

# EVALUATION OF ASCORBIC ACID CONTENT *IN VIVO* AND *IN VITRO* IN *BALANITES AEGYPTIACA* DEL.

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## ABSTRACT

Evaluation of Ascorbic acid content of roots, leaves, fruits and tissue cultures from the selected medicinal plant *Balanites aegyptiaca* growing in arid zone of Rajasthan was carried out. Among all the samples the maximum (92.3 mg/100 g.d.w.) amount of ascorbic acid was found in 6 weeks old tissue cultures, while the leaves of *Balanites aegyptiaca* had minimum concentration (58.2 mg/100 g.d.w.) collected from field area of Bikaner District.

**KEY WORDS:** Ascorbic acid contents, *Balanites aegyptiaca*, *in vivo* and *in vitro*

## INTRODUCTION

The medicinal plant species growing in arid zone of Rajasthan are good and potential source of nutritionally and phytochemically important compounds so these can be used as herbal medicine and livestock feed. Ascorbic acid, also called as anti-scorbutic (Vitamin C), is an important primary product and well known for its property as an electron doner in photophosphorylation.

*Balanites aegyptiaca* Del. (Zygophyllaceae), known as 'desert date,' and locally as Hingota. It is a spiny shrub or tree up to 10 m tall, widely distributed in arid region of Rajasthan. It is traditionally used in treatment of various ailments i.e. jaundice, intestinal worm infection, wounds, malaria, syphilis, epilepsy, dysentery, constipation, diarrhoea, hemorrhoid, stomach aches, asthma, and fever. It is also used as fodder for livestock.

The role of ascorbic acid in plant growth and metabolism has been worked out by various workers [1-5]. Free endogenous ascorbic acid has been recently reported from some arid zone plant species [6-16].

## MATERIALS AND METHODS

Plant parts were collected in polythene bags. The samples were dried, powdered and then used for the estimation of free endogenous ascorbic acid.

The surface sterilized seeds were aseptically placed on hormone free MS medium for germination in the dark at  $28\pm 2^{\circ}\text{C}$ . Cotyledons and radicals from these aseptically grown 15 – 20 days old seedlings were taken as explants.

These explants were then established and maintained by frequent subculturing after 4 weeks on MS Medium supplemented with various concentrations and combinations of kinetin and 2, 4-D for callus induction and kinetin and BAP for induction of multiple shoots. Cultures were maintained in growth chamber with regulated temperature ( $26\pm 2^{\circ}\text{C}$ ), relative humidity ( $55\pm 5\%$ ), 3000 lux light intensity. Data was recorded after 2, 4, 6, 8 and 10 weeks and growth indices were calculated.

Fresh and healthy roots, leaves and fruits of the selected plant collected from Bikaner district and tissue cultures were dried and homogenized in a mortar with 2% metaphosphoric acid (MPA)(10 mg powder: 100 ml MPA) and allow to macerate for one hour. The mixtures were centrifuged at low speed (2500 rpm) and supernatants were used for estimation of ascorbic acid following the colorimetric method [5]. Absorbancy of each of the

sample was measured on a spectronic-20 colorimeter (Bausch & Lomb) set at 546nm against blank. Values are expressed in mg / 100 g.d.w

## RESULTS AND DISCUSSION

Concentration of the ascorbic acid in the various parts (roots, leaves and fruits) and 2, 4, 6, 8, 10 weeks old tissue cultures of the selected plant species *Balanites aegyptiaca* are presented in Table- 1.

**Table 1:** Ascorbic acid contents (mg/100 g.d.w) from plant parts and tissue cultures

Plant	Roots	Leaves	Fruits	Tissue Cultures 2 weeks	Tissue Cultures 4 weeks	Tissue Cultures 6weeks	Tissue Culture s 8 weeks	Tissue Cultures 10 weeks
<i>Balanites aegyptiaca</i>	74.3	58.2	80.8	43.3	90.3	92.3	82.3	81.2

The present investigation shows that among all the samples tested the ascorbic acid content was found to be maximum (80.8 mg/100 g.d.w.) in the fruits, while minimum (58.2 mg/100 g.d.w.) in the leaves of *Balanites aegyptiaca*.

In tissue cultures of *Balanites aegyptiaca* maximum (92.3 mg/100 g.d.w.) ascorbic acid content was found in 6 week old tissue cultures, while minimum (81.2 mg/100 g.d.w.) in the 10 weeks old tissue cultures.

Among all the various plant parts and tissue cultures the maximum (92.3 mg/100 g.d.w.) amount of ascorbic acid was found in 6 weeks old tissue cultures, while the leaves of *Balanites aegyptiaca* had minimum concentration (58.2 mg/100 g.d.w.).

## CONCLUSION

The present study thus indicates that medicinal plants of this arid region of Rajasthan are good source of ascorbic acid (Vitamin C) so these can be used as livestock feed and as herbal medicine for human welfare.

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## REFERENCES

1. Arnon D I, Whatley FR and Allen M B, (1954), Photosynthesis by isolated chloroplast II, Photosynthetic Phosphorylation and the conversion of light into phosphate bound energy .*J. Amer.Chem. Soc.* 76: 6324-6329.
2. Aberg B, (1958), Ascorbic acid formation, storage, mobilisation and transformation of carbohydrates. In: *Encyclopedia of Plant Physiology*, Springer Verlag. Berlin. 6:479-499.
3. Mitsui A and Oi Y, (1961), Endogenous changes of photochemical activities of Spinach leaves. *Plant Cell Physiol.* Tokyo. 2: 45-50.
4. Isherwood FA and Mapson LW, (1962), Ascorbic acid metabolism in plants: Part II. Biosynthesis. *Ann. Rev. Plant Physiol.* 13:329-350.
5. Jenson WA, (1962), *Botanical Histochemistry – Principles and Practice*. W.H. Freeman and Company. San Fransisco. pp201.
6. Kapoor BBS, (1989), Free endogenous ascorbic acid from *Argemone mexicana* growing in Arid Zone of Rajasthan. *Oikoassay.* 6 (2): 83.
7. Harsh ML and Ahmed S, (1994), *Maytenus emarginata*, *Parkinsonia aculeate* and *Tecomella undulata* : New Sources of ascorbic acid . *Oikoassay.* 11: 5.
8. Kapoor BBS and Ritu, (1996), Comparative evaluation of ascorbic acid from some trees growing in arid zone of Rajasthan. *Oikoassay.* 13 (1&2): 29.
9. Tyagi S and Nag TN, (2002), Endogenous level of ascorbic acid in moth bean cultivar in vivo and in tissue culture. *J. Indian Bot. Sci.* 8: 94-98.
10. Kapoor BBS and Priydershan Ranga, (2003), Ascorbic acid contents from asteraceous medicinal plants of Rajasthan desert. *Indian J. of environmental sciences.* 7 (2): 173 – 174.
11. Kapoor BBS, Khatri JS, Bhumika and Priydershan Ranga, (2004), Herbal plants of Hanumangarh district: New sources of ascorbic acid. *J. Phytol.Res.* 17(1): 111-112.
12. Kapoor BBS, JS Khatri, Bhumika and Priydershan Ranga, (2005), Evaluation of ascorbic acid contents in some arid zone tree species. *Indian J. of environmental sciences.* 9(1): 31-32.
13. Kapoor BBS and Mishra Raksha, (2013), Capparidaceous Medicinal Plants of North-west Rajasthan: Good Sources of Ascorbic Acid. *Indian Journal of Pharmaceutical and Biological Research.* 1(2): 20 – 22.
14. Kapoor BBS and Swati Lakher,a (2013), Ascorbic Acid Contents from Some Medicinal Plant Species of the Jodhpur District of Rajasthan: *International Journal of Universal Pharmacy and Biosciences.* Vol. 2 No.4, pp 128-132.
15. Kapoor BBS and Kumar Sunil, (2014), Ascorbic acid contents from some medicinal plants of Barmer district of Rajasthan. *International Journal of Universal Pharmacy and Bio Sciences* 3(5): 80-84.
16. Kapoor BBS and Manju Saharan, (2016), Comparative Evaluation of Ascorbic acid Contents from Some Plants of Barmer District of Rajasthan. *CBITECH Journal of Basic and Applied Chemical Sciences.* 5 (2): 10 – 12.