receptors (MR) include microbial polysaccharides, proteoglycans, and glycolipids, as well as mammalian glycoproteins containing N-linked high-mannose. These receptors contain another carbohydrate-binding domain, the cysteine-rich domain, which recognizes several endogenous sulfated glycoconjugates, in addition to the C-type lectin domains. Among the ligands containing sulfated oligosaccharides are pituitary hormones, such as lutropin, (Fiete et al. 1998; Simpson et al. 1999).

Antigen mannosylation gives a higher presentation quality of the order of 100, to 10000-fold to T cells.^[21] Antigen administration towards the MR has been shown to be successful in inducing robust immune responses in a number of ways. As a result, when MR-targeted vaccines were given in combination with medicines complimentary activation signals got acativated, which are most beneficial *in vivo*.

3.5 Scavenger receptors

According to Goldstein, scavenger receptors (SRs) were identified from their binding ability and internalizing modified lipoproteins (Goldstein et al. 1979). SR involves a wide range of molecules, including modified low-density lipoproteins, participating in receptor-mediated endocytosis of selected polyanionic ligands (LDL). Many scavenger receptors take part in apoptotic cell, bacterial phagocytosis and cell adhesion. These trans-membrane receptors vary in their structure, including C-type-lectin, collagenous, cysteine-rich and other domain molecules. Scavanger receptors plays significant role in an absorption and removal of damaged sections i.e. changed host molecules or apoptotic cells.

4.0 Mechanism of action and role of immunopotentiators in innate immunity processing

The processing of adaptive immunity is regulated by APCs presenting protein antigens to T- or B-lymphocytes, which inturn depends upon whether antigen is endogenous or exogenous [Perkin and Cohen 2001]. Adjuvants, like amphipathic non-ionic block copolymers that binds exogenous antigens during their internalization by APCs, retain their three-dimensional structure (necessary in the development of neutralizing antibodies) (Cox and Coulter 1997; Hunter 2002). These Adjuvants deliver exogenous antigens to class II (phagosomes) or class I (cytoplasm) pathways for processing (Hunter 2002) that depends upon their size and the conjunction of hydrophilic to hydrophobic chains. These are very efficient in antigen delivery which can also be used with several other immunostimulators. The production of cytokines and chemokines by activated CD4+ helper T cells influence pathogen life or enhance the stimulation of CD8+ T cells and/or B cells. The contact of TCR with the antigen-MHC complex produces one signal, and other from the engagement of T-cell CD28 receptors with B7-1 or B7-2 ligands.

Studies have been revealed that B7-1 and B7-2 promote the development of Th1 cells and Th2 cells respectively (Harris and Ronchese 1999). These co-stimulatory signals provide an outlet for adjuvant immune modulation which can replace the B7-1 or B7-2 ligands (Rhodes 1996). Two main types of effector cells are produced by activated T cells are helper (Th) and cytotoxic (Tc) cells which came from CD4 and CD8 cells.

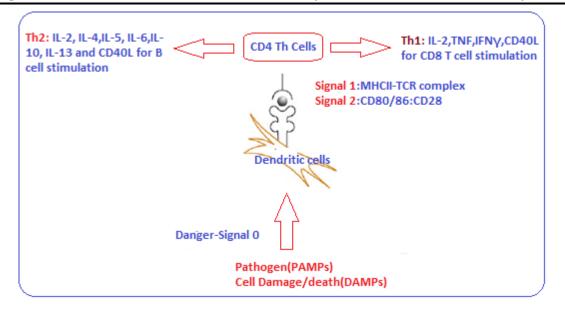


Figure 5: CD4 helper T-cell priming Schematic diagram of signaling during antigen presentation and Th priming.

Besides, either Th1/Th2 or both are activated by activated CD4 cells. Interferon-gamma (IFN- γ), tumor necrosis factor- β (TNF- β) and IL-2 cytokines are produced by Th1 cells and induce cytotoxic T lymphocyte production (CTL). Cytokines IL-4 and IL-10 were produced by Th2 cells that favor the antibody production. The class switch seen in figure 5 (Murphy and Reiner 2002).

5.0 Commonly used vaccine adjuvants

5.1 Aluminum salts

The clinically accepted alum adjuvants are consisted with aluminum hydroxide and aluminum phosphate. Initially, they were assumed to act mainly by building a long-lasting antigen depot and facilitating their absorption by APCs. It is clearly understood that innate immunity plays a primary role in alum's adjuvanticity. [23] Alum is used mainly to stimulate the production of antibodies and does not use TLR in vivo for its function. However, a highly polarized Th2 cell reaction persuades in mice alum in the human response to proteins with alum appears to be a combination of Th1 and Th2 cell. [24] Antibody isotypes were needed for Th2 cell polarization with all protein antigens. Studies *in vitro* employing DCs and macrophages have demonstrated after the lipopolysaccharide (LPS) priming. Inspiration of the Th2 cell promoting cytokines IL-4, IL-25 and IL-6 from innate cells by alum has been planned. The adjuvants also induce the death of necrotic cells and the discharge of uric acid's endogenous threat signal. Indeed, the uric acid injection has been exposed to inhibit the immuno-potentiating activity of intraperitoneal administration of alum. [25]

5.2 Oil in Water Emulsions

AS03 and MF59 are both squalene-based emulsions. Squalene oil that easily metabolized than paraffin oil is used in the MF59. In almost all of Europe, MF59 is authorized in normal flu shots in the elderly, and both have been used within global flu shots. As a result, there's sufficient human research evaluating the influenza vaccine with all these adjuvants to the same vaccine without additives or aluminum. ^[26] Such emulsions induce remarkable memory reactions with a mixed phenotype of Th1 and Th2 cells, allow for lower

dosages and reduce antigen doses, and cause greater antibody responses. With considerable local stimulation, MF59 increases DC recruitment. The sequence of mediated genes elicited by MF59 intramuscular injection is indeed higher and unique from the immune response induced by alum or a TLR9 ligand. [27]

5.3 α**-D-Galactosylceramide**

The glycolipid α -D-galactosylceramide as well as its analogues are class of immune enhancers derived by MHC-I-like compounds. CD1d were identified by the TCRs of natural killer T cells (NKT cells), which are stimulated either by release of IFN- γ or IL-4.^[28] Glyclipids attach to CD1d having low affinity and specificity through hydrphobic connectins to build a complex, as per study, employed dissolved forms of α - β -TCR and CD1d. Utilising rigid interaction surfaces involving the glyclipids carbohydrate moiety and constant 14 chains of the TCR, this CD1d glyclipid complex then attaches to the / TCR with strong affinity (Figure 6). [29]

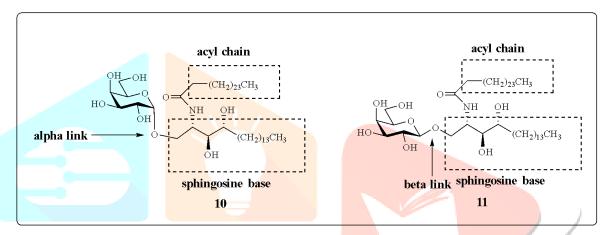


Figure 6: Structure of α and β -D-Galactosylceramide

The galactose molecule's anomeric configuration is crucial for interacting. [30] Upon initial activation, NKT cells become polarised towards the release of IL-4, which aids in the conversion of naive antigen-specific CD4+ T cells leading to Th2 immunity. As a result,α-galactosylceramides may contribute in the induction of Th2 immunity toward external pathogens or even in the defence against chronic illnesses caused by Th1 pathogenic immunity. [31]

5.4 Saponins

Saponins are steroids or triterpenoid glycosides found in natural and agricultural plants, lower marine creatures, and certain microbes. [32] Saponins, which have one or even more sugar molecules connected to them, hold a triterpenoid or steroidal aglycone. These are commonly found in the plant kingdom, mainly in processed crops, with triterpenoid saponins predominant. Many legumes, including peas, maize, soybeans, lucerne, etc., as well as spinach, tea, sugar beet, sunflower, licorice, quinoa, horse chestnut, and ginseng, have been found to contain steroid saponins, as per Fenwick. Oats, tomato seeds, aubergines, alliums, capsicum peppers and ginseng also contain steroidal saponins. Saponins are tensioactive glycosides covering the triterpenoid form of a hydrophobic nucleus with nucleus-bound carbohydrate chains.

5.5 Quillaja saponin (QS-21) and Quill

The adjuvant activity was observed in the bark extract of the *Quillaja saponaria* tree. The extract comprises some compounds that have been isolated and confirmed to have distinct adjuvant properties. Quillaja and other saponins have been reported to improve *in vitro* immune cell proliferation either as crude mixtures or as purifying compounds. Purified Quillaja saponins also improved the production of antibodies without producing any regional antibodies. Campbell and Chavali observed that in spleen and mesenteric lymph nodes, pests fed Quillaja saponins showed enhanced and long-drawn-out natural killer (NK) cell involvement in expanded cell proliferation. The saponins engage directly with T - lymphocytes of mucosal immune cells, promoting the production of soluble factors. According to other study, Quillaja saponins have mitogenic properties and enhance B and T cellular proliferation. [34] QS-21 was initially investigated as just a substitute for alum when a certain vaccination required significant cellular functions. Adjuvants are exogenous active ingredients which adhere to hydrophobic surfaces that convey immunomodulatory characteristics via cell surface-free energy. Water insoluble motifs are responsible for the outward reactive characteristics of QS-21 and Quill. For veterinary applications, Quill has been used productively. It's a natural compound that's constituted of even more than 23 saponins and is commonly hazardous to human health.

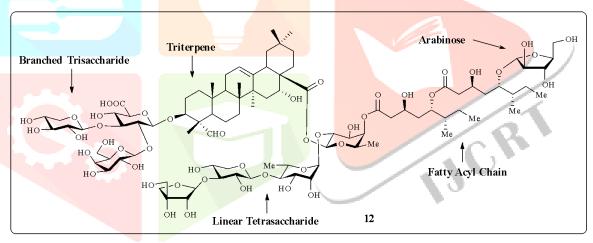


Figure 7: Structure of potent QS-21 (12)

Toxicity includes severe hemolysis, which reflects the similarity of cholesterol saponins found in erythrocyte membranes, in addition to harsh local reactions. As a result of membrane solubilization and hemolysis, alienated fractions have been tested. Saponin, QS-21, and QS-7 have been found to reduce toxicity increased antigen-specific IgG responses which were experiential compared to antigen alone in intradermal antigen-vaccinated mice, and 20 μg of QS-7, QS-17, QS-21 or QS-18. QS-21 is less poisonous than Quill itself. QS-21 is a potent adjuvant for the release of Th1 cytokine-induced CTL (IFN-γ and IL-2) and IgG2a isotype antibodies. According to Evans, QS-21 was capable of permitting considerable dosage reductions for HIV-1 antigen and also increased proliferative T-cell activation, but not CTL, in a testing with the antigen. Following both universal and mucosal management, QS-21 has been preserved to function as a DNA vaccine adjuvant. Quillaja saponins were restricted to be used in human vaccination due to their high toxicity, hemolytic effect, and instability (Figure 7).^[37]

5.6 Monophosphoryl lipid A

Figure 8: Structure of Monophosphoryl lipid A (13)

Monophosphoryl lipid (MPL) was reported to exhibit potent adjuvanticity and no effect of toxicity. The MPL's structural activity relationship showed that adjuvanticity includes a hexaacylated β (1 - 6) diglucosamine with three 3-alkanoyloxytetradecanoyl residues or six fatty acid groups.^[38] MPL's adjuvant characteristics were investigated, and it was discovered that this is an excellent adjuvant in both humoral and cell-mediated immune activation. In the global and mucosal compartments of the immune system, MPL potentially induce simultaneous Th1 and Th2 responses. MPL was approved for allergic treatment in European countries because of its ability to minimize Th2 regulatory responsiveness to allergens. ^[39] MPL has been given to thousands of healthy, well-tolerated, and potent adjuvant constituents of individuals in several formulations (Figure 8).

5.7. Muramyl dipeptide (MDP)

Flower and Jeanlozs reported the first N-acetylmuramic acid synthesis in 1963.^[40] MDP is an important adjuvant for improving the potency of vaccines and medications. The expression surface markers essential for cell adhesion and presentation of antigen l, thus increasing anti-microbial and phagocytic activity promoting cytotoxicity mediated antibody.^[41] MDP or other muropeptides also elicit immunological responses, promoting cell differentiation and proliferation, by producing IFN- γ as well as other cytokines.

Figure 9: Structure of Muramyl dipeptide (MDP) (14)

The separation of white blood cells plays an integral function in protection against foreign intruders. [42] In general, three sub-units, N-acetyl-D-glucosamine, lactic acid, and dipeptide, will synthesize MDP **14** by coupling reaction. The N-acetylmuramic acid molecule is mainly described in a synthesis. MDP does have the (*R*)-configuration of lactic acid, while N-acetylisomuramyl-L-alanyl-D-isoglutamine does have the (*S*)-configuration. An adjuvanticity of the (*S*)-isomer in the formation of delayed-type N-acetyl-3-(4-arsonophenylazo)-L-tyrosine hypersensitivity in animals was decreased drastically in a biological investigation. It is important to note that certain chirality seems to have an effect on molecule stabilization as well as biological properties. MDP has also been proven to be the minimal structure essential for cellular stimulation both in vitro and in vivo. MDP was introduced to a synergistic effect with LPS (lipopolysaccharides) present in Gram-negative bacteria outer membranes and considered to be the TLR4 cell surface receptor. This synergy was found in vitro in whole blood human primary cells, peripheral blood mononuclear cells (PBMCs), numerous human monocytes, and rodent cell lines were purified in vivo in a rat anorexia model (Figure 9). [43]

5.8 Other particulate adjuvants

While they have earned less importance to date, several other particulate adjuvants are mentioned.

- Calcium salts; Calcium salts have less immunomodulatory properties but are safe to use in humans. [44]
- **Proteosomes;** They may not have an immunomodulatory activity, but they provide excellent immunogen presentation and targeting with minimal CTL activation. [45]
- Virosomes; "virus-derived trans-membrane proteins" e.g. influenza haemagglutinin. [46]
- Stearyl tyrosine; these adjuvants forms hydrophilic protein complex that functions as a medium-term depot and activates responses to Thl. Also, its safety profile is reportedly good. [47]
- Algammulin. In clinical testing, a combo of aluminium hydroxide and –inulin with combined characteristics were discovered. [48]

6.0 Features of an ideal adjuvant

Because every adjuvant and its specific antigen have its unique requirements, a "perfect" adjuvant is unlikely to exist. [49] The generic qualities listed below, however, would just be preferable. Yet, no adjuvant has achieved most of these objectives.

- Chemically and biologically, it must be established that there shouldn't be lot-to-lot variation in the formulated product so that maintaining stable vaccine responses between studies and over time.
- It must also be disposable or quickly excreted by the kidneys once its adjuvant effect has worn off to prevent the danger of long-term negative consequences. Combined with the antigen, it can induce a more intense defensive or therapeutic immune response than when the antigen is given alone.
- That must be risk-free, with no immediate or long-term adverse effects.
- It should be possible to achieve potency with few doses and/or lower antigen concentrations. To be economically and clinically beneficial, it must be shelf durable.

b435

The adjuvant should be cost-effective.

7.0 Summary of rational adjuvant development

There is an intense and increasing interest in the discovery of vaccine adjuvants because many of the current candidates for vaccine subunits lack adequate immunogenicity to be clinically effective. The production of adjuvants for vaccines is now moving from 'reality to hypothesis' in the area of immunology. The logical conception of effective vaccine adjuvants has been enabled by our enhanced understanding of the innate immune system and the molecular pathways involved in the antigen-specific immune response. Potent immunoadjuvants are particularly important in the form of novel recombinant antigens and also to overcome the poor immunogenicity of existing antigens include carbohydrates, with the right formulation become the basis of an effective immunization technique. As a result, a better and more effective adjuvant susceptible of strengthening both body's immune responses is desperately needed in clinical care.^[50]

8.0 Reference

- [1] D. Mullin, The Journal of allergy and clinical immunology **2003**, 112, 810-814.
- [2] J. A. McCullers, J. D. Dunn, P T 2008, 33, 35-41.
- [3] A. Adamo, R. L. Beingessner, M. Behnam, J. Chen, T. F. Jamison, K. F. Jensen, J. C. Monbaliu, A. S. Myerson, E. M. Revalor, D. R. Snead, T. Stelzer, N. Weeranoppanant, S. Y. Wong, P. Zhang, *Science* **2016**, *352*, 61-67.
- [4] A. T. Glenny, B. E. Hopkins, *Br J Exp Pathol* **1923**, *4*, 283-288.
- [5] S. Plotkin, *Proc Natl Acad Sci U S A* **2014**, *111*, 12283-12287.
- [6] S. H. E. Kaufmann, Frontiers in Immunology 2019, 10.
- [7] A. T. Glenny, J. Pathol. Bacteriol. 1926, 29, 38-45.
- [8] S. E. Girardin, L. H. Travassos, M. Hervé, D. Blanot, I. G. Boneca, D. J. Philpott, P. J. Sansonetti, D. Mengin-Lecreulx, *The Journal of biological chemistry* **2003**, 278, 41702-41708.
- [9] S. Kumar, A. Roychowdhury, B. Ember, Q. Wang, R. Guan, R. A. Mariuzza, G. J. Boons, *The Journal of biological chemistry* **2005**, *280*, 37005-37012.
- [10] Y. Kitaura, O. Nakaguchi, H. Takeno, S. Okada, S. Yonishi, K. Hemmi, J. Mori, H. Senoh, Y. Mine, M. Hashimoto, *Journal of Medicinal Chemistry* **1982**, 25, 335-337.
- [11] Y. Fujimoto, A. R. Pradipta, N. Inohara, K. Fukase, *Natural Product Reports* **2012**, 29, 568-579.
- [12] A. REITERMANN, J. METZGER, K.-H. WIESMÜLLER, G. JUNG, W. G. BESSLER, **1989**, *370*, 343-352.
- [13] J. H. Fritz, R. L. Ferrero, D. J. Philpott, S. E. Girardin, *Nature Immunology* **2006**, *7*, 1250-1257.
- [14] H. A. Youssef, T. A. Omar, H. A. Fouad, S. M. El-Sheikh, S. El Achy, M. M. Afifi, *Alexandria Dental Journal* **2018**, *43*, 65-69.
- [15] N. Siegmund-Schultze, *Dtsch Arztebl International* **2007**, *104*, A-1072.
- [16] B. Lemaitre, E. Nicolas, L. Michaut, J. M. Reichhart, J. A. Hoffmann, *Cell* **1996**, *86*, 973-983.
- [17] T. Kawai, S. Akira, *Nat Immunol* **2010**, *11*, 373-384.

- P. Henneke, S. Dramsi, G. Mancuso, K. Chraibi, E. Pellegrini, C. Theilacker, J. Hübner, S. Santos-Sierra, G. Teti, D. T. Golenbock, C. Poyart, P. Trieu-Cuot, *Journal of immunology (Baltimore, Md. : 1950)* **2008**, *180*, 6149-6158.
- [19] S. Rakoff-Nahoum, R. Medzhitov, *Nature reviews. Cancer* **2009**, 9, 57-63.
- [20] Y. Gao, H. Xiao, Y. Wang, F. Xu, Medicine (Baltimore) 2017, 96, e6822-e6822.
- [21] A. Aderem, R. J. Ulevitch, *Nature* **2000**, *406*, 782-787.
- [22] R. Ahmed, D. Gray, Science **1996**, 272, 54-60.
- [23] A. M. Didierlaurent, S. Morel, L. Lockman, S. L. Giannini, M. Bisteau, H. Carlsen, A. Kielland, O. Vosters, N. Vanderheyde, F. Schiavetti, D. Larocque, M. Van Mechelen, N. Garçon, *The Journal of Immunology* 2009, 183, 6186-6197.
- [24] M. L. Mbow, E. De Gregorio, N. M. Valiante, R. Rappuoli, *Current Opinion in Immunology* **2010**, 22, 411-416.
- [25] B. N. Lambrecht, M. Kool, M. A. M. Willart, H. Hammad, Current Opinion in Immunology 2009, 21, 23-29.
- [26] P. Marrack, A. S. McKee, M. W. Munks, *Nature Reviews Immunology* **2009**, *9*, 287-293.
- [27] F. Mosca, E. Tritto, A. Muzzi, E. Monaci, F. Bagnoli, C. Iavarone, D. O'Hagan, R. Rappuoli, E. De Gregorio, Proceedings of the National Academy of Sciences 2008, 105, 10501-10506.
- [28] N. Singh, S. Hong, D. C. Scherer, I. Serizawa, N. Burdin, M. Kronenberg, Y. Koezuka, L. Van Kaer, *The Journal of Immunology* **1999**, *163*, 2373-2377.
- [29] S. Sidobre, O. V. Naidenko, B.-C. Sim, N. R. J. Gascoigne, K. C. Garcia, M. Kronenberg, *The Journal of Immunology* **2002**, *169*, 1340-1348.
- [30] L. Brossay, O. Naidenko, N. Burdin, J. Matsuda, T. Sakai, M. Kronenberg, The Journal of Immunology 1998, 161, 5124-5128.
- [31] K. J. L. Hammond, D. I. Godfrey, *Tissue Antigens* **2002**, *59*, 353-363.
- [32] R. Riguera, Journal of Marine Biotechnology 1997, 5, 187-193.
- [33] A. M. Mowat, R. E. Smith, A. M. Donachie, E. Furrie, D. Grdic, N. Lycke, *Immunology letters* **1999**, *65*, 133-140.
- [34] N. E. Jacobsen, W. J. Fairbrother, C. R. Kensil, A. Lim, D. A. Wheeler, M. F. Powell, *Carbohydrate Research* **1996**, 280, 1-14.
- [35] A. M. Mowat, A. M. Donachie, G. Reid, O. Jarrett, *Immunology* **1991**, 72, 317-322.
- [36] A. C. Allison, N. E. Byars, *Molecular Immunology* **1991**, 28, 279-284.
- [37] C. R. Kensil, U. Patel, M. Lennick, D. Marciani, *The Journal of Immunology* **1991**, *146*, 431-437.
- [38] J. Ismaili, J. Rennesson, E. Aksoy, J. Vekemans, B. Vincart, Z. Amraoui, F. Van Laethem, M. Goldman, P. M. Dubois, *The Journal of Immunology* **2002**, *168*, 926-932.
- [39] A. W. Wheeler, S. R. Woroniecki, Expert Opinion on Biological Therapy 2004, 4, 1473-1481.
- [40] B. Loev, M. F. Kormendy, *The Journal of Organic Chemistry* **1963**, 28, 3421-3426.

1JCR

- [41] G. R. Dreesman, Y. Sanchez, I. Ionescu-Matiu, J. T. Sparrow, H. R. Six, D. L. Peterson, F. B. Hollinger, J. L. Melnick, *Nature* **1982**, *295*, 158-160.
- [42] V. Souvannavong, S. Brown, A. Adam, *Cellular Immunology* **1990**, *126*, 106-116.
- [43] J. E. Wang, P. F. Jørgensen, M. Almlöf, C. Thiemermann, S. J. Foster, A. O. Aasen, R. Solberg, *Infection and immunity* **2000**, *68*, 3965-3970.
- [44] E. A. Miao, C. M. Alpuche-Aranda, M. Dors, A. E. Clark, M. W. Bader, S. I. Miller, A. Aderem, *Nature Immunology* **2006**, *7*, 569-575.
- [45] T. H. Mogensen, S. R. Paludan, Journal of Molecular Medicine 2005, 83, 180-192.
- [46] A. B. Molofsky, B. G. Byrne, N. N. Whitfield, C. A. Madigan, E. T. Fuse, K. Tateda, M. S. Swanson, *J Exp Med* **2006**, *203*, 1093-1104.
- [47] M. Morr, O. Takeuchi, S. Akira, M. M. Simon, P. F. Mühlradt, European Journal of Immunology 2002, 32, 3337-3347.
- [48] K. M. Murphy, S. L. Reiner, *Nature Reviews Immunology* **2002**, 2, 933-944.
- [49] T. Nishiya, A. L. DeFranco, *Journal of Biological Chemistry* **2004**, 279, 19008-19017.
- [50] S. Okahira, F. Nishikawa, S. Nishikawa, T. Akazawa, T. Seya, M. Matsumoto, *DNA and cell biology* **2005**, 24, 614-623.