



# Detection of Phospholipase Activity in Clinical Isolates of *Candida* species in a Tertiary Care Hospital

1Asim Sarkar, 2Sharvari Samant

1Post Graduate Student, 2Associate Professor

1MGM Institute of Health Sciences, Navi Mumbai,

2MGM Institute of Health Sciences, Navi Mumbai

**Abstract:** *Candida* is a yeast-like fungus and omnipresent in the environment, which makes it one of the most common pathogens for opportunistic infections. *Candida* usually causes mucosal Candidiasis but may also cause systemic and disseminated infections in debilitated patients. Phospholipase production has been reported to be one of the virulence factors in *Candida*. This study was carried out to isolate various species of *Candida* from different clinical specimens and to check their Phospholipase activity. All the clinical specimens suspected of fungal aetiology were analysed and subjected to Microbiological identification of *Candida* species. *Candida* species were subjected to Phospholipase activity by Egg Yolk Agar Plate method. A total of 7 varieties of *Candida* species ( *C. albicans* 25.45%, *C. tropicalis* 32.72%, *C. krusei* 12.72%, *C. kefyr* 12.72%, *C. dubliniensis* 10.90%, *C. glabrata* 3.63%, *C. utilis* 1.81%) were isolated from various clinical samples. Out of total 54 isolates, 29(53.70%) showed extensive Phospholipase activity.

**Keywords:** *Candida albicans*, *Candida-non-albicans*, Phospholipase activity, Candidiasis, Egg Yolk.

**Introduction:** Yeasts are omnipresent in the environment and some belong to the normal human flora. *Candida* is a yeast like fungus and is one of the normal flora of human body residing mostly in vagina, mouth, intestinal tract, skin and other mucous membranes. It changes in the internal and external environment from a completely harmless saprophyte into a virulent pathogen especially in immunocompromised individuals. Morphologically *Candida* are oval yeast cells, with pseudo-hyphae. When grown in the laboratory, *Candida* appears as large, round, white or cream colonies with a yeasty odour on agar plates. *C. albicans* ferments glucose and maltose producing acid and gas, sucrose with acid and without gas production and fails to ferment lactose which helps it to distinguish *C. albicans* from other *Candida* species.

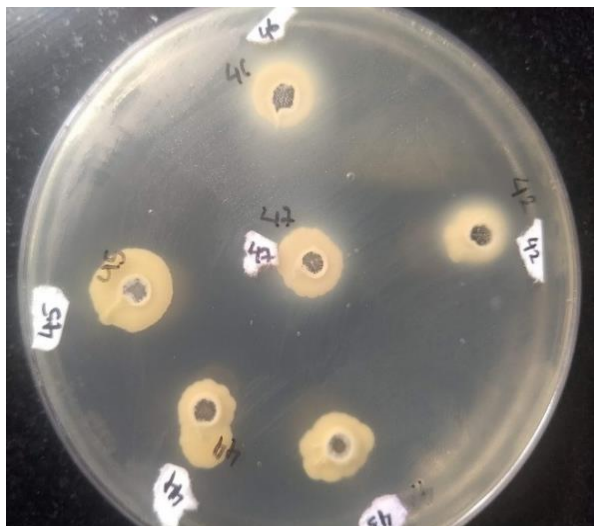
Although most infections of Candidiasis are caused by *C. albicans*, there has been striking increase in the frequency with *non-albicans Candida* species in last few years as well.<sup>[1]</sup> There has been an increase in treatment failure because of drug resistance.<sup>[2]</sup> It can also be attributed to some virulence factors. These putative virulence factors include secretion of proteinase, adherence to host cell surface, germination, and Phospholipases, though not even a single putative factor has been clearly reported with pathogenicity.<sup>[3]</sup> There are reports showing Phospholipase activity as one of the important virulence factors of *Candida* species.<sup>[4]</sup> There is study showing co-relation between Phospholipase activity with mucosal invasion<sup>[5]</sup>. *Candida* isolates producing greater number of Phospholipases leading to increased mortality in rat models which has been reported<sup>[6]</sup>.

Phospholipases contribute to pathogenicity of *bacteria*<sup>[7]</sup>, *protozoa*<sup>[8]</sup>, and *rickettsiae*<sup>[9]</sup> the role of these membrane damaging enzymes is proposed in the pathogenicity of *C. albicans* as well. Other than Phospholipase, Secreted Aspartyl Proteinases, Germ tube formation, adherence to host tissue and phenotypic switching are also factors contributing to the virulence of *Candida*. Both hydrolytic enzymes cause cellular damage. Different species of *Candida* produce varying degree of Phospholipases; hence speciation of the *Candida* till species level becomes important. *Vidotto et al.*<sup>[10]</sup> have reported that the correlation between Phospholipase activity and high Germ tube formation can facilitate the mucosa penetration. A comparison between the level of production of Phospholipase of *C. albicans* isolated from cases of infections and commensals<sup>[11]</sup> has been described.

Therefore, the present study was planned to isolate *Candida spp.* in various clinical specimens and to detect their Phospholipase activity

**Material and Methods:** This study was carried out in a period of 1 year from October 2018-October 2019 in the Department of Microbiology, MGM Medical College and Hospital, Kamothe, Navi Mumbai, India. A total of 50 *Candida* were isolated from 167 clinical specimen suspected of fungal aetiology. All the samples were processed as under:

1. Microscopy
  - a. KOH wet mount- For rapid detection of Pseudo-hyphae and budding yeast cell and to dissolve the organic matter such as Hair and Nail.
  - b. Grams staining- for identification of budding yeast cells with or without Pseudo-hyphae.
2. Culture on SDA- The specimens were then inoculated onto Sabouraud Dextrose Agar (SDA- Hi-media) and incubated at 37°C for 24-48 hours and observed for typical colonies of *Candida species*.
3. Species identification
  - a. Urea hydrolysis- To rule out the possibility of *Cryptococcus*
  - b. Germ tube (GT) test- To differentiate between *albicans* and *non-albicans* group.
  - c. Sugar assimilation<sup>[12]</sup>- Yeast Nitrogen plates (Hi media laboratories) were made and flooded with yeast suspension, air dried and then sugar impregnated discs (Hi-media laboratories) were put and incubated 37°C for 24-48 hours and observed for the hazy zones around.
4. Speciation by Chrom Agar- Final step in the species identification by distinct colour formation on CHROM agar plates (HiCHROM by Hi media laboratories) were used.
5. Demonstration of extracellular Phospholipase activity- Extracellular Phospholipase activity was determined by a method described Price *et al.*<sup>[13]</sup> that is Egg yolk agar Plate method. All the isolates were then subjected for Phospholipase production, by creating wells (keeping the diameter same) onto the plates, as it gives better infusibility, 10µ of the yeast suspension was inoculated and incubated at 37°C for 4 days and observed for precipitation zone around the colonies. Egg yolk agar was prepared by 6.5gms of SDA, 5.84gms of NaCl and 0.55gms of CaCl<sub>2</sub> dissolved in 98ml of distilled water and sterilised by autoclaving at 121°C for 15 minutes. Egg yolk was separated from Egg white and centrifuged at 4500 rpm for 45 minutes, 2ml of supernatant was then added to cooled medium (45-50°C).



Phospholipase Activity



Candida spp. on Chrome Agar

**Results and Observations:** Microbiological analysis of all the specimens received for fungal cultures were processed and 54 isolates of yeasts were identified for further study. The isolation was done from various specimens including Sputum, Urine, Blood, High Vaginal Swab (HVS), Pus, Nail clipping and Foley's tip. It was also correlated with duration of infection as well as the Phospholipase activity of each isolate.

Fig. 1: Rate of Isolation of various Candida species in the Tertiary Care Hospital

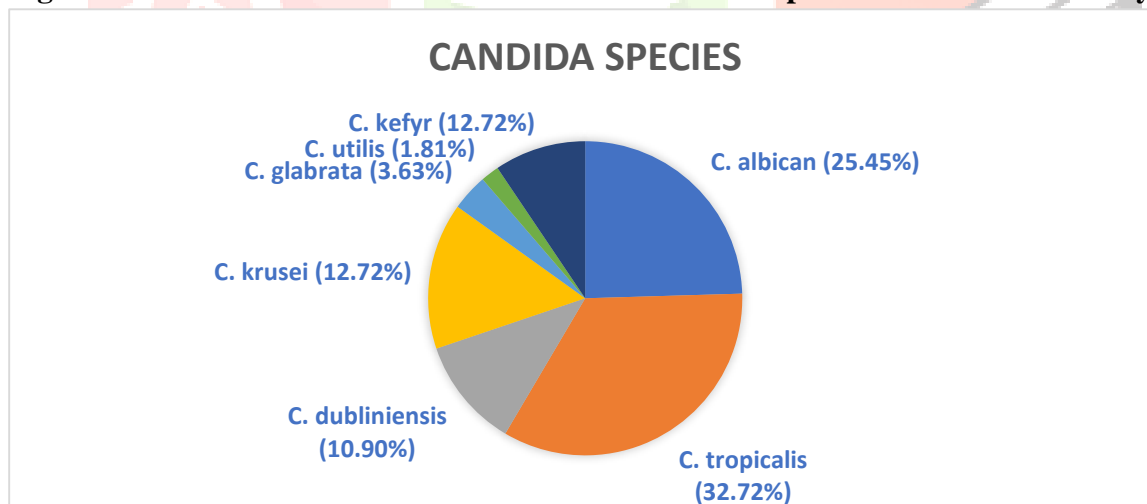
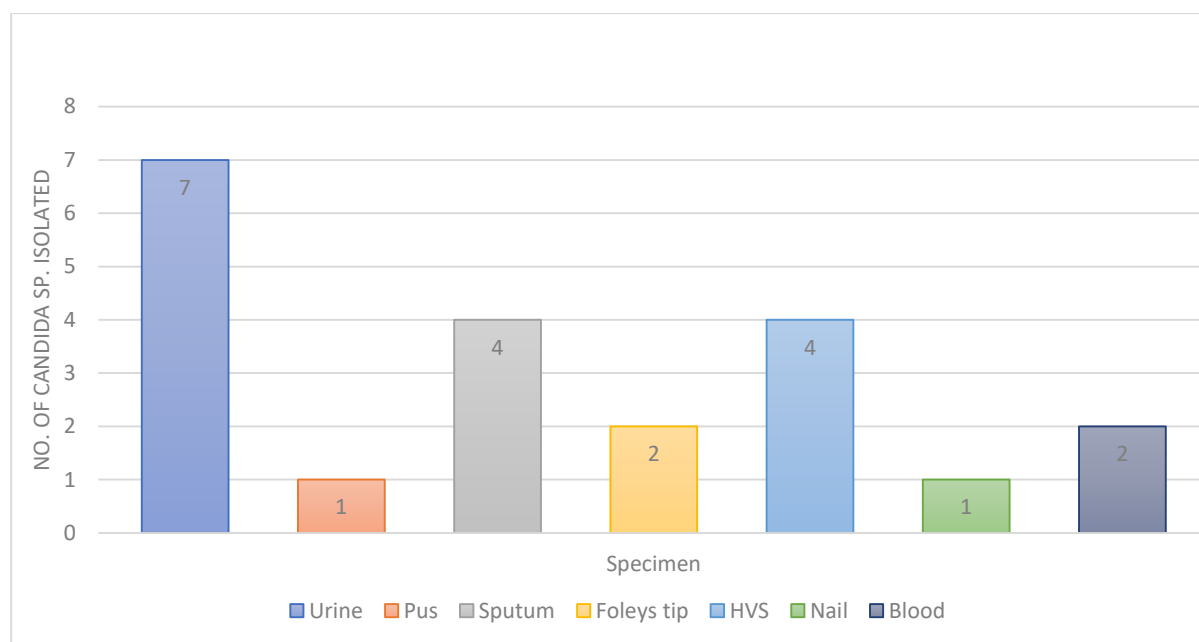


Figure 1 depicts the no. of species isolated and its percentage with respect to the total no. of *Candida* isolated. *Candida tropicalis* (32.72%) was found to be the predominant species whereas as *Candida utilis* (1.81%) was found to be the least common species in this study.

**Graph 1: Variety of *Candida* species isolated in Various Specimens-**

The above Graph depicts rate of isolation of various species of *Candida* in various clinical specimens. A total of 7 varieties of *Candida* were isolated from Urine whereas only 1 type of species were isolated from Pus and Nail Clipping.

All the isolates were also subjected to check Phospholipase activity. The Phospholipase activity was calculated in terms of Pz Value which was determined by the following formula<sup>[13]</sup>:

$$Pz = \frac{\text{Colony diameter}}{\text{Colony diameter} + \text{Zone of precipitation}}$$

The Pz value was classified in following based on intensity:

Pz value	Intensity
1 (Negative)	0
<0.90-0.99	+
0.80-0.89	++
0.70-0.79	+++
<0.70	++++

It was found that 29 (53.70%) isolates were positive for Phospholipase activity and all showed Strong Phospholipase activity i.e., Pz value <0.70, whereas the rest 25 (46.29%) were negative for phospholipase activity.

**Table 1: Average Pz value with Mean Percentage of Various *Candida species* Isolated from Various Specimens-**

<i>Candida species</i>	Average Pz Value	Mean %
<i>C. albicans</i>	0.0442	4.42
<i>C. tropicalis</i>	0.046	4.65
<i>C. dubliniensis</i>	0.045	4.5
<i>C. krusei</i>	0.067	6.73
<i>C. kefyr</i>	0.041	4.1
<i>C. utilis</i>	0.055	5.5
<i>C. glabrata</i>	0.00	0

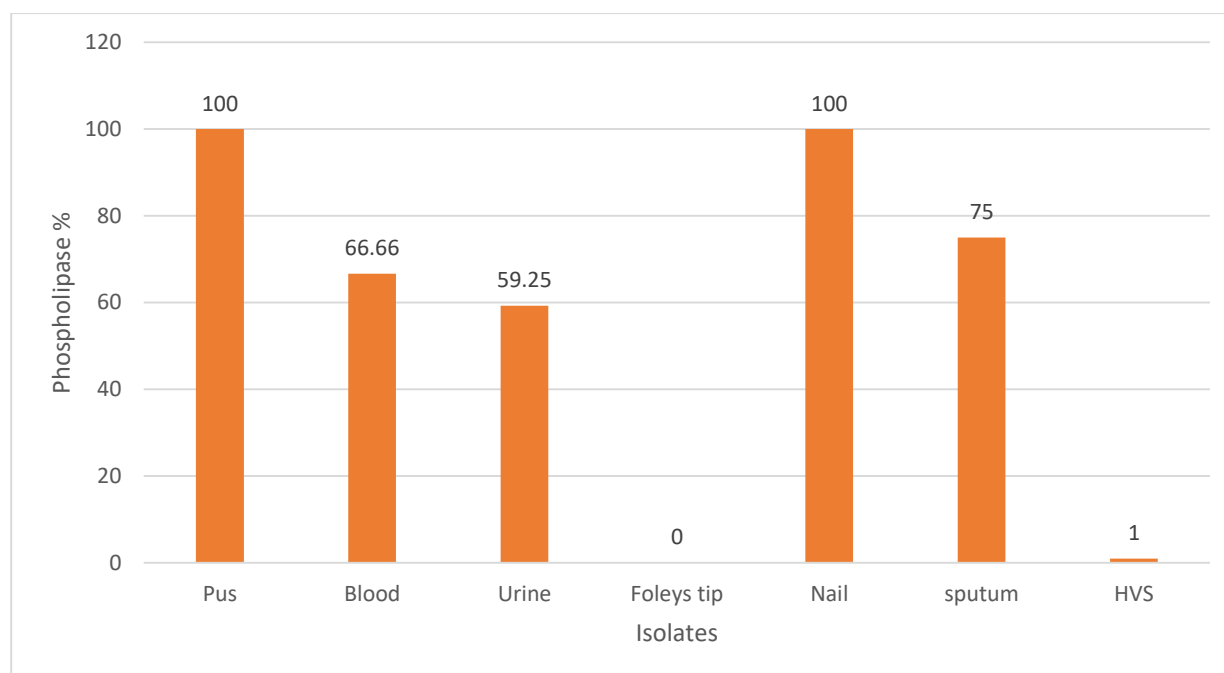
It was found that 29 of 54 isolates were positive for Phospholipase activity and they all showed strong Phospholipase activity i.e., Phospholipase positive (Pz <0.70).

The above Table shows that Pz value of *Candida* isolates range between 0.041-0.067. Highest activity was recorded in the isolates of *C. krusei* and lowest in *C. kefyr*. The commonly isolated *Candida species* were *C. albicans* and *C. tropicalis* which showed Phospholipase activity in the range of 0.44-0.046.

**Table 2: Phospholipase Activity of Various *Candida species*:**

<i>Candida species</i>	Total isolate	Phospholipase (+)	Percentage (%)
<i>Candida albicans</i>	13	8	69.23
<i>Candida tropicalis</i>	18	12	66.66
<i>Candida dubliniensis</i>	6	1	16.66
<i>Candida krusei</i>	8	3	37.5
<i>Candida kefyr</i>	5	3	60
<i>Candida utilis</i>	1	1	100
<i>Candida glabrata</i>	2	0	NA

The Table 2 shows percentage of Phospholipase activity in different species of *Candida*. It was observed that both *Candida tropicalis* and *Candida albicans* were potential Phospholipase producers with more than 60% isolates showing Phospholipase production.

**Graph 2:Percentage of Phospholipase Activity in *Candida* Isolated from Various Clinical Samples.**

Majority of *Candida* isolated from Nail, Pus and Sputum samples showed phospholipase activity.

**Discussion:** *Candida albicans* is the most common cause of Candidiasis, whereas Other *Candida species* are now replacing *C. albicans* in recent years. The extracellular Phospholipase has been known for its pathogenicity causing membrane disruption and colonisation, which was even reported in few cases [5]. The state of host is of primary importance in determining *Candida* Pathogenicity [14]. This study was done on various clinical specimens isolated from suspected case of Candidiasis.

In this study maximum *Candida* were isolated from cases of Urinary tract infection (UTI) that is from Urine (56%) followed by Sputum (20%), High Vaginal Swab (8%), Blood (6%), Tip and Pus (8%) and Nail (2%). It shows that *Candida* is major cause of UTI followed by Respiratory Infections and vulvovaginal infections. Similar findings were reported by Das Mohandaset al. [4]

In majority of the studies done across the Globe showed *C. albicans* as the major pathogen. (4,15) However in the present study we found *C. tropicalis* as most frequently isolated species. In a study by Sudhason Ye [16] and Shivprakasha [17] other *Candida species* were more predominant. This shows that *non-albicans-candida* was more prevalent in our set up.

This study shows the rate of isolation of various species of *Candida*, in which *Candida albicans* was 14 (25.45%), whereas *Candida tropicalis* was 18 (32.72%). Amongst the *non-albicans species*, *C. tropicalis* 18 (32.72%) was the most predominant followed by *C. krusei* 7 (12.72%) and *C. kefyr* 7 (12.72%), *C. dubliniensis* 6 (10.90%), *C. glabrata* 2 (3.63%) and *C. utilis* 1 (1.81%). The ratio of *C. albicans* to *C. non-albicans* group was found to be 2.8:11 in this study, whereas in the study done by SR Fule [14], the ratio was found to be 1.68:1, which is much lesser than what we have got in our study.

There are many reports describing the features of *C. albicans* with probable virulence factors that enables the organism to cause infection in a susceptible host. [14] A study has been done on Role of Phospholipase in virulence and fungal pathogenesis. [18] The highest phospholipase intensity was seen in *C. albicans*, whereas it was minimal in *C. krusei*. All the clinical isolates in this study showed either intense Phospholipase activity or no Phospholipase activity at all, which confers its virulence and pathogenicity as an opportunistic pathogen. Lower is the Pz value, more is the enzymatic activity, and more is the pathogenicity.

The average Pz value ranged from 0.041-0.067. *C. albicans*, *C.dubliniensis* and *C. tropicalis* were found nearly equal in terms of Phospholipase activity this could be due to varying no. of species isolated. More than 60% of *C.albicans* and *C. tropicalis* isolates were Phospholipase producers. In this study, 9 of 13 (69.23%) *C. albicans* showed the enzymatic activity whereas, *C.tropicalis* 12 of 18 (66.66%), 1 isolate of *C. utilis*, *C. dubliniensis* 1 of 6 (16.66%), *C. krusei* 3 of 8 (37.5%) and *C. kefyr* 3 of 5 (60%) and in *C. glabrata* none of the strains showed Phospholipase activity. *Albicans* and *non-albicans Candida* showed this activity, whereas as Fule SR<sup>[14]</sup> reported that none of the *non-albicans* group showed the Phospholipase activity. Another study by Vinitha M<sup>[4]</sup> showed that Phospholipase activity was shown by both *albicans* (46.93%) and *non-albicans* (42%) group which correlates with our study.

One important observation noted in this study was 69.23 % *C. albicans* (n= 13) and 66.66% of *C.tropicalis* (n=18) were Phospholipase positive. This indicates that though non albicans *Candida* were isolated in higher number as compared to *Candida albicans*, phospholipase activity was the common factor contributing to their pathogenicity.

**Conclusion:** This study was carried out for a period of one year with 50 isolates of *Candida* due to time constrain. However, it clearly highlights the fact that Phospholipase activity could be one of the important factors for pathogenicity of *Candida*. The Authors would like to take this study further to study the effect of phospholipase activity on prognosis of infection.

## References:

1. Falagas ME, Roussos N, Vardakas KZ. Relative frequency of albicans and the various non-albicans *Candida* spp among candidemia isolates from inpatients in various parts of the world: a systematic review. *International Journal of Infectious Diseases*. 2010 Nov 1;14(11):e954-66.
2. Sanguinetti M, Posteraro B, Lass-Flörl C. Antifungal drug resistance among *Candida* species: mechanisms and clinical impact. *Mycoses*. 2015 Jun;58:2-13.
3. Yang YL. Virulence factors of *Candida* species. *Journal of Microbiology Immunology and Infection*. 2003 Dec;36(4):223-8.
4. das Mohan V, Ballal M. Proteinase and phospholipase activity as virulence factors in *Candida* species isolated from blood. *Revistaiberoamericana de micologia*. 2008 Dec;25(4):208-10.
5. BARRETT-BEE KE, HAYES Y, WILSON RG, RYLEY JF. A comparison of Phospholipase activity, cellular adherence and pathogenicity of yeasts. *Microbiology*. 1985 May 1;131(5):1217-21.
6. Ibrahim AS, Mirbod F, Filler SG, Banno Y, Cole GT, Kitajima Y, Edwards JE, Nozawa Y, Ghannoum MA. Evidence implicating Phospholipase as a virulence factor of *Candida albicans*. *Infection and immunity*. 1995 May 1;63(5):1993-8.
7. Wright GC, Weiss J, Kim KS, Verheij H, Elsbach P. Bacterial phospholipid hydrolysis enhances the destruction of *Escherichia coli* ingested by rabbit neutrophils. Role of cellular and extracellular Phospholipases. *The Journal of clinical investigation*. 1990 Jun 1;85(6):1925-35.
8. Silverman DJ, Santucci LA, Meyers NA, Sekeyova ZU. Penetration of host cells by *Rickettsia rickettsii* appears to be mediated by a Phospholipase of rickettsial origin. *Infection and immunity*. 1992 Jul 1;60(7):2733-40.
9. Saffer LD, Krug SA, Schwartzman JD. The role of Phospholipase in host cell penetration by *Toxoplasma gondii*. *The American journal of tropical medicine and hygiene*. 1989 Feb 1;40(2):145-9.
10. Vidotto V, Koga-Ito CY, Milano R, Fianchino B, Pontón J. Correlation between germ tube production, Phospholipase activity and serotype distribution in *Candida albicans*. *Revistaiberoamericana de micologia*. 1999 Dec;16(4):208-10.
11. McCool L. The Discovery and Naming of *Candida albicans*.

12. Mukhia RK. Virulence factors molecular characterization and clinical correction of candida species isolated from various specimens in a tertiary care hospital.2016;53-55.
13. Price MF, Wilkinson ID, Gentry LO. Plate method for detection of Phospholipase activity in *Candida albicans*. *Sabouraudia: Journal of Medical and Veterinary Mycology*. 1982 Jan 1;20(1):7-14.
14. Fule SR, Das D, Fule RP. Detection of Phospholipase activity of *Candida albicans* and non *albicans* isolated from women of reproductive age with vulvovaginal Candidiasis in rural area. *Indian journal of medical microbiology*. 2015 Jan 1;33(1):92.
15. Sharma Y, Chumber SK, Kaur M. Studying the prevalence, species distribution, and detection of in vitro production of Phospholipase from *Candida* isolated from cases of invasive Candidiasis. *Journal of global infectious diseases*. 2017 Jan;9(1):8.
16. YeSudhaSon BL, MohanraM K. *Candida tropicalis* as a predominant isolate from clinical specimens and its antifungal susceptibility pattern in a tertiary care hospital in Southern India. *Journal of clinical and diagnostic research: JCDR*. 2015 Jul;9(7):DC14.
17. Shivaprakasha S, Radhakrishnan K, Karim PM. *Candida* spp. other than *Candida albicans*: a major cause of fungaemia in a tertiary care centre. *Indian journal of medical microbiology*. 2007 Oct 1;25(4):405.
18. Ghannoum MA, Abu-Elteen KH. Pathogenicity determinants of *Candida*. *Mycoses*. 1990 Jun;33(6):265-82.

