



COMPARATIVE STUDY OF BANANA PEEL AND BLACK TEA EXTRACTS AGAINST PERIODONTITIS CAUSING *Staphylococcus aureus*

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Abstract: Black tea and banana (*Musa paradisiaca*) is to contribute natural remedies and useful in curing bacterial infections. Banana plants (*Musa paradisiaca*) are grown in worldwide and all parts of it have medicinal application. The extract of banana peels and black tea was detected from phytochemical analysis. These can also be applied for dental caries and are of great significance in therapeutic treatments. This study aimed to compare the activity of banana peel and black tea against periodontitis causing *S.aureus*.

Keywords – Black tea and banana peel extract, Fluoride, Phytochemical, Antibacterial, Agar well diffusion.

INTRODUCTION

Various plants and their products have been in use for their medicinal value since ancient times (Meckes *et al.*, 1995).Natural products have been an integral part of the ancient traditional medicine system (Sarkar *et al.*, 2007). Use of herbal remedies for prevention and cure of ailments is of increasing interest due to the superiority and efficiency of activity provided by phytoconstituents in herbs and undesirable effects of modern medicine. Plants are still major sources of the foods worldwide and phytochemicals can play a significant role in modern food and pharmaceutical industries (Asif Ahmed Kibria *et al.*, 2019).

Periodontitis, the multifactorial diseases are mainly derived by interaction between invasions of causative bacteria and host immune response of varied degrees (Jenabian *et al.*, 2012).Usually gram negative bacteria play a central role and gram positive bacteria are rare. Among, *Staphylococcus aureus* plays a rare role in causing Periodontitis as compared to gram negative organism.

The unripe peel of banana and of the leaves are taken in dysentery and diarrhea and used for treating malignant ulcers (Girish *et al.*,2008). The waste material of banana peel has medicinal properties and various bioactive compounds flavonoids, tannins, alkaloids, glycosides and terpenoids are present in banana peel which exerts a pharmacological effect, especially antioxidant, antibiotic, anti-inflammatory (Imam *et al.*,2011 and Chabuck *et al.*,2013).

Black tea is more oxidised than all other types of teas. It contains antioxidants and other substances that might help protect the heart and blood vessels (Diane *et al.*,2007).Tea is one of most popular and liked beverages worldwide. Tea is known to possess various biological properties (Daljit Singh Arora *et al.*,2007).The biological properties of tea and coffee are accredited mainly to their polyphenolic contents(flavonoids, catechins, tannins).The ability of these components to scavenge reactive oxygen species impart them potent antioxidant activities *in vitro* and *in vivo*. In the oral tract, *S. aureus* has been associated with dentoalveolar infections, and oral mucosal lesions (Ga-Yeon Kim *et al.*,2015).

MATERIALS AND METHODS

BACTERIAL CULTURE

The bacterial culture *Staphylococcus aureus* were obtained from Microbiological Laboratory of Kovai Medical Center and Hospital, Coimbatore. Those bacterial cultures were maintained by regular sub culturing on nutrient agar slants and stored at 4°C.

SAMPLE COLLECTION AND PREPARATION

The banana peels used were collected from banana (*Musa paradisiacal*) brought from market. They were dried in hot air oven at 120°C for 3h 40min and ground into powder with mechanical blender and sieved. The obtained powdered samples were soaked in 70% methanol, ethanol, hexane and aqueous are placed on the shaker for 72hrs at room temperature. The sample and extracts used for the extraction is 1:3(ratio) respectively.

The black tea is collected from market and blended into fine powder. Then soaked in 70% methanol, ethanol, hexane and aqueous and placed on the shaker for 72hrs or three days at room temperature. The sample and solvent was used for the extraction is 1:3 ratio respectively.

PHYTOCHEMICAL ANALYSIS OF EXTRACTS

Qualitative phytochemical analysis of the crude extracts of the banana peel and black tea collected were determined as follows

- 1) Alkaloids:** To 2ml of extract add few ml of dil. Hcl and filtered and add few drops of Mayer's reagent (saturated aqueous solution of picric acid). Yellow precipitate confirms presence of alkaloids.
- 2) Flavonoids:** To the few ml of extracts, few drops of NaOH and few ml of dil.Hcl was added. Yellow color is obtained when NaOH added become colorless after adding dil.Hcl which confirms the presence of flavonoids.
- 3) Saponins:** To 2 ml of dis H₂O add 5ml of extract. If foam persists for 10 minutes it indicates the presence of saponins.
- 4) Tannin:** Few drop of FeCl₃ (10%) solution was added with 2-3ml of extract and greenish grey or dark blue color indicates positive.

5) Phenols: 2ml of extract mixed to 5% FeCl₃ solution and deep blue color indicates presence of phenols.

ANTIBACTERIAL ACTIVITY

The banana peel and black tea extracts were used to test the inhibitory activity against the Periodontitis causing *Staphylococcus aureus*. The formation of clear zone was used to determine the efficacy of banana peel and black tea extracts. Bacterial inoculums were grown in nutrient broth for overnight or 24hrs.

SCREENING OF ANTIBACTERIAL ACTIVITY USING WELL DIFFUSION METHOD

The antimicrobial activity of *Musa paradisiaca* (Banana) peel and black tea extracted with different solvent (methanol, ethanol, hexane, aqueous) were tested against Periodontitis causing *Staphylococcus aureus* was measured using the well diffusion method. Muller Hinton Agar plates were prepared. Log phase test specimen *Staphylococcus aureus* were swabbed on the Muller Hinton Agar plate surface using the sterile cotton swab. Wells were made with the sterile gel puncture (cork borer) on Muller Hinton Agar plate and the 550mg/ml of the *Musa paradisiaca* and 50mg/ml of black tea solvent extracts were loaded on to the wells with MIC. The plates were incubated at 37°C for 24hrs and zone of inhibition around the well was measured in millimeters (mm).

MINIMUM INHIBITORY CONCENTRATION

Minimum inhibitory concentration (MIC) is defined as the lowest concentration of antimicrobial that will inhibit the visible growth of microorganism after overnight incubation. Bacterial inoculums were grown in nutrient broth for overnight. The extract of *Musa paradisiaca* (banana) peel and black tea of different solvent were subjected into the determination of MIC using the tube dilution technique. The ethanolic extract of banana peel and methanolic extract of black tea were taken. A double fold serial dilution was made using Muller Hinton Broth (MHB). The concentration obtained for Black tea was 50mg/ml, 25mg/ml, 10mg/ml, 7mg/ml, 3.5mg/ml, 3mg/ml, 2.5mg/ml, 2mg/ml, 1.6mg/ml and the concentration obtained for banana peel was 550mg/ml, 520mg/ml, 500mg/ml, 480mg/ml, 250mg/ml, 120mg/ml, 60mg/ml, 32mg/ml and 8mg/ml. Equal volume of extract and Muller Hinton broth (2ml) was dispensed into sterilized test tubes. A quantity (0.1ml) of standardized inoculums (0.9×10^8 cfu/ml) was added to each of the test tubes which were incubated aerobically at 37°C for each 24hrs. The lowest concentration of the extracts which inhibited microbial growth (no turbidity) was recorded as the (MIC) minimum inhibitory concentration.

RESULT AND DISCUSSION

PHYTOCHEMICAL ANALYSIS

The phytochemical active compounds were qualitatively analyzed from the four types of extracts such as methanolic, ethanolic, hexane and the aqueous of both *Musa paradisiaca* (banana) peel and black tea.

TABLE:1 Extracts of *Musa paradisiaca* and Black tea

Phytochemical	Methanol Extract		Ethanol Extract		Hexane Extract		Aqueous Extract	
	Banana peel	Black tea	Banana peel	Black tea	Banana peel	Black tea	Banana peel	Black tea
Flavonoids	–	+	+	+	+	+	–	–
Saponins	+	–	+	–	–	+	–	+
Tannins	+	+	+	+	–	–	–	+
Phenol	+	–	–	+	–	–	+	+
Alkaloids	+	+	+	+	+	+	+	+

ANTIBACTERIAL ASSAY

The antibacterial assay for the eluted fractions was measured using well diffusion method. The zone of inhibition was measured and tabulated.

TABLE: 2

Solvents	<i>Musa paradisiaca</i> (banana peel)	Black tea
Methanolic extract	–	5.66±1.24
Ethanolic extract	3.66±1.24	3.66±0.471
Hexane extract	–	–
Aqueous extract	–	–
DMSO (Negative control)	–	–
Streptomycin (Positive control)	7±0.816	6.33±1.247

The ethanolic extract of banana peel (*Musa paradisiaca*) and methanolic extract of black tea showed effective inhibition against *Staphylococcus aureus*.



PLATE:1 Antibacterial activity of black tea against *S.aureus*

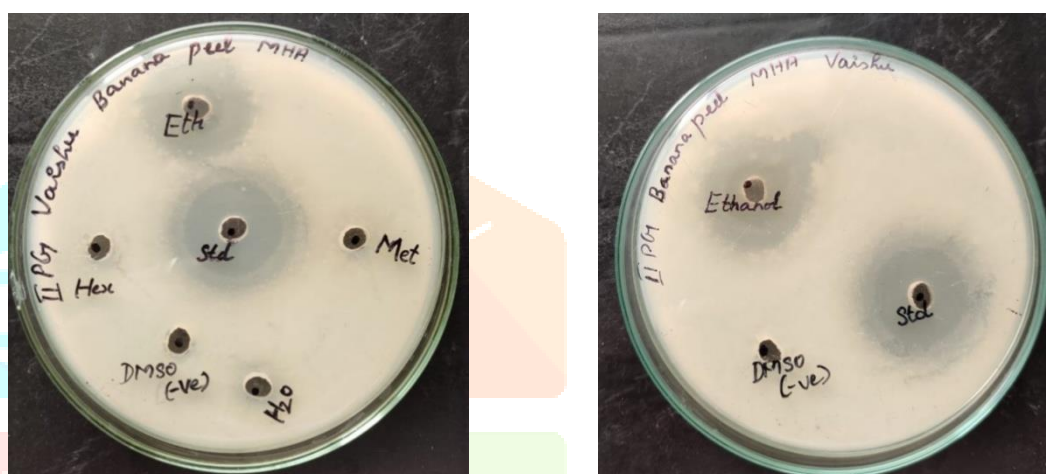


PLATE:2 Antibacterial activity of banana peel(*Musa paradisiaca*) against *S.aureus*

MINIMUM INHIBITORY CONCENTRATION

The minimum inhibitory concentration is the lowest concentration of extract to kill or inhibit the growth of microorganisms. The MIC was performed for *Staphylococcus aureus* on ethanolic extract of *Musa paradisiaca* (banana) peel and methanolic extract of black tea. The turbidity is checked to identify the MIC. The minimal inhibitory concentration of Ethanolic extract of banana peel is 500mg/ml and methanolic extract of black tea is 7mg/ml.

TABLE: 3

Extracts	MIC
Banana peel(Ethanolic extract)	500mg/ml
Black tea(Methanolic extract)	7mg/ml



PLATE:3 MIC of banana peel extract

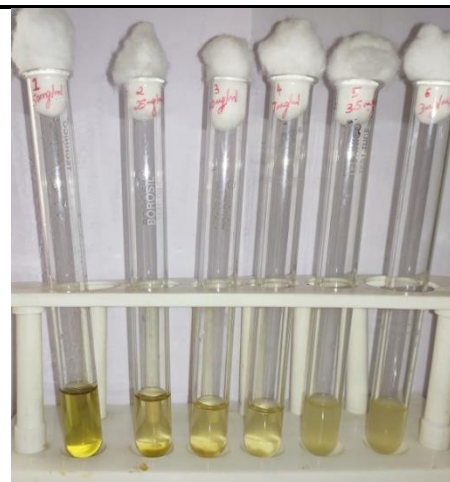


PLATE:4 MIC of black tea extract

The preliminary phytochemical screening showed *Musa paradisiaca*(banana)peel and black tea contain some secondary metabolites such as alkaloids, phenols, flavonoids, Saponins and tannins. Secondary metabolites present in plants have been reported by Rabe (2000) to be responsible for the antimicrobial activity associated with some ethnomedicinal plants.

Black tea and banana (*Musa paradisiaca*) is to contribute natural remedies and useful in curing bacterial infections. Where, the black tea contributes fluoride equal to the adequate amount as 0.05mg F/kg body weight/day. Where fluoride in tea could be beneficial for the prevention of dental caries but excessive intake is not by children at the age of risk to dental caries should be avoided. The black tea is only effective when it is not added with milk, sugar and it has active compounds only when it is taken alone without any additives.

The antibacterial efficacy of both banana peel and black tea extracts were detected with measuring the zone of inhibition. The positive control used is Streptomycin and DMSO as negative control.

CONCLUSIONS

The study showed the difference that the ethanolic extract of banana peel (*Musa paradisiaca*) was effective than its methanolic extract where the methanolic extract of black tea was effective than its ethanolic extract. The ethanolic extract of banana peel shown highest MIC of 7mg/ml and the MIC of black tea methanolic extract against *S.aureus* was 500mg/ml. In this present study, banana peel (*Musa paradisiaca*) and black tea extract to screen the efficacy on periodontitis causing *S.aureus* as a comparative study and revealed both samples are effective on *S.aureus* but of different solvents.

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CONFLICTS OF INTEREST

The authors have no conflicts of interest to publish this research article in this journal.

REFERENCE

- 1) Bushman, J. (1998). Green tea and cancer in humans: a review of the literature. *Nutrition and Cancer*, 31, 151-159.
- 2) Yang, C.; Landua, J. 2000 Effects of tea consumption on nutrition and health. *Journal of Nutrition*, 130, 2409-2412.
- 3) McKay, D.L.; Blumberg, J.B. 2002. The role of tea in human health: an update. *Journal of American college of Nutrition*, 21, 1-13.
- 4) Sesso, H.; Gaziano, J.; Buring, J.; Hennekens, C. 1999. Coffee and tea intake and risk of myocardial infarction. *American Journal of Epidemiology*, 149, 162-167.
- 5) Tariq, A.L., and Reyaz, A.L. (2012). *International Journal of Drug Development and Research*, vol. 4, no 4, 311-316.
- 6) Sarkar, S.D., and Nahar, L. 2007. *Chemistry for pharmacy students General, Organic and Natural Product Chemistry*. England: John Wiley and sons. Pp. 283-359.
- 7) Ehiowemwenguan, G., Emoghene, A. O. and Inetianbor, J. E. 2014. 8 August, Volume 4, *IOSR Journal of Pharmacy*.
- 8) Asif Ahmed Kibria, kamrunnessa, Md. Mahmudur Rahman, Annanya kar. 2019 volume 2, *Malaysian Journal of Halal Research Journal*.
- 9) Sumathy, V., 2011. In vitro bioactivity and phytochemical screening of *Musa acuminata* flower, *Pharmacology online* 2, 118-127.
- 10) J. A. Shagana, Dr. R. Geetha. 2017 *Journal of Pharmaceutical Sciences and research*, vol 9(9).
- 11) Pinky Goswami, Chandanakalita, Atoolchandue Bhuyan. J. 2020. *Evolution Med. Dent. sci*; 9 (01); 18-22.
- 12) Ehiowemwenguan G., Emoghene A., Inetianbor J. 2014 Antibacterial and phytochemical analysis of Banana fruit peel. *IOSR J. Pharm.* 4 (8), 18.
- 13) Chabuck Z. A. G., Al-Charrakh A. H., Hindi N. K. K., Hindi S. K. K. 2013. Antimicrobial effect of aqueous banana peel extract, *Iraq. Res. Gate: Pharm. Sci.* 1, 73.
- 14) Parashar S., Sharma H., Garg M. 2014. Antimicrobial and antioxidant activities of fruits and vegetable peels: A review. *J. Pharmacogn. Phytochem.* 3 (1).
- 15) Abo, K. A., Ogunleye, V. O., and Ashidi, J. S. 1991. Antimicrobial potential of *Spondias mombin*, *croton zambesicus* and *Zygotritonia crocea*. *Journal of Pharmacological Research*. 5(13): 494-497.
- 16) Liu, R. H., 2004. Potential synergy of phytochemicals in cancer prevention: mechanism of Action. *Journal of Nutrition*. 134(12): 3479-3485.
- 17) Nweze, E. L., Okafor, J. I. and Njoku O. 2004. Antimicrobial Activities of Methanolic extracts of *Traumequinesis* (scchumn and Thorn) and *Morinda Lucinda* used in Nigerian Herbal Medical practice, *Journal of Biological Research and Biotechnology*. 2(1): 34-46.
- 18) Doughari, J. H., Human, J. S., Bennade, S. and Ndakidemi, P. A. 2009. Phytochemicals as chemotherapeutic agents and antioxidants: possible solution to the control of antibiotic resistant verocytotoxin producing bacteria. *Journal of Medicinal Plants Research*. 3(11): 839-848.
- 19) Tiwari, B. K., Brunton, N. P., Brennan, C. 2013. *Handbook of plant food phytochemicals: sources, stability and extraction*. John Wiley and Sons, 1-4.

- 20) Tiwari, B.K., Valdramidis, V.P. 2009. "Application of Nutritional Antimicrobials for Food Preservation". *Journal of Agricultural and Food Chemistry* 57 (14), 5987-6000.
- 20) Karakaya, S. Kavas, A. 1999. Antimutagenic Activities in some foods, *Journal of Science of Food and Agriculture*, 79(2),237-242.
- 21) Belitz, H.D., Grosch, W. 2004. *Aroma compounds, Food chemistry*. Springer, Berlin, Heidelberg, 342-408.
- 22) Akhter, A.Zaman, S.Umar, M. Ali, M.Y., Ali, Jalil Miah, M.A. 2010. Isolation of Polyphenolic Compounds from the Green Coconut (*cocos nucifera*) Shell and Characterization of their Benzoyl Ester Derivatives. *Journal of Science and Research*. 2 (1), 186-190
- 23) Schieber, A., Stintzing, F.C. 2001. "By products of plant food processing as a source of functional compounds-recent developments" *Trends in food science and technology* 12 (11),401-413.
- 24) Cowan, M.M., 1999. Plant Products as Antimicrobial Agents. *Clinical Microbiology Review*, 12 (4) 564-582.
- 25) Van V, Suliman, S.F.S., 2009. "The antimicrobial activity of four commercial essential oils in combination with conventional antimicrobials." *Letters in Applied Microbiology*, 48 (4), 440-446
- 26) Monika. S, 2014. Antioxidant properties of extracts from selected plant materials (*Caesalpinia spinosa*, *Perilla frutescens*, *Artemisia annua* and *Viola wittrockiana*) in vitro and in model food systems, *Universitat Politecnica De Catalunya, Barcelonatech*. Pp.97-102
- 27) Novak, F. J. 1992. *Musa (Bananas and Plantains)*. In: Hammerschlag, F.A. and Litz., R.E.(eds), *Biotechnology of Perennial Fruit Crops*. CAB International, University Press, Cambridge. U.K. pp. 449-488.
- 28) Husain, M.D., and William, R. 2010. Status of banana cultivation and disease incidences in Malaysia. *Crop Protection and Plant Quarantine Division, Department of Agriculture, Malaysia*, Pp.: 60.
- 29) FAO (Food and Agricultural Organization). 2004. *Production Yearbook 2003*, Rome, Italy
- 30) Hossain, M.F., 2014, A Study of Banana Production in Bangladesh: Area, yield and major constraints. *ARPN Journal of Agricultural and Biological Science*. . 9(6), 205-210.
- 31) Haque M.A., 1988. *Kolar Bagan (In Bengali)*. 3rd Edition. Banana Research Project, Bangladesh Agricultural University, Mymensingh. p. 24.
- 32) Mukul, A.Z.A., Rahman, A. 2013. Production and profitability of banana in Bangladesh-an economic analysis. *International Journal of Economics, Finance and Management Sciences*. 1(3): 159-165.
- 33) Chung, S.L., and Meyers, S.P., 1979. Bioprotein from banana waste. *Development in Industrial Microbiology*, 20, 723-731.
- 34) Krishna, C. 1999. Production of bacterial cellulose by solid state bioprocessing of banana waste. *Bio resource Technology*, 69, 231-239.